Third-harmonic generation for the study of Caenorhabditis elegans embryogenesis

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Abstract. Live microscopy techniques (i.e., differential interference contrast, confocal microscopy, etc.) have enabled the understanding of the mechanisms involved in cells and tissue formation. In long-term studies, special care must be taken in order to avoid sample damage, restricting the applicability of the different microscopy techniques. We demonstrate the potential of using third-harmonic generation (THG) microscopy for morphogenesis/embryogenesis studies in living Caenorhabditis elegans (C. elegans). Moreover, we show that the THG signal is obtained in all the embryo development stages, showing different tissue/structure information. For this research, we employ a 1550-nm femtosecond fiber laser and demonstrate that the expected water absorption at this wavelength does not severely compromise sample viability. Additionally, this has the important advantage that the THG signal is emitted at visible wavelengths (516 nm). Therefore, standard collection optics and detectors operating near maximum efficiency enable an optimal signal reconstruction. All this, to the best of our knowledge, demonstrates for the first time the noninvasiveness and strong potential of this particular wavelength to be used for high-resolution four-dimensional imaging of embryogenesis using unstained C. elegans in vivo samples. © 2010 Society of Photo-Optical Instrumentation Engineers. [DOI: 10.1117/1.3477535]

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1 Introduction

Live microscopic recordings have enabled researchers to understand the developing mechanisms involved in cells and tissue formation. One particular case is the study of embryogenesis using the soil nematode Caenorhabditis elegans (C. elegans). This has been addressed employing different microscopy techniques. To perform these recordings, for example, a combination of differential interference contrast (DIC)/Nomarski and fluorescence microscopy has been used to track morphogenesis processes in vivo. DIC microscopy uses polarized light allowing it to capture intrinsic features of thin transparent specimens (preferentially). This produces a contrast that is generated from the encoded phases inside the sample, but it lacks being specific to structures, and it possesses limited axial resolution. If the development of a specific structure needs to be recorded individually, other techniques, such as fluorescence microscopy, have to be...
used. In such a case, the use of fluorescent markers or vital dyes endogenously expressed is required for a selective observation. This has, however, time, technical, and sample viability limitations besides the inability to produce high-resolution three-dimensional (3-D) images.

Given this, alternative research works have proposed the use of confocal laser scanning microscopy to compare wild-type organisms, studying a large number of existent mutant alleles and transgenic reporter strains. All this has been used to perform lineage analysis and recordings of embryogenesis in *C. elegans*.

However, this technique requires the use of fluorescent reporters. Moreover, as the technique is based on linear excitation, care should be taken to minimize photodamage effects (i.e., photobleaching and phototoxicity). In addition, the excitation power must be dynamically adjusted to allow for long-term recordings in order to reduce exposure effects on the sample. As a consequence, the dim signal of the system can limit the cell identification task. Therefore, to track cells and morphological processes, the development of relatively complex algorithms or specialized cell tracking software is required (i.e., Endrov and Simi-Biocell).

A successful implementation using two-photon excited fluorescence (TPEF) to perform four-dimensional (4-D) developmental analyses in *C. elegans* was reported by in Ref. 13. Due to the reduced sample damage (based on the nonlinear effect using excitation wavelengths in the near infrared), it is possible to image living specimens for long periods. However, the use of fluorescent probes is still required.

Alternative methods, such as second-harmonic generation (SHG), and third-harmonic generation (THG), offer the advantage of being label-free techniques. These valuable tools have been used to study *in vivo* biological samples (i.e., *C. elegans*, *Drosophila melanogaster*, and Zebrafish). For instance, these studies aim to establish higher harmonic imaging modalities as label-free techniques showing a great potential to perform different developmental studies in a less invasive way. The generation of SHG signal requires a noncentrosymmetric structure, limiting its applicability in tissue mainly to collagen, muscle, and microtubules. In contrast, for the case of THG, the only imposition to generate a signal is the existence of interfaces (change in the refractive index or in the $\chi^3$ nonlinear coefficient). Additionally, a significant THG signal has been reported to arise from lipid bodies in different samples such as *Drosophila melanogaster*, fresh mouse lung tissue, *Arabidopsis thaliana*, etc.

In particular, in the past, the use of wavelengths within the range of the 1500 nm from a synchronously pumped optical parametric oscillator (OPO) has been reported. In these experiments, fixed samples such as neurons and epithelial cells have been imaged using the THG technique. Recently, the development of fiber-based femtosecond lasers has simplified the integration of nonlinear microscopy (NLM) techniques in biology, due to the compact design, output wavelength, pulse duration, and repetition rates of these new generation sources. These lasers use reliable state-of-the-art telecommunications components that have been developed for long-term operation, minimum maintenance, and moderate cost. Thus, a fiber laser operating at 1560 nm has been used, e.g., to produce THG images of chloroplasts and flowing erythrocytes.

In this work, we show the potential of the THG technique to be used for time-lapse studies in developmental processes. This is done by using a fiber-based mode-locked femtosecond telecommunications laser operating at 1550 nm. We show that employing this wavelength, it is possible to produce THG images of embryogenesis/morphogenesis process *in vivo* in *C. elegans* embryos in a noninvasive (label-free) way. The use of this specific wavelength has the advantage that the emitted THG signal is located at the visible range of the spectrum (516 nm). As a result, the requirement of UV-grade optics is eliminated, and therefore, standard photo multiplier tubes (PMTs) can be used, as they have a maximum detection efficiency located within the visible range. This wavelength has not been used in living samples or for long-term exposure experiments, and therefore, the sample viability is unknown. On one hand, UV light is not involved in the imaging process, increasing the sample viability. On the other hand, water absorption caused by the excitation wavelength starts to be important. For this reason, in this work, and in addition to the demonstration of THG microscopy at $\lambda=1550$ nm, we confirm that it is possible to perform 4-D (time-lapse 3-D) studies for several hours in living organisms.

The emitted THG signal was present in all the developmental stages of the embryo, enabling the identification of tissues/structures and showing its potential for embryogenesis/morphogenesis studies. As a result, to the best of our knowledge, this is the first time that THG imaging at this wavelength is used to obtain 4-D embryogenesis/morphogenesis information of *C. elegans* *in vivo* samples.

### 2 Materials and Methods

#### 2.1 Experimental Setup

The experimental setup was based on an inverted microscope (Nikon, Eclipse TE 2000U) modified to work as a laser scanning THG microscope.

A compact fiber-based pulsed laser system (Toptica Photonics, FemtoFiber FFS laser) operated at a central wavelength of 1550 nm, having a pulse duration of 100 fs and repetition rate of 107 MHz, was employed to excite the sample. The laser delivered an output average power up to 350 mW. An RG1000 filter (Schott; transmittance: 830 to 2000 nm) was placed at the laser output to block any spurious signal below 1000 nm from the laser itself. After passing through all the optical elements (see Fig. 1), the measured average power was 4.9 mW (measured at the sample plane), which corresponds to 460-W peak power.

The microscope is equipped with a pair of $x$-$y$ galvanometric mirrors (GMs) (Cambridge Technology, 6215H) used to scan the excitation beam over the sample at the maximum scanning speed of $\sim$300 lines/s. A filter cube, containing a silver-coated mirror to direct the excitation beam toward the sample, was used. A 40× oil immersion microscope objective with NA=1.3 (Nikon, Plan Fluor) was used during the experiments. The THG signal was collected by an oil immersion condenser, NA=1.4 (Nikon). A custom-made forward-detection mount using an attached photomultiplier tube (PMT; Hamamatsu, H9305-04) to detect the THG signal at the central wavelength of 516 nm was implemented. (No requirement on specialized collection optics are imposed.) This

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mount contained a bandpass filter to separate the fundamental beam from the generated THG signal (Semrock; transmittance = 512–521 nm); and PMT, photomultiplier tube.

2.2 Biological Sample

Wild-type (N2) C. elegans embryo samples were used to perform the THG 4-D recordings. This nematode was selected because it is a widely used model organism for many different research fields. Its short life cycle, small size, transparency, invariant cell lineage, and ease of maintenance in the lab make this model organism highly attractive for microscopy studies such as embryogenesis. The C. elegans completes its life cycle in three and a half days at 20 °C (Ref. [28]). Therefore, its development is relatively fast (once the eggs are laid). Given this, hatching will take place in approximately 13 h (Ref. [28]).

N2 wild-type C. elegans were grown in nematode growth media and fed with OP50 (Escherichia coli). Worms were synchronized, and embryos were obtained for the imaging experiments. Embryos from different developmental stages were mounted on a poly-L-lysine-Coated (Poly-L-lysine hydrobromide, Sigma Aldrich) thin coverslip (No. 0 thickness) with 10 μl of M9 buffer. The embryos where sandwiched between two thin glass coverslips, which were separated with 50-μm polystyrene microspheres (polymer microsphere suspension, Thermo Scientific). All this was done to avoid damaging and altering the embryos’ original shape. Samples were sealed with melted paraffin for sample stability and were imaged at constant room temperature (20 °C).

Control dishes containing nonirradiated C. elegans eggs were used to take into account other constraints that might prevent the embryos from developing. The survival rate for a total of 416 unexposed embryos was 89.2%.

2.3 Two- and Three-Dimensional Time-Lapse Imaging

Given that the maximum scanning speed of our galvanometric mirrors is \( \sim 300 \) lines/s, a single 2-D image of 500 \( \times 500 \) pixels could be acquired in less than 2 s. For the purpose of this study and in order to improve the signal-to-noise ratio, one 2-D optical section is made of 10 averaged scans. (The whole process takes less than 20 s.)

To create a 3-D reconstruction of our specimen, a series of 2-D optical sections separated 1.3 μm apart were acquired. The choice of this parameter was based on the theoretical point spread function (PSF) axial resolution calculation (resulting value of 1.3 μm). Given that the average sizes of the eggs are 60 \( \times 30 \times 30 \) μm, 24 planes are required to cover the whole specimen in the axial direction. In practice, 52 planes were taken into account for possible displacements of the sample. Thus, one full 3-D image was taken in approximately 15 min.

As no reference for the stress induced in the embryo or of sample deterioration at this specific wavelength is known, sample exposition and signal quality were adjusted according to the setup capabilities.

The images were volume rendered to allow for 3-D reconstruction employing ImageJ software. This procedure was repeated for different time points to enable a time-lapse 3-D reconstruction and further analysis.

3 Results and Discussion

A total of 27 C. elegans embryos, at different developmental stages, were imaged. An emitted THG signal was observed in all the development stages as well as dynamic changes in its position and intensity along the different phases of the C. elegans wild-type embryos. Figure 3 depicts the C. elegans stages, starting from the cell division process inside the hermaphrodite, passing through the gastrula, coma, and multiple-fold stages (previous to the hatching process), and the L1 larva (after the hatching process).

At the early development stages, the emitted signal was generated from the individual cells (see Fig. 3a). In later developmental stages occurring between thecoma and the two-fold stages (see Fig. 3b), the nerve and muscle cells start dif-

Fig. 2 THG images taken from several C. elegans, all at different developmental stages during the morphogenesis process. The THG signal is depicted in yellow. (Color online only.)

Fig. 3 THG images of several C. elegans embryos, all in the cell division stage. These images show identifiable dividing cells and nuclei (black circles inside delimited cells). The images depict (a) four-cell-stage egg; (b) and (c) multiple-cell-stage eggs. THG signal is depicted in yellow. (Color online only.)
ferentiating. At this stage, the embryo begins to move inside the egg. In this case, the signal arose mainly from the outer part of the embryo, becoming more restricted to tissue/structure. In what follows, all this will be further explained.

Figure 3 shows different embryos at the cell division stage (i.e., before gastrulation occurs). The emitted THG signal was observed in all the embryo cells (having the nuclei as a dark region due to its homogeneous nature); thus, every cell can be outlined. During these stages, the cells contain lipid and carbohydrate depositions (i.e., vacuoles shown in bright yellow spots), which are internal storages that allow the embryos’ development in the absence of external sources of energy. Due to these depositions, the embryos are able to complete the morphogenesis/embryogenesis process. These lipid depositions have already been suggested to give rise to the THG signal.

In Fig. 4 we show a 2-D time-lapse recording (see Video 1 for the animated time-lapse sequence) of a group of embryos at different developmental stages. (The background of the image is due to the M9 buffer–glass interface.) At the beginning of this time-lapse experiment, the acquired images allowed us to easily classify the embryos in two groups: the early stages, embryos labeled a and b, and the late stages (after ventral enclosure took place), embryos labeled c to f.

In this set of images, part of the C. elegans embryos’ elongation process at the different developmental stages can be imaged. For the embryos in the early stage, this is particularly clear in embryo a, as it has been strategically positioned ventrally. For the more developed embryos, c to f (two- and threefold-stage embryos), this process can also be appreciated, although a detailed evolution cannot be imaged due to the intense movement, which is faster than our acquisition time (this will be further discussed later).

Moving forward in the developmental stages (starting from twofold stages, embryos c, d, e, and f in Fig. 4), the observed THG signal arose from a more superficial section of the embryo. This signal might be generated (in the twofold-stage embryo f) by tissues that could be body wall muscles or hypodermal cells (skin cells, ectoderm, etc.). For the more developed embryos c, d, and e (in threefold stage), such signal could possibly be attributed to the cuticle formation. The correct identification of the exact signal source is not within the scope of this work, which aims only to show the potential of the technique.

To further show the potential of this methodology, we proceed to obtain 4-D images of developing embryos. Figure 5 is a volume-rendered model of a C. elegans sample (rotated for a perspective view) taken at different times. In this figure, the 3-D time-lapse sequence enables us to follow the morphological evolution of five embryos (a and b earlier developed, c to e late developed).
In embryos a and b, the THG signal arose from the inner spots, which had a turning on and off process. This could be caused by the cell deaths, migrations, or fusions. All these processes occur particularly during the morphogenesis processes, or due to the loss of membrane integrity. The THG signal has enormous potential, not only because we are obtaining intrinsic signal from different structures inside the embryo, but also because we are able to record embryogenesis/morphogenesis processes in vivo. This work, in agreement with previous studies, has proven that THG modality at 1550 nm can be used to follow developmental processes of living unstained specimens (Drosophila melanogaster and Zebrafish) for several hours. This is possible due to its optical noninvasive nature.

To further explore the induced effects of the 1550-nm wavelength, the survival rates of the embryos after long-term imaging exposure were studied. These rates were strongly dependent on the actual embryo development stage and the beginning of the time-lapse experiments. Thus, several long-term exposure trials (including the samples shown in this work) evaluating embryos’ survival rates were performed. The samples were imaged for several hours and afterward left unexposed for another period of time to be able to hatch.

Based on our results, we were able to classify the specimen behavior in two main groups: embryos before twofold stage (N=12), and embryos after twofold stage (N=15). On one hand, the embryos before twofold had a survival rate of 41% (compared to our control sample). During the first hours, these embryos presented a normal development. Morphological processes such as gastrulation, cell migration, and fusion were recorded. However, in some cases, after long-term sample exposition, there was some slow down in development. A possible cause for this performance could be that during this stage, the embryo lacks differentiated tissue (i.e., hypodermic/ectodermic), making the differentiating cells more vulnerable to this kind of radiation.

On the other hand, and contrary to the less developed embryos, the twofold- and threefold-stage embryos had a survival rate of 73% (compared to our control sample). This was confirmed as the more developed embryos presented continuous muscular movement during the whole imaging process. In these experiments, the embryos presented a normal development without any detectable morphological defects (observable in the 2-D and 3-D volume-rendered images of Figs. 3-5 respectively). Moreover, as previously mentioned, even when threefold stage embryos were exposed up to 6 h, they were able to hatch normally. Based on these results, a possible cause for this performance could be that the development of structures like hypodermal/ectodermic tissue acts as a protection to infrared (IR) light exposure.

Although the hatching and the viability of the embryos and larvae cannot be considered an ideal indicator for the absence of possibly induced damage with this particular wavelength, our survival rate percentages, for the purpose of this work, were used as indicators of inflicted embryo damage. Moreover, a close anatomical observation of L1 larvae with DIC post-4-D imaging and the integrity of anatomical and behavioral characteristics of the larvae sustain our previous statement of reduced damage.

This work, in agreement with previous studies, has proven that THG modality at 1550 nm can be used to follow developmental processes of living unstained specimens (Drosophila melanogaster and Zebrafish) for several hours. This is possible due to its optical noninvasive nature.

The application of this laser wavelength in vivo, and more importantly, its use for long-term 4-D imaging experiments (to record developing organisms such as C. elegans embryos) has enormous potential, not only because we are obtaining intrinsic signal from different structures inside the embryo (employing the sample interfaces as a contrast agent), but also because we are able to record embryogenesis/morphogenesis processes in vivo.

As previously demonstrated, embryogenesis/morphogenesis processes can take several hours. However, employing this particular wavelength, no apparent damage
was caused to the sample. These results were not expected, because it is known that IR wavelengths above 1500 nm are associated with increased water absorption in tissue, and may lead to unwanted heating of biological specimens. Additionally, due to the employed wavelength, the lateral and axial resolutions are reduced approximately by a factor of 2 when compared to that obtained with a Ti:Sapphire laser (at 800 nm and the same objective lens, NA=1.3). Given this, one might think that this wavelength cannot be used for high-resolution imaging of living samples, nor for long-term imaging experiments. However, in our case, and in agreement with other works, the laser used is able to produce an efficient THG signal from the sample using very low average powers. All this helps reduce the expected damage. In addition, the generated THG signal (516 nm) falls in the peak sensitivity region of most PMTs (i.e., Hamamatsu, H9305-04 peak sensitivity at 530 nm), allowing for an efficient signal collection. Another important and practical issue is that by imaging in living samples, e.g., Caenorhabditis elegans, this technique has the potential to provide enough information to easily identify precursors of different tissues. Importantly, this technique has the potential to be used as a tool for cell tracking when no fluorescent markers are available or desired for a particular reason. This type of information is relevant for cell lineage studies where these are tracked to determine which cells are precursors of different tissues. Importantly, this technique has the potential to provide enough information to easily identify the imaged structures without having to employ the previously mentioned complex image analysis algorithms to track the development of each structure.

Note that a frequency-doubled version of the same fiber laser source could be used for a multimodal microscopy approach, allowing the implementation of other techniques such as TPEF, SHG, etc. Adding these techniques will comprise an improved and supporting tool that would provide complementary information useful to investigate the source of the THG signal.

Last, the exact sources of the obtained THG signal in all development stages in the C. elegans embryos shown in this study still need to be investigated, and this is certainly a challenge in future work. Once the THG signal sources (in all the embryos’ developmental stages and in the larvae) are identified, the requirement of using different reporters expressed in different cell types to track these types of processes could be bypassed. Then, the use of this technique, and this particular wavelength, could be used as a noninvasive technique to study different dynamic processes such as the development of certain cell/tissue precursors.

4 Conclusions

To the best of our knowledge, this is the first time that this wavelength is applied in THG imaging to perform morphogenesis/embryogenesis studies in living C. elegans embryos. We have demonstrated that using the 1550-nm wavelength, we are able to track in vivo morphological processes for several hours.

The fact that the THG emission is produced at 516 nm makes this wavelength very attractive for THG microscopy because many conventional detectors have their peak sensitivity within this range. Therefore, it enables the third-harmonic signal collection without having to employ specialized or UV-grade optics. In addition, the use of fiber-based laser systems will enable a simpler implementation on NLM, as they are compact, robust, and inexpensive.

THG microscopy provides the exact spatial localization of individual cells due to the 3-D sectioning capability. Importantly, its resolution enables the identification of individual cells and different structures present in the whole development cycle of the C. elegans worm. Therefore, it has the potential to be used to track the cells’ positions and divisions during the morphological process. Given this, it could be possible to apply this methodology to perform cell lineage studies.

Certainly, this technique reduces the complexity of sample preparation required by other imaging techniques (fluorescence, confocal, and TPEF), as well as the interference with the specimen (genetic modification, fluorescent transgenes, dyes, etc.). Although the source of the THG signal has not been identified in all the developmental stages of the C. elegans embryo, we have shown that it is possible to outline a great potential use of this technique.

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