Journal of Biomedical Optics

SPIEDigitalLibrary.org/jbo

Low-level laser therapy for spinal cord injury in rats: effects of polarization

Takahiro Ando
Shunichi Sato
Hiroaki Kobayashi
Hiroshi Nawashiro
Hiroshi Ashida
Michael R. Hamblin
Minoru Obara

Low-level laser therapy for spinal cord injury in rats: effects of polarization

Takahiro Ando,^a Shunichi Sato,^b Hiroaki Kobayashi,^c Hiroshi Nawashiro,^{c*} Hiroshi Ashida,^b Michael R. Hamblin,^{d,e,f} and Minoru Obara^a

^aKeio University, Department of Electronics and Electrical Engineering, 3-14-1 Hiyoshi, Kohoku-ku, Yokohama, Kanagawa 223-8522, Japan

^bNational Defense Medical College Research Institute, Division of Biomedical Information Sciences, 3-2 Namiki, Tokorozawa, Saitama 359-8513, Japan

^cNational Defense Medical College, Department of Neurosurgery, 3-2 Namiki, Tokorozawa, Saitama 359-8513, Japan

^dMassachusetts General Hospital, Wellman Center for Photomedicine, 40 Blossom Street, Boston, Massachusetts 02114

^eHarvard Medical School, Department of Dermatology, 55 Fruit Street, Boston, Massachusetts 02115

^fHarvard-Massachusetts Institute of Technology (MIT) Division of Health Sciences and Technology, 65 Landsdowne Street, Cambridge, Massachusetts 02139

Abstract. The effects of laser polarization on the efficacy of near-infrared low-level laser therapy for spinal cord injury (SCI) are presented. Rat spinal cords were injured with a weight-drop device, and the lesion sites were directly irradiated with a linearly polarized 808-nm diode laser positioned either perpendicular or parallel to the spine immediately after the injury and daily for five consecutive days. Functional recovery was assessed daily by an open-field test. Regardless of the polarization direction, functional scores of SCI rats that were treated with the 808-nm laser irradiation were significantly higher than those of SCI alone group (Group 1) from day 5 after injury. The locomotive function of SCI rats irradiated parallel to the spinal column (Group 3) was significantly improved from day 10 after injury, compared to SCI rats treated with the linear polarization perpendicular to the spinal column (Group 2). There were no significant differences in ATP contents in the injured tissue among the three groups. We speculate that the higher efficacy with parallel irradiation is attributable to the deeper light penetration into tissue with anisotropic scattering. © The Authors. Published by SPIE under a Creative Commons Attribution 3.0 Unported License. Distribution or reproduction of this work in whole or in part requires full attribution of the original publication, including its DOI. [DOI: [10.1117/1.JBO.18.9.098002](https://doi.org/10.1117/1.JBO.18.9.098002)]

Keywords: low-level laser therapy; photobiomodulation; polarization; spinal cord injury; functional evaluation.

Paper 130302PR received May 1, 2013; revised manuscript received Jul. 11, 2013; accepted for publication Aug. 7, 2013; published online Sep. 12, 2013.

1 Introduction

In spinal cord injury (SCI), complete or partial loss of autonomic, sensory, and motor functions is caused by interruption of neural signal conduction along the axonal tracts. There is generally poor recovery of these functions because of the difficulty of tissue regeneration in the central nervous system. Thus, SCI patients are left with serious residual disabilities, such as paralysis, respiratory difficulty, chronic pain, urinary problems, and neurologic decline, leading to considerable decrease in quality of life. Various strategies have been examined for repair of SCI in animal models, including blockage of the endogenous growth inhibitory factors,^{1,2} infusion of neurotrophic factors,^{3,4} and transplantation of growth promoting cells.^{5–7} However, no effective treatment for SCI has yet been established.

Low-level laser therapy (LLLT) is a promising approach to treat SCI. LLLT has been clinically applied to the treatment of rheumatoid arthritis and periodontal disease, pain management, and healing of wounds and burns.^{8–10} LLLT is also currently used for the treatment of various neurological diseases such as stroke, neurodegenerative diseases, and brain injury.^{11–16} Several studies have shown that near-infrared LLLT has the potential to be an effective noninvasive therapy for SCI.^{17–20}

Rochkind et al. demonstrated that transplantation of embryonal spinal cord nerve cells followed by 780-nm laser irradiation enhanced axonal sprouting and spinal cord repair in a completely transected rat SCI model.¹⁷ In two different rat models of hemisection SCI and contusion SCI, Anders et al. transcutaneously applied an 810-nm laser, which penetrated to the depth of the injured spinal cord and promoted axonal regeneration and functional recovery.^{18,19} Their study demonstrated that near-infrared laser irradiation significantly suppressed immune cell activation and cytokine/chemokine expression, suggesting that a decrease in the inflammatory response is one of the recovery mechanisms in LLLT for spinal cord repair.

The detailed mechanisms of LLLT are still under investigation. However, the therapeutic efficacy relies fundamentally on the initial photochemical event, i.e., absorption of photons by photoacceptors or chromophores such as cytochrome *c* oxidase in the tissue.^{21,22} Karu et al. showed in an *in vitro* study that the basic processes of LLLT occurring in HeLa cells were light absorption and photochemistry but that the incident characteristics of photons, such as degree of light polarization, did not affect the biological reactions in LLLT.²³ However, scattering of photons *in vivo* depends on the microstructure of tissue, and light propagation into biological tissue would therefore change the healing property. For instance, Ribeiro et al. investigated the repair of skin burns in rats with a linearly polarized He–Ne laser beam, which was parallel or perpendicular to the direction of the spinal column, at the same laser dose.^{24,25} Their results showed that the healing process was dependent on the polarization orientation; lesions irradiated with parallel

*Present address: Tokorozawa Central Hospital, Division of Neurosurgery, 753-2 Kitaakitsu, Tokorozawa, Saitama 359-0038, Japan.

Address all correspondence to: Shunichi Sato, National Defense Medical College Research Institute, Division of Biomedical Information Sciences, 3-2 Namiki, Tokorozawa, Saitama 359-8513, Japan. Tel: +81-42-995-1211; Fax: +81-42-991-1757; E-mail: shunsato@ndmc.ac.jp

polarization were completely repaired 17 days after wound creation, while those with perpendicularly polarized irradiation showed a moderate degree of healing in the same period. They attributed these results to the fact that the parallel polarization was aligned with the predominant orientation of collagen fibers in the dermis, which was confirmed by histological analysis. This alignment would reduce photon scattering and thus increase optical penetration depth in the tissue, leading to the acceleration and improvement of cutaneous wound repair.²⁴

It is widely known that photon scattering by aligned cylindrical structures, such as myofibrils, axons, and collagen fibers, results in anisotropic light reflection and propagation in the tissue.^{26–30} These characteristics are often used for diagnosis of the tissue abnormalities and for mapping of specific structures in the tissue.^{31–33} Since the spinal cord has a fibrous structure, photon migration should be affected by polarization of incident light in the tissue. However, the effect of polarization on efficacy of LLLT for SCI has not been elucidated. In the present study, we examined the effect of relative orientation of laser polarization on efficacy of near-infrared LLLT for contused spinal cords in rats.

2 Materials and Methods

2.1 Spinal Cord Injury Model

The protocol used in this study was approved by the Committee on Ethics of Animal Experiments in the National Defense Medical College. We used female Sprague-Dawley rats (Japan SLC Inc., Shizuoka, Japan) weighing 180 to 270 g. Before the operation, they were anesthetized by intraperitoneal injection of pentobarbital sodium (50 mg/kg animal weight). During all of the experiments using anesthesia, body temperature was monitored with a rectal probe and maintained at 37.0 to 37.5°C. The lamina of the tenth thoracic vertebra was surgically removed, and the spinal cord was exposed. A New York University weight-drop device was used to make a severe spinal cord contusion.^{34–36} A 10-g metal rod with a flat circular impact surface 2.5 mm in diameter was dropped from a height of 25 mm onto the exposed spinal cord.

2.2 Laser Irradiation

Immediately after making a contusion in the spinal cord, the lesion site was directly irradiated with an 808-nm diode laser beam (B&W Tek Inc., Newark, Delaware) that was transmitted through a polarizer (SPFN-30C-26, Sigma Koki Co., Ltd., Tokyo, Japan). The polarizer was held with a rotatable holder to change the direction of incident polarization onto the tissue. The incised skin was closed with sutures after laser irradiation. Exposure of the spinal cord, irradiation, and suturing were repeated daily for the following five consecutive days. The control animals (Group 1) received a weight-drop injury and the lesion was exposed daily but not irradiated with the laser beam. The injured tissue of Group 2 rats was irradiated with a linearly polarized laser perpendicular to the spinal direction (hereafter called perpendicular polarization) and that of Group 3 rats was treated with parallel-aligned polarization (hereafter called parallel polarization). The laser power measured at the injured spinal cord surface was 25 mW, with a spot diameter of 20 mm giving a power density of 8 mW/cm². Using an irradiation duration of 20 min, the light fluence per day was 9.6 J/cm².

2.3 Functional Evaluation

The motor function of hind limbs was evaluated by open-field testing and scored on the basis of the Basso-Beattie-Bresnahan (BBB) scale^{35,36} ($n = 12$ in each group); a score of 0 means no spontaneous movement, while a score of 21 indicates normal locomotion. Assessment of the animals was performed before laminectomy and 1, 2, 3, 5, 7, 10, 14, and 21 days after injury. The open field consisted of a squared arena (45 cm × 90 cm) with 20-cm-high walls. All rats received manual bladder expression before the open-field test to eliminate possible behavior differences due to bladder fullness.

2.4 Histological Analysis

After spinal injury, glial cells are intrinsically activated with enlarged somas and intensive expression of intermediate filament proteins over time as the inflammatory response.^{37,38} The activated astrocytes compose a glial scar, forming a cystic cavity in the region surrounded by the scar.^{37,38} This neurodegenerative nature leads to a progressive increase in the size of the cavitation area.³⁹ Thus, a cavity is closely connected with inflammation. Since the locomotor recovery is critically correlated with the percentage of remaining normal nerve fibers in spinal tissue,^{40,41} suppression of the excessive inflammatory responses and progressive increase in the lesion sizes is necessary. Thus, we evaluated cavity area as the most important histopathological outcome.

Under systemic anesthesia, rats were euthanized 21 days after injury by transcardial perfusion with 150 mL physiological saline followed by further perfusion with 200 mL 4% paraformaldehyde in physiological saline. Segments of the spinal cords centering on the injury were removed and postfixed in the same fixative overnight. The tissues were then frozen in an optimal cutting temperature compound (Sakura Finetek USA Inc., Torrance, California) and sectioned to 10- μ m-thick slices with a cryostat microtome. For histological images (HE staining) of the longitudinal sections at the lesion epicenter, cavity areas were manually outlined and quantified by image analysis using Adobe Photoshop 7.0 imaging software (Adobe Systems, San Jose, California) ($n = 9$ in each group).

2.5 ATP Content Measurement

Increase in ATP synthesis is one of the important indicators for evaluating the effect of LLLT on enhancement of mitochondrial function.²² Immediately after near-infrared laser irradiation, we harvested traumatized spinal tissues (length, ~ 1 cm) and measured ATP contents in the tissues with an ATP assay kit (TA100, Toyo Ink., Tokyo, Japan) according to the manufacturer's instructions ($n = 6$ in each group). Intact spinal tissues (normal) and injured tissues after laminectomy without laser irradiation (Group 1) were also harvested and analyzed for comparison. Spinal tissue was homogenized in 1 mL of 4-(2-hydroxyethyl)-1-piperazine-ethanesulfonic acid buffer, and the solution was centrifuged at 15000 rpm for 15 min at 4°C to pellet insoluble materials, followed by addition of the ATP-extraction buffer provided in the kit to a portion of the lysis solution. After shaking and incubating for 30 min at room temperature, the supernatant was mixed with luciferin, and then luminescence of an aliquot was measured with a luminometer (LB955, Berthold Technologies, Bad Wildbad, Germany). The concentration of ATP was calculated using the ATP standard curve

and expressed as nmol per mg protein. Protein concentrations in all spinal samples were determined using a protein assay system (500-0112, BioRad, Richmond, California).

2.6 Distribution of Light Transmitted Through Spinal Tissue

To compare penetrations of light with different polarization directions through the spinal cord, light transmitted through excised spinal tissue was imaged with a CCD camera (XC-7500, Sony Corp., Tokyo, Japan). The experimental setup is schematically shown in Fig. 1. A fresh spinal column removed from an uninjured rat (diameter, ~5 mm; length, 12 mm) was placed on a black plastic sheet (thickness, 0.7 mm) with a rectangular hole (3 mm × 9 mm) through which polarized laser light was directed onto the bottom surface of the spinal column ($n = 2$). The transmitted light was detected from the top. The polarization direction was changed by rotating a polarizer that was placed between the fiber output end and the spinal tissue; 0 and 180 deg means the incident direction of the linear polarized laser was parallel to the spinal column, while 90 deg indicates the polarization direction was perpendicular to the spinal column. The laser power measured at the bottom surface of the spinal cord was 25 mW. The transmitted light was quantified by calculating white-colored pixels in the regions of interest (ROIs, 3 mm × 9 mm), corresponding to the size of a rectangular hole in the plastic sheet.

2.7 Statistical Analysis

The results of functional evaluation were compared between the groups using two-way repeated analysis of variance (ANOVA) with Tukey's *post hoc* test. Statistical analysis for the results of cavity area and ATP content measurement was performed using one-way factorial ANOVA followed by Tukey's *post hoc* test. A value of $P < 0.05$ was regarded as statistically significant.

3 Results

3.1 Functional Recovery

Figure 2 shows the BBB scores for rats in the three groups as a function of time after injury. Regardless of the polarization direction, the BBB scores of the rats receiving 808-nm laser irradiation (Groups 2 and 3) were significantly higher than those of

Fig. 1 Experimental setup for measurement of light transmitted through an excised spinal tissue.

Fig. 2 Locomotive scores based on an open-field test as a function of days after SCI. Values are expressed as means + standard error of the mean (SEM) ($n = 12$). Asterisks indicate significant differences ($P < 0.05$).

the SCI alone group (Group 1) from five days after injury (Group 1 versus Group 2, $P = 0.003$ at day 5; $P = 0.002$ at days 7 and 10; $P = 0.000$ at days 14 and 21, Group 1 versus Group 3, $P = 0.000$ at days 5, 7, 10, 14, and 21). In addition, BBB scores of the rats irradiated with parallel polarization (Group 3) were significantly higher than those of the rats treated with perpendicular polarization (Group 2) from 10 days after SCI ($P = 0.029$ at day 10; $P = 0.005$ at day 14; $P = 0.003$ at day 21). The averaged BBB scores at three weeks post-SCI were 7.5 ± 1.2 for Group 1, 9.8 ± 1.0 for Group 2, and 10.9 ± 1.0 for Group 3.

3.2 Histologic Evaluation

Figure 3 shows histological images (HE staining) of longitudinal sections of the spinal cords of rats in all groups at three weeks after injury. Formation of a cavity was observed in the spinal cord tissue of all rats. Figure 4 shows the results of quantitative analysis of the cavity areas in injured spinal cords on the basis of histological images. Cavity areas for the rats in Groups 2 and 3 were significantly smaller than those for the rats in Group 1; $P = 0.050$ between Groups 1 and 2 and $P = 0.006$ between Groups 1 and 3, while $P = 0.639$ between Groups 1 and 2. These results show that irradiation with both the parallel and perpendicular polarized laser can lead to reduced formation of glial scar and cavity. However, there was no significant difference in cavity area between Groups 2 and 3.

3.3 ATP Content

Figure 5 shows the ATP content in the spinal cords of rats in Groups 1, 2, and 3, where the value for normal rats is also shown for comparison. In the normal rats, baseline ATP content in the spinal tissues was 0.13 ± 0.02 nmol per mg protein, compared with 0.07 ± 0.02 nmol per mg protein in the untreated SCI rats (Group 1). The ATP contents in rats of Groups 2 and 3 were 0.08 ± 0.02 and 0.09 ± 0.02 nmol per protein, respectively. However, there was no significant difference in ATP content between Groups 1, 2, and 3, indicating that

Fig. 3 Histological images (HE staining) of longitudinal sections of injured spinal cords at three weeks after injury: (a) Rat with SCI alone (Group 1). (b) SCI rat treated with perpendicularly polarized laser to the spinal column (Group 2). (c) SCI rat treated with parallel polarization (Group 3). Scale bars indicate 200 μm .

ATP synthesis immediately after LLLT was not associated with improved motor function by near-infrared light either with parallel or perpendicular polarization.

3.4 Light Transmitted Through the Spinal Cord

Figure 6 shows the distributions of light transmitted through an excised spinal cord under two incident polarization conditions: (a) perpendicular and (b) parallel to the spinal direction. Figure 6(c) shows the amount of transmitted light from the ROI as a function of incident laser polarization direction. The amount of transmitted light for parallel polarization was ~ 1.8 -fold higher than that for perpendicular polarization, indicating that light with parallel polarization penetrated deeper in the spinal tissue than did light with perpendicular polarization.

4 Discussion

The current study has shown that locomotive scores of SCI rats with 808-nm laser treatment were significantly higher than those of SCI rats without light irradiation from day 5 after injury onward regardless of the incident polarization direction (Fig. 2). Anders et al. demonstrated that transcutaneous application of 810-nm nonpolarized laser significantly promoted axonal

Fig. 4 Results of quantitative analysis of injury cavity areas in spinal cords on the basis of histological images. Values are expressed as means $\pm$ SEM ($n = 9$ in each group). * $P < 0.05$, ** $P < 0.01$.

regrowth at six weeks postinjury in a rat hemisection SCI model¹⁸ and functional recovery at three weeks after injury in a rat contused SCI model.¹⁹ There are differences in the time course of treatment efficacy between our study and their studies. In their experiments, the incident laser power and daily dosage at the skin surface overlying the lesion site were 150 mW and 1589 J/cm² (irradiation duration, 2997 s), respectively, 6% of which (9 mW and 95 J/cm²) penetrated to the spinal cord depth. The irradiation was applied daily for 14 consecutive days after SCI.^{18,19} They concluded that the improved axonal regeneration was caused by inhibiting inflammatory cell activity due to laser irradiation at high dosage per day (> 10 J/cm²). In our study, on the other hand, a linearly polarized laser was directly applied to the exposed spinal cord lesion immediately after trauma and then daily for the following five days at the fluence of 9.6 J/cm² (power, 25 mW; irradiation duration, 1200 s). The therapeutic effects of near-infrared laser irradiation have been reported to be dependent on dosage, being associated with production of anti-apoptotic, pro-proliferative, antioxidant, and angiogenic factors.^{22,42–44} Although further study is needed to clarify the therapeutic mechanisms, as well as the optimum irradiation conditions for treating SCI, a different mechanism might work for treatment under the laser irradiation conditions in the present study.

Locomotor function of SCI rats treated with parallel polarization (Group 3) was significantly improved when compared

Fig. 5 ATP contents in normal and injured spinal cords (Groups 1, 2, and 3). Results are expressed as means $\pm$ SEM ($n = 6$ in each group).

with that irradiated with perpendicular polarization (Group 2) from day 10 after trauma (Fig. 2). To investigate the reason for the polarization-dependent treatment efficacy, we evaluated the cavity area 21 days after injury (Figs. 3 and 4), ATP content immediately after SCI (Fig. 5), and light penetration in the spinal tissue (Fig. 6). There was no statistically significant difference either in cavity area or in ATP content between the two polarization groups. On the other hand, the polarization dependence of light transmission through tissue was remarkable; the amount of light transmitted through the spinal cord with perpendicular polarization was $\sim 40\%$ less than that with parallel polarization [Fig. 6(c)]. Thus, we speculate that the higher treatment efficacy with parallel polarization is attributable to the more efficient light propagation through tissue, which is consistent with the results reported by Hebeda et al.²⁶ The penetration depth of red light (wavelength, 632.8 nm) with polarization parallel to white matter tracts in a fresh bovine brain was significantly greater than that of red light polarized perpendicular to myelinated fiber tracts; the effective attenuation coefficients of light (μ_{eff}) were $0.47 \pm 0.06 \text{ mm}^{-1}$ with parallel polarization and $0.63 \pm 0.13 \text{ mm}^{-1}$ with perpendicular direction ($P < 0.05$).²⁶ It is known that locomotor recovery after SCI is highly correlated with the volume of remaining normal nerve fibers in spinal tissue.^{40,41} In the present study, LLLT with parallel polarization would have provided more efficient protection of neural cells from apoptosis or necrosis in deep-located anterior horns than LLLT with perpendicular polarization. Further investigation is needed to elucidate the detailed physiological contributions of linearly polarized light, but the present study clearly demonstrated the importance of light polarization in LLLT for SCI.

Moreover, Silva et al. recently reported that when the rat tendon was irradiated with linearly polarized 632.8-nm laser aligned in parallel with the tendon long axis, the tissue showed higher birefringence and greater nonsusceptibility when compared with those of the nonirradiated tissue. They attributed

this to the changes in molecular order and aggregation states of the collagen fibers.⁴⁵ With linearly polarized laser, the tendon was irradiated once in their work, while the spinal cord was irradiated daily for five consecutive days in our work. Similar alteration of the fibrotic structure, which affects direction of photon scattering, could take place in the spinal cord examined in this study.

Open surgery for direct laser irradiation to the spinal cord is invasive, which is not practical in clinical situations. However, there seems to be a chance for direct irradiation during surgical exposure for spinal decompression and endoscopic discectomy, for which laser delivery through an optical fiber should be useful. For fiber-based irradiation with a linearly polarized laser, a polarization filter can be placed at the laser output fiber end, or a polarization-maintaining fiber can also be used.

Although we observed a significantly higher treatment efficacy with parallel polarization than with perpendicular polarization, the difference is not large enough to indicate medical relevance, although our results would provide important data concerning effect of light polarization in LLLT for fibrotic tissue. It should also be noted that our data strongly support the efficacy of LLLT for SCI regardless of the polarization direction. In addition to transcutaneous LLLT,^{18,19} LLLT based on direct laser irradiation can also provide a therapeutic option for SCI since open surgery is often performed for decompression in conventional SCI treatment strategy.

5 Conclusion

We investigated the effects of polarization on efficacy of 808-nm LLLT for contusion SCI in rats. Rats treated with light for which polarization was parallel to the spinal direction showed significantly faster recovery of locomotor function than did rats treated with perpendicular polarization. We speculate that this is attributable to deeper photon penetration through spinal tissue with parallel polarization than with perpendicular polarization.

Acknowledgments

This work was supported in part by the Special Research Program of the National Defense Medical College. T. Ando is grateful for the JSPS Research Fellows for Young Scientists. M. R. Hamblin was supported by US NIH R01AI050875.

References

1. A. J. Kastin and W. Pan, "Targeting neurite growth inhibitors to induce CNS regeneration," *Curr. Pharm. Des.* **11**(10), 1247–1253 (2005).
2. B. Atalay et al., "Antibodies neutralizing Nogo-A increase pancytadherin expression and motor recovery following spinal cord injury in rats," *Spinal Cord* **45**(12), 780–786 (2007).
3. H. S. Sharma, "A select combination of neurotrophins enhances neuroprotection and functional recovery following spinal cord injury," *Ann. NY Acad. Sci.* **1122**, 95–111 (2007).
4. E. R. Hollis, II and M. H. Tuszynski, "Neurotrophins: potential therapeutic tools for the treatment of spinal cord injury," *Neurotherapeutics* **8**(4), 694–703 (2011).
5. D. D. Pearce et al., "cAMP and Schwann cells promote axonal growth and functional recovery after spinal cord injury," *Nat. Med.* **10**(6), 610–616 (2004).
6. D. Cizkova et al., "Transplants of human mesenchymal stem cells improve functional recovery after spinal cord injury in the rat," *Cell Mol. Neurobiol.* **26**(7–8), 1167–1180 (2006).
7. O. Tsuji et al., "Therapeutic potential of appropriately evaluated safe-induced pluripotent stem cells for spinal cord injury," *Proc. Natl. Acad. Sci. USA* **107**(28), 12704–12709 (2010).

Fig. 6 Images of laser light transmitted through excised spinal tissue: directions of incident linearly polarized laser were (a) perpendicular and (b) parallel to the spinal column. (c) Transmitted light distribution in ROIs [areas indicated by a broken line in (a) and (b)] as a function of the direction of incident linearly polarized laser with respect to spinal orientation. Values are expressed as means $\pm$ SD ($n = 2$).

8. A. Ekim et al., "Effect of low level laser therapy in rheumatoid arthritis patients with carpal tunnel syndrome," *Swiss Med. Wkly.* **137**(23–24), 347–352 (2007).
9. A. M. Shirani et al., "Low-level laser therapy and myofascial pain dysfunction syndrome: a randomized controlled clinical trial," *Lasers Med. Sci.* **24**(5), 715–720 (2009).
10. Z. Simunovic, A. D. Ivanovich, and A. Depolo, "Wound healing of animal and human body sport and traffic accident injuries using low-level laser therapy treatment: a randomized clinical study of seventy-four patients with control group," *J. Clin. Laser Med. Surg.* **18**(2), 67–73 (2000).
11. J. J. Anders, "The potential of light therapy for central nervous system injury and disease," *Photomed. Laser Surg.* **27**(3), 379–380 (2009).
12. L. Detaboada et al., "Transcranial application of low-energy laser irradiation improves neurological deficits in rats following acute stroke," *Lasers Surg. Med.* **38**(1), 70–73 (2006).
13. P. A. Lapchak et al., "Safety profile of transcranial near-infrared laser therapy administered in combination with thrombolytic therapy to embolized rabbits," *Stroke* **39**(11), 3073–3078 (2008).
14. A. Oron et al., "Low-level laser therapy applied transcranially to mice following traumatic brain injury significantly reduces long-term neurological deficits," *J. Neurotrauma* **24**(4), 651–656 (2007).
15. T. Ando et al., "Comparison of therapeutic effects between pulsed and continuous wave 810-nm wavelength laser irradiation for traumatic brain injury in mice," *PLoS One* **6**(10), e26212 (2011).
16. X. Wu et al., "Pulsed light irradiation improves behavioral outcome in a rat model of chronic mild stress," *Lasers Surg. Med.* **44**(3), 227–232 (2012).
17. S. Rochkind et al., "Transplantation of embryonal spinal cord nerve cells cultured on biodegradable microcarriers followed by low power laser irradiation for the treatment of traumatic paraplegia in rats," *Neurol. Res.* **24**(4), 355–360 (2002).
18. K. R. Byrnes et al., "Light promotes regeneration and functional recovery and alters the immune response after spinal cord injury," *Lasers Surg. Med.* **36**(3), 171–185 (2005).
19. X. Wu et al., "810 nm wavelength light: an effective therapy for transected or contused rat spinal cord," *Lasers Surg. Med.* **41**(1), 36–41 (2009).
20. S. Rochkind, "Photoengineering of neural tissue repair processes in peripheral nerves and the spinal cord: research development with clinical applications," *Photomed. Laser Surg.* **24**(2), 151–157 (2006).
21. T. I. Karu, "Primary and secondary mechanisms of action of visible to near-IR radiation on cells," *J. Photochem. Photobiol. B* **49**(1), 1–17 (1999).
22. Y. Y. Huang et al., "Biphasic dose response in low level light therapy," *Dose Response* **7**(4), 358–383 (2009).
23. T. I. Karu et al., "Elementary processes in cells after light absorption do not depend on the degree of polarization: implications for the mechanisms of laser phototherapy," *Photomed. Laser Surg.* **26**(2), 77–82 (2008).
24. M. S. Ribeiro et al., "Effects of low-intensity polarized visible laser radiation on skin burns: a light microscopy study," *J. Clin. Laser Med. Surg.* **22**(1), 59–66 (2004).
25. F. da Silva Dde et al., "Collagen birefringence in skin repair in response to red polarized-laser therapy," *J. Biomed. Opt.* **11**(2), 024002 (2006).
26. K. M. Hebeda et al., "Light propagation in the brain depends on nerve fiber orientation," *Neurosurgery* **35**(4), 720–722 (1994).
27. S. Nickell et al., "Anisotropy of light propagation in human skin," *Phys. Med. Biol.* **45**(10), 2873–2886 (2000).
28. A. Kienle, F. K. Forster, and R. Hibst, "Anisotropy of light propagation in biological tissue," *Opt. Lett.* **29**(22), 2617–2619 (2004).
29. A. Kienle and R. Hibst, "Light guiding in biological tissue due to scattering," *Phys. Rev. Lett.* **97**(1), 018104 (2006).
30. T. Yun et al., "Monte Carlo simulation of polarized photon scattering in anisotropic media," *Opt. Express* **17**(19), 16590–16602 (2009).
31. L. Larsen et al., "Polarized light imaging of white matter architecture," *Microsc. Res. Tech.* **70**(10), 851–863 (2007).
32. R. Liao et al., "Rotating linear polarization imaging technique for anisotropic tissues," *J. Biomed. Opt.* **15**(3), 036014 (2010).
33. R. Liao et al., "Penetration depth of linear polarization imaging for two-layer anisotropic samples," *Appl. Opt.* **50**(23), 4681–4687 (2011).
34. J. A. Gruner, "A monitored contusion model of spinal cord injury in the rat," *J. Neurotrauma* **9**(2), 126–128 (1992).
35. D. M. Basso, M. S. Beattie, and J. C. Bresnahan, "A sensitive and reliable locomotor rating scale for open field testing in rats," *J. Neurotrauma* **12**(1), 1–21 (1995).
36. D. M. Basso, M. S. Beattie, and J. C. Bresnahan, "Graded histological and locomotor outcomes after spinal cord contusion using the NYU weight-drop device versus transection," *Exp. Neurol.* **139**(2), 244–256 (1996).
37. M. Pekny and M. Nilsson, "Astrocyte activation and reactive gliosis," *Glia* **50**(4), 427–434 (2005).
38. M. V. Sofroniew, "Molecular dissection of reactive astrogliosis and glial scar formation," *Trends Neurosci.* **32**(12), 638–647 (2009).
39. C. J. Ek et al., "Spatio-temporal progression of grey and white matter damage following contusion injury in rat spinal cord," *PLoS One* **5**(8), e12021 (2010).
40. P. Schucht et al., "Anatomical correlates of locomotor recovery following dorsal and ventral lesions of the rat spinal cord," *Exp. Neurol.* **176**(1), 143–153 (2002).
41. S. W. You et al., "Spontaneous recovery of locomotion induced by remaining fibers after spinal cord transection in adult rats," *Restor. Neurol. Neurosci.* **21**(1–2), 39–45 (2003).
42. Y. Y. Huang et al., "Biphasic dose response in low level light therapy—an update," *Dose Response* **9**(4), 602–618 (2011).
43. H. Chung et al., "The nuts and bolts of low-level laser (light) therapy," *Ann. Biomed. Eng.* **40**(2), 516–533 (2012).
44. W. Xuan et al., "Transcranial low-level laser therapy improves neurological performance in traumatic brain injury in mice: effect of treatment repetition regimen," *PLoS One* **8**(1), e53454 (2013).
45. D. F. Silva et al., "Birefringence and second harmonic generation on tendon collagen following red linearly polarized laser irradiation," *Ann. Biomed. Eng.* **41**(4), 752–762 (2013).