Multipoint fluorescence correlation spectroscopy with total internal reflection fluorescence microscope

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Abstract. We report simultaneous determination of diffusion coefficients at different points of a cell membrane using a multipoint fluorescence correlation spectroscopy (FCS) system. A system carrying seven detection areas in the evanescent field is achieved by using seven optical fibers on the image plane in the detection port of an objective-type total internal reflection flow correlation spectroscopy (TIR-FCS) system. Fluctuation of fluorescence intensity is monitored and evaluated using seven photomultiplier tubes (PMTs) and a newly constructed multichannel correlator. We demonstrate simultaneous-multipoint FCS, with a 3-µs time resolution, to investigate heterogeneous structures such as cell membranes and membrane-binding molecular dynamics near glass surfaces in live cells. © 2009 Society of Photo-Optical Instrumentation Engineers. [DOI: 10.1117/1.3080723]

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1 Introduction

Fluorescence correlation spectroscopy1 (FCS) is a powerful tool to investigate molecular behavior such as diffusional mobility, photophysical properties, and reaction kinetics in vitro and in vivo. In spite of their wide range of application, FCS measurements are restricted to monitoring at only one volume element defined by both a focused laser beam and a pinhole. Although there have been a few reports on FCS experiments with multiple detection areas, further advances in the number of detection areas, sensitivity, time resolution, and methods are required.

Heterogeneous structures of cell surfaces and the motion of membrane-binding molecules are thought to be dynamic and presumed to modulate the formation of cell-signaling molecular complexes at the membranes. Hence, simultaneous FCS measurement with multiple detection areas would be an attractive tool for understanding of the heterogeneous diffusion properties of cell membranes and such cell signaling.

Recently, some research groups reported total internal reflection fluorescence correlation spectroscopy2–10 (TIR-FCS) and its application to living cells.11 Although the limited-time temporal resolution of 4 ms, electron-multiplying CCD (EMCCD)-camera-based multipoint TIR-FCS measurement was achieved.12 Extension of the multipoint FCS system will be spatially resolved FCS such as image correlation spectroscopy (ICS), however the temporal resolution should be improved13 (see more detailed in the review article in Ref. 14). In previous work, we also reported the multipoint FCS measurement made by confocal laser-scanning microscopy. The detection areas were focused in the cytoplasm that located deeper from the plasma membrane though the measurements were not completely simultaneously.15

In this paper, we describe the high temporal resolution of multipoint TIR-FCS (M-TIR-FCS). TIR illumination produces an evanescent field. The intensity decreases exponentially along the optical axis but is widely spreading on the surface. Therefore, as shown in Fig. 1(a), multipoint FCS detection areas in the evanescent field were achieved by putting multiple optical fibers on the image plane.

The optics of M-TIR-FCS is based on an objective-type TIR-FCS, which is combined with objective-type total internal reflection fluorescence microscopy (TIRFM, TE2000, Nikon) with a ×100 oil-immersion objective [Plan Apo, numerical aperture (NA) 1.49, Nikon] for TIR illumination and a 488-nm semiconductor laser (488-20CDRH, Coherent, California) for the excitation. On the other hand, a bundle of seven multimode fibers with a 50-µm-diam core (Seiko-Giken, Chiba, Japan) to produce seven detection areas in the evanescent field was placed on an image plane in its detection port. Seven photomultiplier tubes (PMTs, H7421, Hamamatsu Photonics K. K., Shizuoka, Japan) to detect fluorescence signals in parallel through each optical fiber and a multichannel correlator (Hamamatsu Photonics K. K.) to simultaneously calculate seven autocorrelation functions were set up as shown in Fig. 1(b). The sensitivity of the PMT is a little lower than that of an avalanche photodiode (APD), but the price of PMT is lower than that of the APD and the PMT has larger detection area so that it is easy to align; these properties are benefits for a multiple-detection system.
For the calibration control experiments on 100-nM fluorescein (FITC) in 10-mM Tris buffer (pH 8.0) were carried out according to previous study. By monitoring seven detection areas at the same time, seven individual fluorescence autocorrelation functions (FAFs) \( G(\tau) \) were acquired online with a digital multichannel correlator (Hamamatsu Photonics K. K., Shizuoka, Japan). As shown in Fig. 2, seven autocorrelation functions (dotted lines) were simultaneously obtained. To calibrate the size distribution among the seven detection volumes, fitting by a one-component model \(^1\) (solid lines), [Eq. (1)], was conducted to obtain the height \( h \) and the radius \( \omega_{xy} \) of each detection volume. The structure parameter is defined \(^6\) by \( \omega = h/\omega_{xy} \).

The analysis yielded diffusion time along the \( z \) axis, \( \tau_z = 3.5 \mu s \), which shows that M-TIR-FCS has a 3-\( \mu s \) time resolution at least. However, time resolution is not clear in this study because the time resolution should take into account the resolution of the detector, amplifier, and correlator system. In a small lag time range, a high peak signal were observed and fitted by the triplet term by using Eq. (1). Though the after pulse could be effected to the signal at small lag time, the effect will be small composed to the diffusion signal part. The difference in the values of \( h \) and \( \omega_{xy} \) might be due to the deviation of the core sizes of optical fibers, the \( z \) axis position at the image plane, and alignment of the fibers (Fig. 2). The height and radius of the detection volumes were about 60 and 330 nm, respectively. These parameters were influenced by TIR angle as follows. According to the increase in the TIR angle from critical angle, \( \tau_z \) decreases to 4.3 ± 0.9, 3.8 ± 0.8, 3.1 ± 0.5, and 2.6 ± 0.3 \( \mu s \), and \( h \) decreases to 72 ± 7.6, 67 ± 7.9, 60 ± 4.5, and 56 ± 3.5 nm, respectively. On the other hand \( \omega_{xy} \) is constant to 387 ± 33.4, 386 ± 28.7, 387 ± 30.5, and 385 ± 31.6 nm. Finally, \( N \) decreases 19.6 ± 0.9, 19.8 ± 1.0, 19.1 ± 1.1, and 18.0 ± 2.0. The distances between each core of the optical fibers on the image plane, and alignment of the fibers (Fig. 2).
plane was about 250 μm, as shown in Fig. 1(b). In this system, the distances between each M-TIR-FCS detection spots were about 2.5 μm because we used a ×100 objective. The distance of 2.5 μm was distant enough for the observation area of 660 nm not to detect crosstalk signals from neighboring detection spots. Also, the cross-correlation between signal from each spot was not detected (data not shown). From these results, we confirmed that M-TIR-FCS worked successfully.

Next, we carried out measurement of membrane-binding fluorescent proteins expressed in a living COS7 cell using M-TIR-FCS. Expression vectors encoding farnesylated monomeric-type EGFP A206K variants, which are abbrevi-
ated mGFP-F, were constructed by mutating sequences of pEGFP-F (Clontech) with site-directed mutagenesis (Quick-Change, Stratagene). For M-TIR-FCS measurement, COS7 cells were grown on glass-based dishes (φ 12 mm, Asahi Technoglass, Chiba, Japan) 2 days before measurements. The cells were transiently transfected with 0.5 μg of plasmid DNA of mGFP-F and 4.0 μg of opitект (Invitrogen) 17 h before M-TIR-FCS measurements. The left photograph in Fig. 3(a) shows TIRFM image of a COS7 cell expressing farnesylated monomeric type (A206K) enhanced GFP, mGFP-F. The broken white line shows the border of the plasma membrane of a COS7 cell attached to the glass surface. The width of the cell is about 15 μm. The right photograph in Fig. 3(a) shows the end face of the bundle of optical fibers used as seven pinholes. The value 4.4 μm is the length of the projected image in the object port. Figure 3(b) shows the seven autocorrelation functions (dotted lines) obtained by M-TIR-FCS of mGFP-F in the cellular membrane of a COS7 cell and its fitting results with a lateral diffusion parallel to the surface and three dimensional diffusion through evanescent profile

\[ G(\tau) = 1 + \frac{\gamma}{N} \left[ F_1 \left( 1 + \frac{\omega^2 \tau}{\tau_{1,2}} \right)^{-1} \left( 1 - \frac{\tau}{2 \tau_{1,2}} \right) \right] + \frac{\tau}{\pi \tau_{1,2}} + F_2 \left( 1 + \frac{\tau}{\tau_{2,xy}} \right)^{-1}, \]

where \( F_1 \) and \( F_2 \) are each fraction, respectively.

The time range of the correlation function is much slower than that obtained by measurements of fluorescein in aqueous solution. These results show that TIR-FCS covers a wide range of diffusion time induced by large molecular complex in cell membrane. Diffusion constants of mGFP-F in the cell membrane were calculated in the same way as described in a previous report. The 3-D diffusion can be related to free-moving protein in cytosol because the obtained diffusion constant is comparable with previous report. The difference in values of \( D_2 \) may reflect the heterogeneous structure of cellular membranes, and its average value is almost the same as the result in a previous work. At the lag time, the measurement curves seem to decay much faster than the model. This may be the result of the instability of the measurement conditions; this should be answered in near future.

We reported simultaneous diffusion measurement at multiple tiny areas close to surfaces in not only aqueous solution but also in live cells using multipoint FCS with a wide time resolution from the microsecond to the millisecond range. Because the mGFP-F membrane-binding protein we measured here is a model protein, the variation of its diffusion rate on membranes would still be small. Thus, M-TIR-FCS can be a powerful technique for the study of more complex and heterogeneous specimen, i.e., membrane-receptor complex in live cells. Furthermore, the spatial cross-correlation method would be applicable to this system and make M-TIR-FCS versatile and more oriented toward FCS imaging. A much more interesting application would be to exploit simultaneously the spatial and temporal correlation between different measurement positions. This could enable us to measure drift, diffusional anisotropy, direction flow, and other effects. However, these applications should be performed in the near future.

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References