Multimodality Raman and photoacoustic imaging of surface-enhanced-Raman-scattering-targeted tumor cells

Wei Shi
Robert J. Paproski
Peng Shao
Alexander Forbrich
John D. Lewis
Roger J. Zemp
Multimodality Raman and photoacoustic imaging of surface-enhanced-Raman-scattering-targeted tumor cells

Wei Shi,a Robert J. Paproski,a,b Peng Shao,a Alexander Forbrich,a John D. Lewis,b and Roger J. Zempa,*

aUniversity of Alberta, Department of Electrical and Computer Engineering, Second Floor ECERF, 9107-116 Street, Edmonton, Alberta, T6G 2V4, Canada
bUniversity of Alberta, Department of Oncology, 114 Street and 87 Avenue, Edmonton, Alberta, T6G 2E1, Canada

Abstract. A multimodality Raman and photoacoustic imaging system is presented. This system has ultralow background and can detect tumor cells labeled with modified surface-enhanced-Raman-scattering (SERS) nanoparticles in vivo. Photoacoustic imaging provides microvascular context and can potentially be used to guide magnetic trapping of circulating tumor cells for SERS detection in animal models. © The Authors. Published by SPIE under a Creative Commons Attribution 3.0 Unported License. Distribution or reproduction of this work in whole or in part requires full attribution of the original publication, including its DOI. [DOI: 10.1117/1.JBO.21.2.020503]

Keywords: multimodality imaging; Raman spectroscopy; surface-enhanced Raman scattering; nanoparticles; photoacoustic; biomedical optics.

Paper 150833LR received Dec. 15, 2015; accepted for publication Jan. 26, 2016; published online Feb. 25, 2016.

Specific and sensitive detection of individual tumor cells in vivo with ultralow background detection is challenging despite many recent attempts. In vivo fluorescence-based detection suffers from significant background autofluorescence. Bioluminescence offers ultralow background but requires genetic prelabeling of cells. Photoacoustic flow cytometry was developed to image hemoglobin oxygen saturation ($\text{SO}_2$) and oxygen partial pressure ($\text{pO}_2$) in vivo in single blood vessels with high spatial resolution. However, they did not demonstrate the capability to detect CTCs. Recently, we demonstrated SERS-based detection of magnetically trapped CTCs in vitro. Here we extend this work to include a multimodality photoacoustic-SERS imaging platform and to demonstrate ultralow-background Raman-detection of SERS-targeted tumor cells in vivo. The PAM subsystem enables detailed visualization of the microvascular structure and potentially can guide the placement of a trapping magnet. The SERS subsystem is capable of detecting single tumor cells if they are targeted with more than 200 SERS nanoparticles per cell.

To target widely overexpressed folate receptors on cancer cells, we conjugated both SERS nanoparticles (S440 & S421, BD Technologies) and magnetic fluorescent silica particles (screenMAG-Thiol, chemiccil GmbH) with folate. Simply, folate-PEG5k-maleimide (0.8 mM, PG2-FAML-5k, Nanocs Inc.) and methoxy-PEG2k-maleimide (4 mM, Sigma Aldrich) were added to thiolated SERS nanoparticle solution (1.3 nM, a 60 nm diameter Au core, and a 30 nm silica shell). After mixing and rocking the solution overnight at room temperature, excess folate-PEG-maleimide was removed by three runs of centrifugation (14,000 RPM, 5 min). Similar process was applied to magnetic nanoparticles (1.25 nM, 500 nm diameter magnetic fluorescent silica particles).

The diagram of our imaging system is demonstrated in Fig. 1. A 785 nm Raman laser RL (FB-785-350-FS-FS-1-1, RGBLase LLC) with 0.4 nm 20 dB spectral line-width and up to 350 mW output (while the power used in our experiment was 54 mW) was used as excitation laser source for SERS imaging. The excitation laser beam transmitted through a dichroic beam splitter BS (DMSP805L, Thorlabs) toward mirror M and was then focused onto the sample by an aspheric lens L2 (A240TM-B, Thorlabs, 0.5 NA, 8 mm focal length) with large NA to enable high collection efficiency needed for sensitive SERS detection. Sample was placed on a three-axis translation stage, controlled by a four-axis PCI stepper/servo motion controller (NI PCI-7354, National Instruments). A 1 in. tall, 1 in. diameter N50 cone magnet (Cone0100N, SuperMagnetMan) was pointed at the sample, generating strong magnetic field with high magnetic field gradient for magnetic trapping. SERS generated by sample was collected by lens L2, reflected by mirror M, and beam splitter BS. A 0.3 NA lens (AC127-019-B-ML, Thorlabs) focused SERS beam into a custom Raman imaging spectrometer (RASPEC-785-HR, P&P Optica Inc.), with a 15 μm × 26 mm slit as input aperture (f#: 3.5). A 10 cm−1 spectral resolution within spectral range of 815 to 895 nm was obtained using a 900 l/mm gel-grating in the Raman imaging spectrometer. A liquid-cooled electron-multiplying (EM) CCD camera (DU971N-UV, Andor Technology) was implemented for detection and signal acquisition, with very low dark current noise (0.0002 e-/pix/s at −100°C) and ~50% quantum efficiency at peak wavelengths, ~30% in the wavelength range we used. There were 1600 × 400 pixels on the EMCCD array, with 16 × 16 μm element size. The EM gain was set at 10, integration time was 1 s, and sensor temperature was liquid.

*Address all correspondence to: Roger J. Zempa, E-mail: Zemp@ualberta.ca

Downloaded From: https://www.spiedigitallibrary.org/journals/Journal-of-Biomedical-Optics on 4/8/2018 Terms of Use: https://www.spiedigitallibrary.org/terms-of-use
cooled to −100°C. SERS signals were finally acquired by imaging software (SOLIS, Andor Technology Ltd.). A custom-designed PAM probe was placed on the side of the sample opposite from lens L2 and magnet MN to achieve PAM imaging. The 10 Hz 532 nm laser pulses were generated by a Q-switch laser source (Nd6000, Continuum) and was focused by an objective lens (M Plan Apo, 5x, NA = 0.14, f = 20 cm, Edmund Optics Inc.). A beam combiner consists of a 15 mm aluminum-coated prism (BRAP15-A, Newport Co.) and a 15 mm uncoated prism (RA 15 mm UNCTD TS, Edmund Optics Inc.) was used for reflecting 532 nm laser beam onto the sample, while transmitting generated photoacoustic signals from sample to a 50 MHz ultrasound transducer (V214-BB-RM, Olympus Corp.) for acoustic detection. An acoustic lens L3 (f = 6 mm, NA = 0.5, Edmund Optics Inc.) is attached to the top surface of the beam combiner. Received photoacoustic signals were then amplified by an ultrasound pulse-receiver (5900PR, Olympus NDT Inc.). A high speed data acquisition card (CS8289, Gage Cobra, Gage Applied Systems, Inc.) with 12-bit dynamic range and up to 125 MSamples/s sample rate acquired the PAM laser trigger, the feedback positions of the translation stages, and the photoacoustic signals.

In our experiment, Raman imaging of folate-receptor overexpressing HeLa cells (cervical cancer and folate receptor positive) was performed by labeling HeLa cells with SERS and magnetic nanoparticles (which were both folate-conjugated), magnetically trapping cells in the focal zone of lens 2, and acquiring SERS signals from the magnetic focal zone. Simply, we took SERS spectrum from the SERS nanoparticle solution as a reference spectrum, which was then used for demixing of SERS spectrum obtained from all samples by implementing a classic linear least-squares method with an optional positivity constrain to quantify SERS signals. A concentration unit was used in Ref. 3 for simple comparison. Further data processing and analysis were conducted by MATLAB programs (Mathworks, Inc.) in the data acquisition computer.

In our Raman and photoacoustic imaging system, the Raman imaging subsystem works similar to the single modality Raman imaging system described in our previous publication with ∼1 pM SERS nanoparticle detection sensitivity, single-cell detection capability, high multiplexing ability, and 1:12 nonspecific binding ratio reported between HeLa and ZR-75-1 cell lines. Very little false-positive signal was observed from blood cells. For in vitro detection of CTCs, we used two types of nanoparticles: folate-receptor-targeted SERS nanoparticles and folate-receptor-targeted magnetic nanoparticles. Only cells targeted with both SERS and magnetic nanoparticles were trapped and detected. Previous work, however, used prelabeled tumor cells, did not demonstrate in vivo imaging and lacked another modality for spatial registration. The present work incorporates a photoacoustic imaging subsystem, investigates trapping of CTCs without prelabeling, and illustrates in vivo imaging and CTC detection.

In phantom studies, after incubating with serum-free medium containing folate-conjugated SERS nanoparticles and magnetic nanoparticles at 37 °C for 4 h, ∼5 × 10^4 HeLa cells were mixed with 1 mL rat blood, flowing inside a 200 μm inner-diameter acrylic capillary tubing (CT200-250-5, Paradigm Optics) at 3 cm/s flow velocity controlled by a syringe pump (NE-300, New Era Pump Systems, Inc.). Figure 5A shows a 1 mm × 1 mm coregistered Raman and photoacoustic image of rat blood mixed with SERS/magnetic nanoparticle-labeled HeLa cells flowing and magnetically trapped inside tubing. The cone magnet pointed at the lower left corner of the image. PAM and SERS signals are demonstrated in red and blue pseudocolor maps, respectively. The lateral resolution of the PAM imaging subsystem was measured as ∼50 μm by imaging a ∼7.5 μm carbon fiber.

To further demonstrate the capability of applying our method for the detection of CTCs under native targeting process, instead of incubating HeLa cells with nanoparticles before imaging, we first mixed HeLa cells with rat blood, and then added with folate-conjugated SERS nanoparticles and magnetic nanoparticles. HeLa cells (∼1 × 10^5 or ∼5 × 10^4) were first mixed with 1 mL rat blood for 10 min. Rat blood (1 mL) without HeLa cells was used as a control. The three groups of rat blood samples were then incubated with folate-conjugated SERS nanoparticles (∼0.4 nM, 20 μL) and magnetic nanoparticles (∼0.4 nM, 20 μL) before flowing the samples inside a 200 μm inner-diameter acrylic capillary tubing at 3 cm/s flow velocity. Detected SERS signal versus magnetic trapping time is plotted in Fig. 5B. Rat blood samples mixed with HeLa cells exhibited increasing SERS signals with magnetic trapping time, while SERS signals from rat blood sample without HeLa cells stayed close to the detection limit of our system throughout the experiment. This demonstrated specific targeting of folate-conjugated nanoparticles to folate receptor positive HeLa cells.

For in vivo studies, experimental animal procedures were conducted in conformity with the laboratory animal protocol which is approved by the University of Alberta Animal Use and Care Committee. A breeding anesthesia system (E-Z Anesthesia, Euthanex Corp.) was used to anesthetize the rats with isoﬂuorane during imaging acquisition. After incubated with serum-free medium-containing folate-conjugated SERS and magnetic nanoparticles at 37 °C for 4 h, ∼5 × 10^4 HeLa cells were mixed with 50 μL PBS solution. The solution was then subcutaneously injected into a rat ear. Figure 6A shows a 7 mm × 7 mm rat ear vasculature imaging by PAM, and co-registered Raman imaging of injected HeLa cells, with pseudocolor to illustrate PAM and SERS signals in red and blue color maps, respectively.
In our experiments, the Raman excitation laser power of 54 mW is slightly less than that of 60 mW reported by the other group. In our in vivo experiment with rat ear, to avoid over exposure to the skin, the laser beam was scanned over a 7 mm × 7 mm illuminated area to acquire imaging. Therefore, the corresponding power density by Raman excitation laser used was ∼0.11 W/cm², which is under the suggested ANSI maximum permissible exposure limit of 0.2 mW/cm² for a 785 nm laser with an exposure duration time from 10 s to 8 h. In addition, no apparent skin burns of mice were observed after laser exposure.

To demonstrate the ability to detect magnetically trapped CTCs, we injected ∼1 × 10⁵ HeLa cells into the chorioallantoic membrane of an ex ovo chicken embryo. The cells were prelabeled with both SERS and magnetic nanoparticles. A cone magnet was used for magnetic trapping over large vessels away from the injection point. The vessels used for trapping were visible by eye (such placement is not typically possible in many other organisms with highly scattering skin and tissues, hence the value of the photoacoustic subsystem). The Raman subsystem was used to detect the accumulation of magnetically trapped tumor cells labeled with SERS nanoparticles. While no signal was apparent initially, after 10 min, detectable SERS signals were visible in the trapping region, but not in areas away from the trapping zone. With only SERS or only magnetic nanoparticles, no accumulated SERS signals were measurable, consistent with our previous work. Figures 3(b), 3(c), and 3(d) show the SERS signal spectrum obtained from the magnetic trapping region of chicken embryo membrane, (c) spectrum obtained away from magnetic trapping region, and (d) a photograph of magnetic trapping and SERS detection setup using chicken embryo, respectively.
region, the spectrum obtained away from the magnetic trapping region, and a photograph of magnetic trapping and SERS detection setup with a chicken embryo, respectively.

In summary, we demonstrated the capability of our multimodality SERS and photoacoustic system to specifically trap and detect CTCs in vitro without pretargeting and, to detect trapped CTCs in vivo, and to provide structural information of surrounding vasculature. We previously determined ~pM sensitivities of the Raman detection system. If cells are labeled with more than 200 SERS nanoparticles, single-cell detection is thus possible. Present work focuses on Raman detection of magnetically trapped CTCs. Similar to previous work, photoacoustic imaging could also be used for CTC detection; however, a higher repetition-rate laser may be required. Presently with a 10 Hz laser current imaging time of our system is limited to 160 s for a 2 × 2 mm field of view. Future work would be to combine Raman excitation laser beam with photoacoustic excitation beam so as to co-focus on the sample from the same side of the sample, which will enable applying this system further to imaging thick targets in reflection mode. This technique could be used for studying or detection of CTCs, providing molecular information along with structural or functional vasculature imaging. We believe that the current study offers critical feasibility data to motivate future work on creating new multimodality imaging tools.

Acknowledgments
The authors gratefully acknowledge funding from a Prostate Cancer Canada Movember Discovery Grant, NSERC (355544-2008, 375340-2009, STPGP 396444), the Canadian Cancer Society (CCS 2011-700718), the Canada Foundation for Innovation, Leaders Opportunity Fund (18472), Alberta Advanced Education & Technology, Small Equipment Grants Program (URSI09007SEG), Alberta Ingenuity and Alberta Innovates scholarships for graduate students, Alberta Innovates Technology Futures Postdoctoral Fellowship Program, Canadian Federation of University Women Edmonton Margaret Brine Graduate Scholarships for Women and SPIE Optics and Photonics Education Scholarship.

References

Journal of Biomedical Optics

February 2016 • Vol. 21(2)

020503-4