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After the first use of a surgical microscope in clinics by otolaryngologists in the early 20th century, the surgical microscope was regarded as an essential tool in an operating room.1 Despite significant advances in the surgical microscope technique, typically, the magnified surface image can only be provided by the microscope, resulting in missing subsurface information. However, noninvasive visualization of the subsurface during surgeries is crucial in some clinical applications, such as ophthalmic surgery and neurosurgery.2,3 However, due to the limitations, current surgical procedures heavily rely on the surgeons’ experience. For instance, during penetrating keratoplasty, surgeons typically incise the cornea at 560 μm from the epithelium layer under the guidance of a conventional surgical microscope. Because the surgeons are not able to visualize the clear corneal layers using the surgical microscope, severe side effects, such as corneal perforation and consequent poor eyesight recovery, can be caused if the cross-section area of cornea is inhomogeneous.4,5

Optical coherence tomography (OCT) was first introduced from MIT in the early 1990s.6 The principle of OCT is based on a low-coherence Michelson interferometer, and thus, OCT can provide cross-sectional images of microstructures in biological tissues. OCT offers high-resolution, noninvasive, nondestructive, and real-time imaging capabilities. Many studies have shown that OCT is a powerful tool in ophthalmology, cardiology, gastroenterology, oncology, dermatology, and dentistry.7 As another application, OCT has been used to overcome and/or assist the restriction of the current surgical microscope. In 2005, OCT was utilized as a surgical tool for anterior segment surgery.8 In addition, a handheld OCT probe has been used for macular surgery in 2009.9 However, these systems have only been used to monitor the operating regions not during the surgeries but before and after, resulting in missing the real-time information of lesions. Recently, an integrated OCT and surgical microscope system has been developed to guide vitreoretinal surgery in real time.10,11 Further, a real-time intraoperative OCT system with a graphics processing unit (GPU) was utilized in microsurgery guidance.12,13 Although these integrated systems offered an opportunity to monitor the surgical process in real time, they did not provide the OCT and microscope images at the same time on the same view. Thus, an additional display tool for the OCT images was required, which was cumbersome during the operation.

In this paper, we developed virtual intraoperative surgical OCT (VISOCT) by combining commercial clinical surgical microscope and spectral-domain OCT (SD-OCT). The VISOCT system could simultaneously acquire, process, and display OCT images through a GPU. The processed OCT images were projected back onto the microscope view plane via our homemade optical systems, and the images were visualized through the ocular lenses mounted on the microscope, not through a tabletop display. In our approach, no additional display to show OCT images is required, and the surgical procedures can be much simpler and truly real-time compared to the existing approach. We have successfully monitored and conducted stimulated penetrating keratoplasty of a live rabbit using the VISOCT system in vivo. Potentially, our VISOCT system can accurately guide surgeries in real time in clinics.

Figure 1(a) shows the experimental setup of our VISOCT system. First, the SD-OCT system was composed of an optical fiber-based Michelson interferometer using a broadband superluminescent diode [(SLD), SLD-34-HP, Superlum, Carrigtwohill, Ireland], and a low-coherence interferometer based on the self-heterodyne technique. The SD-OCT system was able to measure the axial resolution, 1.2 μm in air, and the optical bandwidth of 80 nm (centered at 1310 nm). The backscattered light from the sample was collected by an objective lens (20×, NA 0.4, Nikon Inc., Melville, NY), entering into a 50/50 beamsplitter (BS-50/50, Newport, Irvine, CA), and then divided into two branches. One of the branches was sent through a tunable Fabry-Perot interferometer (TFPI) to stabilize the optical path difference (OPD) in the interferometer. The output of the interferometer was a single-frequency light, and its wavelength was controlled by adjusting the temperature of the TFPI. The other branch was sent through a self-heterodyne interferometer (SHI). In the SHI setup, the signal beam and a local oscillator (LO) beam were combined at a home-made BS (BS-HM, Newport, Irvine, CA) and sent through the same optical path as the signal beam. The beam was recombined at BS-HM and was sent through a high-speed photodetector (HSPD, New Focus, San Jose, CA). The reference signal, a signal from the TFPI, was sent through a bandpass filter (BPF) and was used as a trigger signal. After the trigger signal, an integration time of approximately 80 μs was performed, and the power was measured. The measured power was then converted to the reflectance by calibration, and the depth profile of the sample was calculated by the swept-source OCT algorithm. The swept-source OCT system was connected to an OCT microscope (OCT Microscope, Newport, Irvine, CA) via an optical fiber (200 μm, Newport, Irvine, CA). A polarizer (P, Newport, Irvine, CA) was used to control the polarization state of the incident light. A half-wave plate (HWP) (New Focus, San Jose, CA) was used to control the intensity of the incident light. In the OCT microscope, the output light from the swept-source source was sent through a BS (BS-M, Newport, Irvine, CA) and was sent through a 3× objective (f = 10 mm, NA = 0.3, Newport, Irvine, CA). The OCT images were formed on a display screen (D), which was transmitted to the surgeon in real time.

The OCT images were displayed on the display screen (D) of the OCT microscope in real time. However, the OCT images showed limited information from the subsurface of the sample. To overcome this problem, we developed a virtual display system (VDS) that could display and enhance the OCT images using the OCT microscope and a graphics processing unit (GPU) (Nvidia K20, Nvidia, Santa Clara, CA). The optical coherence images of the live rabbit eye were captured by the OCT system and were sent to the SW (Fig. 1). The OCT images and the images of the laser pointer were simultaneously displayed on the display screen (D) of the OCT microscope with a GPU (Nvidia K20). The laser pointer was used to enhance the depth perception of the OCT images. The SW was able to control the laser pointer function, depth perception, and the OCT images.
The OCT images were acquired and processed using a center wavelength of 850 nm and a bandwidth of 50 nm. The SLD light was split into reference and sample arms through a 50:50 optical fiber coupler (FC850-40-10-APC, Thorlabs, Newton, New Jersey). A spectrometer consisted of a transmission type diffraction grating (1800 Ipmm, Wasatch photonics, Logan, Utah), a focusing lens (AC508-075-B, Thorlabs), and a collimator. Interference OCT signals were acquired by a 12-bit line scan CMOS camera with 4096 pixels (Sprint SPL4096-140K, Balser, Ahrensburg, Germany). We utilized a full-range $k$-linearization method to compensate for the spectrometer's nonlinearity. The OCT images were acquired by a frame grabber (PCIe-1429, National Instruments, Austin, Texas). Axial and lateral resolutions were 8.7 and 30.2 μm, respectively.

Then, we built the VISOCT microscope by adapting a commercial ophthalmic surgical microscope as shown in Fig. 1(a). The VISOCT system comprised three main parts: (1) OCT scanning, (2) display, and (3) beam splitting subsystems. Each subsystem is indicated with numbers 1, 2, and 3, respectively. The corresponding photograph is shown in Fig. 1(b). The OCT scanning subsystem 1 contained a collimator, galvo scanner (GVS001, Thorlabs), objective lens (AC508-075-B, Thorlabs), and dichroic mirror (NTS5-233, Edmund, Barrington, New Jersey). The dichroic mirror (750 to 1125 nm) was designed to reflect near-infrared light (i.e., OCT light). Thus, the reflected visible light easily transmitted through the dichroic mirror and an ocular lens, and was visualized by eyes. The display subsystem 2 containing a beam projector (SP-H03, Samsung, Seoul, South Korea) and the beam splitting subsystem 3 were designed to project the OCT image back onto the microscopic view plane via the microscope ocular lens. A beam splitter was located inside a custom-made mount adapted with a standard microscope mount.

To display the OCT images in real time, we coded image processing software based on both a GPU (Geforce GTX480, NVIDIA, Santa Clara, California) and central processing unit (Core 2 Quad Processor Q8200, Intel, Santa Clara, California). The image processing duties, such as $k$-domain linearization, background removal, fast Fourier transformation, and log scaling processes, were performed in the GPU using 480 Compute Unified Device Architecture processors. It was programmed by a C++ programming language. The OCT images with 1024 × 512 pixels along $z$ and $x$ direction, respectively, were recorded at a frame rate of 102 Hz.

To demonstrate the performance of the real-time VISOCT system in vivo, we performed simulated penetrating keratoplasty incising with a surgical blade [Figs. 2(a) and 2(b), Video 1] and suturing with a surgical needle and thread [Figs. 2(c) to 2(f), Video 2 and 3] in the cornea of a rabbit. All animal experimental procedures were conducted under the laboratory animal protocol permitted by the institutional animal care and use committee. A healthy rabbit (*Oryctolagus cuniculus*) weighing ~3.15 kg was utilized for the in vivo animal experiments. The rabbit was anesthetized by intravenous injection of ketamine (50 mg/kg body weight). The scanning range of one B-scan OCT image is 20 mm along the $x$ direction, while the field of view of the conventional microscopy images is 40 × 40 mm along the $x$ and $y$ axes, respectively.

Figures 2(a), 2(c), and 2(e) indicate the screenshots obtained via the ocular lens during the surgery, while Figs. 2(b), 2(d), and 2(f) show the magnified OCT images of them, respectively. First, the cornea incision was processed by the surgical blade. As shown in Video 1, the OCT image was clearly back-projected onto the left side in the microscopic view plane in real time, and we easily incised the rabbit cornea while simultaneously monitoring the cornea structures and the magnified OCT images.

**Fig. 1** (a) Experimental setup of a real-time virtual intraoperative surgical optical coherence tomography (VISOCT) microscope. (b) Photograph of the VISOCT probe. OL, objective lens; L, lens; PC, polarization controller; DM, dichroic mirror; G, galvo scanner; BS, beam splitter; C, collimator; M, mirror; NF, neutron-density filter; BP, beam projector; SM, surgical microscope.

**Fig. 2** Real-time VISOCT in vivo for stimulated penetrating keratoplasty. (a), (c), and (e) are the screenshots, acquired via ocular lens, of overlaid OCT and surgical microscopy images of the presented stimulated keratoplasty procedure in the right cornea of the rabbit (Video 1, MPEG, 3.92 MB) [URL: http://dx.doi.org/10.1117/1.JBO.19.3.030502.1], (Video 2, MPEG, 6.17 MB) [URL: http://dx.doi.org/10.1117/1.JBO.19.3.030502.2], and (Video 3, MPEG, 5.62 MB) [URL: http://dx.doi.org/10.1117/1.JBO.19.3.030502.3]. (b), (d), and (f) are the magnified OCT B-scan image cut from (a), (c), and (e), respectively. MIC, microscope; EP, epithelium; STR, stroma; EN, endothelium; IS, incision site; I, iris; F, forceps; N, needle; ST, surgical thread.
microscope images. After incising the cornea, watery fluid of the anterior chamber came out immediately. Then, an iris and cornea were attached. Moreover, the condition of the incised area was clearly visualized in the OCT image in real time [Figs. 2(a) and 2(b)]. Second, the incised area was sutured by the surgical needle and thread. As shown in Video 2, thanks to the real-time OCT image in the surgical view, we correctly aligned a forceps on the incised cornea layer. Then, we punctured the cornea layer using the surgical needle and thread [Figs. 2(c) and 2(d)]. The OCT image provided the correct position information of the cornea layer and precisely guided the needle insertion. Finally, under the guidance of real-time OCT imaging and display, the surgical thread was precisely strung together. As shown in Video 3, the OCT image showed the alignment of the cornea layer by tightening the threads [Figs. 2(e) and 2(f)]. We could control the strength of a knot based on the real-time OCT and finished the surgery by cutting the remaining thread. Note that the OCT image provided subsurface information of the cornea structures and the movement of the surgical instruments, whereas the microscopy image supplied only surface information. These results imply that our VISOCT system will be extremely useful in the practical surgical operating situation by minimizing unnecessary procedures during the surgeries.

In this study, we demonstrated a new concept of real-time VISOCT system for real-time OCT image projection on the microscopic view plane via the surgical ocular lens. The performance of VISOCT system was validated by showing a simple surgery in vivo. We believe that these developments will be crucial to enhance the usability of the surgical OCT system in the real surgical operating environment. Currently, our VISOCT system shows only two-dimensional B-mode OCT images. In this case, it is possible to miss the correct surgical position of the needle of blade. Future possible solutions are as follows: (1) We will add an aiming beam to guide the needle or blade intervention. (2) We will provide real-time three-dimensional OCT images by enhancing the image acquisition and display rates. Then, the VISOCT system would be significantly beneficial in neuroscience and ophthalmology.

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