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Abstract. This study is to test anatomic correlates, including connecting cilium (CC) and inner segment (IS) ellipsoid, to the hyper-reflective band visualized by optical coherence tomography (OCT) and commonly attributed to the photoreceptor inner/outer segment (IS/OS) junction. A line-scan OCT (LS-OCT) was constructed to achieve sub-cellular resolution (lateral: ~2 μm; axial: ~4 μm) of excised living frog retinas. An electro-optic phase modulator was employed for rapid and vibration-free phase modulation. Comparison of normalized distance measurements between LS-OCT images and histological images revealed that the dominant source of the signal reported as the IS/OS OCT band actually originates from the IS. © 2012 Society of Photo-Optical Instrumentation Engineers (SPIE). DOI: 10.1117/1.JBO.17.6.060504

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Given the excellent capability to identify morphological changes at individual functional layers of the retina, optical coherence tomography (OCT) has an increasing application in eye disease detection. In-depth understanding of anatomic sources of the OCT signal is essential for accurate interpretation of clinical outcomes, and is valuable for instrument optimization to improve imaging sensitivity and selectivity. Early studies have disclosed four hyper-reflective OCT bands at the outer retina, i.e., photoreceptor side. Anatomic sources of these four OCT bands have been typically attributed as follows: first (1st) band at the outer limiting membrane (OLM); second (2nd) band at the photoreceptor inner/outer-segment (IS/OS) junction; third (3rd) band at the posterior tip of the OS; and fourth (4th) band at the retinal pigment epithelium (RPE). Anatomic correlates of the 2nd band remains controversial. The 2nd OCT band is widely attributed to the IS/OS junction, which cell biologists consider the connecting cilium (CC) between these structures. However, comparative alignment of OCT bands with an anatomically correct model of the outer retina suggested an alternative correlate, i.e., the IS ellipsoid to the 2nd OCT band. Previous investigation has indicated that the physiological condition may affect the 2nd OCT band recording. Therefore, freshly isolated living retinas were used for OCT imaging.

Leopard (Rana pipiens) frog was selected for this study. Large photoreceptors (cone: ~3 μm and rod: ~6 μm) of the frog enabled accurate OCT recording at sub-cellular resolution. The experimental procedures were approved by the Institutional Animal Care and Use Committee of the University of Alabama at Birmingham. Without complications of ocular aberrations and signal contamination of other ocular tissues, isolated retinas provided a simple preparation to allow high resolution OCT characterization of the retina. Details of the preparation procedures of freshly isolated living retinas, which have been used to investigate stimulus-evoked retinal neural activities, have been previously reported. During the OCT imaging, the isolated retina was immersed in oxygenated Ringer’s solution to maintain its viability.

In order to achieve sub-cellular resolution in both lateral and axial directions, a rapid line-scan OCT (LS-OCT) was developed. The LS-OCT combined technical merits of our recently demonstrated electro-optic phase modulator (EOPM) based functional OCT and line-scan confocal microscopy. Figure 1 shows a simplified diagram of the LS-OCT. In this system, a near infrared superluminescent laser diode (SLD-35-HP, Superlum) with center wavelength λ = 830 nm and bandwidth Δλ = 60 nm was used for light illumination. A water immersion objective (10×, NA = 0.3) was applied for OCT imaging. The lateral resolution of the system was estimated at ~2 μm (0.61λ/NA), and the axial resolution of the LS-OCT was ~4 μm (0.44λ2/nΔλ), where refractive index n of the sample was assumed ~1.4). Using a mirror immersed in water solution, the actual axial resolution was measured at ~5 μm.

In the illumination path [optics above the beam splitter BS, Fig. 1(a)], a cylindrical lens CL1 was employed to condense the collimated SLD light in one dimension to produce a focused line illumination which was imaged at the retina through lenses L3, L4, L5, and the objective OB. The active area of the high-speed (70,000 lines/s) linear camera (spL2048-140k, Basler Sprint) was conjugated to the focused line illumination. The line-sensor (1 ⨰ 2048 pixels, 10 × 10 μm2) of the linear camera naturally acted as a spatial-filter to reject out-of-focus light. Thus, a LS confocal mechanism was achieved to reduce background light and therefore maximize effective OCT contrast and dynamic range.

In the reference path [optics below the BS, Fig. 1(a)], another cylindrical lens CL2 was used to convert the focused light back to a collimated light beam before entering the EOPM (Model 350-50, Conoptics) that was used to generate rapid vibration- and inertia-free phase modulation. A four-step phase-shifting strategy was employed to retrieve OCT images. The glass
Fig. 1 (a) Schematic diagram of the LS OCT. BS: beam splitter; CO: collimator; L1 ~ L5: lenses; OB: objective; CL1 and CL2: cylindrical lenses. The focal length of CL1, CL2, L1 ~ L5 was 40, 40, 40, 80, 80, 40, and 75, respectively. (b) The side view of the optical geometry of the illumination and reference paths (dashed square in a). The focal length of CL1, CL2, L1 ~ L5 was 40, 40, 40, 80, 80, 40, and 75, respectively. (b) The side view of the optical geometry of the illumination and reference paths (dashed square in Fig. 1(a)). In the side view, the cylindrical lenses CL1 and CL2 acted as a pair of glass plates. Our custom-designed LS-OCT provided both en-face and B-scan recording capabilities. The galvo scanning mirror (GVS001, Thorlabs) could provide rapid en-face [Fig. 2(a)] imaging up to 400 Hz; while the motorized sample platform (Z825B, Thorlabs) could scan in the z-direction to achieve B-scan recording [Fig. 2(b)] of the cross-section of flat-mount retinas.

Figure 2 shows representative en-face and B-scan OCT images of isolated retina in which the retinal pigment epithelium (RPE) was not included. During the recording, the photoreceptors were upward, i.e., facing to the objective, to improve image quality of the outer retina. Given the sub-cellular resolution, the en-face image [Fig. 3(a)] revealed individual photoreceptors. B-scan image [Fig. 3(b)] disclosed individual layers from the photoreceptor to inner retina. After adapting band nomenclature used in high resolution OCT of human retina to frog retina, we labeled individual layers. As shown in Fig. 3(b), the OS, ‘IS/OS’ (presumed), OLM, OPL, inner plexiform layer (IPL), and nerve fiber layer (NFL) were hyper-reflective; while the outer nuclear layer (ONL), inner nuclear layer (INL), and ganglion cell layer (GCL) were hypo-reflective. The bright ‘IS/OS’ band was bumpy in the high resolution OCT image. In order to achieve better visualization of individual OCT bands, Fig. 3(c) shows an averaged cross-section image over 120 μm (200 B-scan images, with 0.6 μm recording interval) along the y-axis. We consistently observed that the whole OS section, not only the OS tip or the ‘IS/OS’ junction, produced OCT signal; and the band of lower reflectivity [red arrow in Fig. 3(c)] was observed located sclerally to the 2nd OCT band. Moreover, bright spots [red arrows in Fig. 3(d) and (e)], with dimension at cellular level, were consistently observed. This might reflect reflectance inhomogeneity of different photoreceptors in the frog retina that has two types each of cones and rods.

Figure 3 shows comparative histological [Fig. 3(a) and 3(b)] and LS-OCT [Fig. 3(c)] images. For histological examination, DNA was stained with DAPI (blue) with Prolong Gold antifade mounting reagent (Invitrogen) in 10-μm thick fixed retinal cryosections. The DNA marked fluorescence image and the differential interference contrast (DIC) image were superimposed. Based on previously reported light [3] and electron [4] microscopy studies, the photoreceptors were categorized into two types: short (S) and long (L) cones. Our LS-OCT imaged the S and L cone photoreceptor mosaic in terms of size distribution and spatial position. The LS-OCT images revealed that the S cone photoreceptors were more closely packed than the L cone photoreceptors. The red arrow points to the local hypo-reflective band, bordering posteriorly on the ‘IS/OS’ band [red arrows in Figs. 3(b) and 3(c)]. Histogram of OCT distance ratio (μ IS/OS − μ OPL (IS)/μ OPL (OS) − μ OPL (IN)) and the OCT distance ratio (μ IS/OS − μ OCT(IS)/μ OCT(OPL) − μ OCT(IN)). Five samples were used for histological and OCT measurements, respectively. For each sample, six positions were measured. Error bars indicate standard deviations.

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microscopy investigations of frog retinas, the IS/OS, OLM, OPL, etc. were identified in Fig. 3(a). For the frog retina, the ONL could be identified as having two rows of nuclei that contains DNA. Therefore, IS ellipsoid portion was also stained by DAPI, and thus could be separated from other IS portions. Figure 4b and 4c show enlarged sub-images of the histological [Fig. 4a] and OCT [Fig. 4c] images. If the 2nd OCT band is correctly attributed to the IS/OS, it would be expected to align with anatomic counterpart in the histological image [Fig. 4d]. However, the center of the 2nd OCT band shifted toward to the IS, mostly like the IS ellipsoid portion. In contrast, the local hypo-reflective band [red arrow in Fig. 4c] might relate to the IS/OS junction.

In order to achieve quantitative analysis, the distance between the well-established OLM and OPL bands were used to normalize the axial position of the 2nd OCT band. Figure 5a shows 3 representative OCT longitudinal reflectance profiles (LRPs) as a function of retinal depth for three zones specified in Fig. 2d. Each zone was averaged over 50 µm along the x-axis to improve signal-to-noise ratio. In theory, the OCT bands could be widened due to the logarithmic transformation or gamma correction of OCT displaying. However, neither the logarithmic transformation nor gamma correction shifts the peaks. Therefore, the positions on the z-axis of peaks of the ‘IS/OS’ (2nd) band, the OLM (1st) band and the OPL band could be defined as their axial positions. Thus, OCT distance ratio (OCT/IS/OS − OCT/OLM)/(OCT/OLM − OCT/OPL) could be quantitatively computed and compared to histological distance ratio of (HIS/OS − HOM)/(HOM − HOP).

Figure 6c shows statistics of OCT and histological distance ratios. Five retinas were used for OCT imaging, and corresponding five retinas were used for histology measurement. For each sample, six lateral positions were measured for estimating the distance ratio. As shown in Fig. 6c, the histological distance ratio of (HIS/OS − HOM)/(HOM − HOP) was 1.0 ± 0.05. In contrast, the mean of (OCT/IS/OS − OCT/OLM)/(OCT/OLM − OCT/OPL) was 0.6 ± 0.1. The significant difference (t-test, p < 0.003) between the (OCT/IS/OS − OCT/OLM)/(OCT/OLM − OCT/OPL) and (HIS/OS − HOM)/(HOM − HOP) suggested that the 2nd OCT band might not actually relate to the IS/OS junction.

In summary, a LS-OCT was developed to demonstrate OCT imaging at sub-cellular resolution in both lateral [Fig. 2a, b] and axial [Fig. 2c, d] directions. Quantitative comparison of histological images and LS-OCT images revealed that the OCT distance ratio (OCT/IS/OS − OCT/OLM)/(OCT/OLM − OCT/OPL) was significantly smaller than the histological distance ratio (HIS/OS − HOM)/(HOM − HOP). The significant difference suggests that dominant source of the signal reported as the ‘IS/OS’ OCT band actually originates from the IS. We speculate that the IS ellipsoid, which consists of abundant mitochondria, or the CC extended into the IS might contribute to the observed 2nd OCT band signal.

Moreover, reflectance inhomogeneity was consistently observed at cellular level [Fig. 2a, b]. The bumpy 2nd OCT band shown in Fig. 2b might attribute to variable lengths and axial locations of different rod/cone photoreceptors. Robust OCT signal was observed through the whole length of the photoreceptor outer segment. Early spectral-domain OCT study (SD-OCT) did not report OCT signal in the outer segment, i.e., the depth between the presumed ‘IS/OS’ band and the posterior tip of the OS. We speculate that this discrepancy might attribute to large (0.3) numeric aperture (NA) of the LS-OCT system, compared to ~0.1 NA in typical SD-OCT system for retinal imaging. Increased NA could improve collection efficiency of reflected light, particularly scaterring light with possible large angle changes, relative to incident light, from outer segment discs. In addition, the large NA system might produce illumination light with incident angle greater than the optimized acceptance of the photoreceptor, which acted as a waveguide. Therefore, more light might be scattered and thus detected from the OS. We expect that further characterization of these observed OCT signals will provide insight for improved instrumental design, validated image attribution, and retinal diagnosis.

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References