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#### Abstract

We propose and objectively evaluate an inverse Monte Carlo model for estimation of absorption and reduced scattering coefficients and similarity parameter $\gamma$ from spatially resolved reflectance (SRR) profiles in the subdiffusive regime. The similarity parameter $\gamma$ carries additional information on the phase function that governs the angular properties of scattering in turbid media. The SRR profiles at five source-detector separations were acquired with an optical fiber probe. The inverse Monte Carlo model was based on a cost function that enabled robust estimation of optical properties from a few SRR measurements without a priori knowledge about spectral dependencies of the optical properties. Validation of the inverse Monte Carlo model was performed on synthetic datasets and measured SRR profiles of turbid phantoms comprising molecular dye and polystyrene microspheres. We observed that the additional similarity parameter $\gamma$ substantially reduced the reflectance variability arising from the phase function properties and significantly improved the accuracy of the inverse Monte Carlo model. However, the observed improvement of the extended inverse Monte Carlo model was limited to reduced scattering coefficients exceeding $\sim 15 \mathrm{~cm}^{-1}$, where the relative root-mean-square errors of the estimated optical properties were well within 10\%. © 2016 Society of Photo-Optical Instrumentation Engineers (SPIE) [DOI: 10.1117/1.JBO.21.9.095003]


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## 1 Introduction

Light that has propagated through a turbid medium carries an abundance of information about the sample structure and chemical composition encountered along the path, which is especially interesting when the turbid medium under investigation is a biological tissue. Reflectance techniques utilizing integrating spheres, ${ }^{1}$ hyperspectral imaging systems, ${ }^{2}$ and optical fibers or optical fiber probes ${ }^{3-5}$ are all frequently used to conduct the measurements. If the profile of the backscattered light is captured as a function of the distance from the illumination source, the measurements are said to be spatially resolved. Spatially resolved reflectance (SRR) measurements can be conducted by optical fiber probes with multiple source-detector separations (SDS). The captured SRR profile depends on the scattering and absorption properties of the turbid medium and is consequently frequently exploited for noninvasive determination of optical properties in biological tissues and other turbid media. ${ }^{6-11}$ Furthermore, different SDS are associated with different sampling depths. In general, a shorter SDS collects photons that propagate nearer to the sample surface and experience fewer scattering interactions with the turbid medium. The collected reflectance signal is considered subdiffusive when the spatial separation between the incident and the collected light becomes comparable to the transport mean free path length. ${ }^{12}$ In contrast, the diffusive photons experience many scattering interactions and lose the information of the initial propagation direction. SRR spectroscopy is said to operate in the subdiffusive regime, when a significant portion of the acquired reflectance signal is represented by the subdiffusive reflectance.

[^0]Due to fewer scattering events, the subdiffusive reflectance largely depends on the scattering phase function of the turbid medium. ${ }^{13-15}$ Since the scattering phase function is tightly related to the medium microstructure (i.e., the microscopic fluctuations of the refractive index ${ }^{16}$ ), the subdiffusive reflectance has the potential to reveal structural characteristics of biological tissues at a cellular level.

Many of the recent studies have focused on the reflectance spectroscopy operating in the subdiffusive regime. ${ }^{16-19}$ Since photons in this regime undergo only a few scattering interactions with the turbid medium, the commonly used diffusion approximation of light transport in turbid media, which assumes reflectance dependence only on the absorption $\mu_{\mathrm{a}}$ and reduced scattering $\mu_{\mathrm{s}}^{\prime}$ coefficients, is insufficient. To overcome these limitations and provide the means to account for the phase function in the light propagation model, direct solutions of the radiative transport equation ${ }^{20}$ or the Monte Carlo (MC) stochastic method ${ }^{21}$ have to be used. However, the full potential of light propagation models can be really appreciated when solving the inverse problem, where the optical properties are estimated from a set of reflectance measurements. In the most simplified case, SRR spectroscopy offers a way to independently estimate the absorption and reduced scattering coefficients for each spectral band. ${ }^{8}$ Nevertheless, in the subdiffusive regime, the inverse model parameters have to be extended to account for the phase function dependence. The existing studies have shown that extending the inverse model by an additional similarity parameter $\gamma=\left(1-g_{2}\right) /\left(1-g_{1}\right)$ should improve the estimation of optical properties. ${ }^{15,22}$ Here, $g_{1}$ and $g_{2}$ denote the first and second Legendre moments of the phase function. Physically,

[^1]$\gamma$ represents the relative contribution of large-angle scattering events and, for biological tissues, amounts to values between 1.6 and 2.4. ${ }^{16,22}$

In this paper, we propose and extensively evaluate an extended inverse Monte Carlo (IMC) model that, in addition to the absorption and reduced scattering coefficients, also incorporates $\gamma$. First, we introduce a new cost function (CF) that enables robust estimation of optical properties from the SRR measurements at five SDS in the subdiffusive regime. Subsequently, we examine the influence of the phase functions on the MC computed SRR. Next, the basic and extended IMC models are evaluated and compared on synthetic datasets of SRR computed by various commonly used phase functions. Finally, the extended IMC model is experimentally evaluated on turbid phantoms comprising molecular absorbers and polystyrene microspheres.

## 2 Materials and Methods

### 2.1 Monte Carlo Simulations

The light propagation was modeled by an MC stochastic approach derived from the code of Wang et al. ${ }^{21}$ and the CUDA-based implementation of Alerstam et al. ${ }^{23}$ In our study, two optical fiber probe geometries that included five SDS were introduced.

The first geometry (SG) was based on a simple laterally uniform probe-tissue interface, which takes into account only the mismatch between the refractive indices of tissue ( $n_{m}=1.33$ ) and optical fibers ( $n_{\text {fib }}=1.45$ ). Each simulation included $10^{8}$ photon packets, which were launched uniformly over the fiber opening and the solid angle defined by the numerical aperture ( $\mathrm{NA}=0.22$ ). Additionally, the detection scheme was modified to account for the numerical aperture of the detector fibers. Due to the uniformity of the probe-tissue interface, the detection scheme around the source fiber was divided into $5-\mu$ m-thick concentric annular rings, from which the number of detected photon packets through a particular detection fiber was estimated in the postprocessing step. In this way, a better signal-to-noise ratio was obtained compared to the exact geometry of the individual detection fibers.

The second geometry (RG) was based on a realistic probetissue interface that additionally considers (1) the reflections from the stainless steel probe tip and (2) the refractive index mismatch between the black epoxy resin ( $n_{\text {epoxy }}=1.6$ ) and the tissue (Fig. 1). Stainless steel reflectivity was set to $57 \%$ (measured value). Its effect on the SRR was already investigated and found significant. ${ }^{24}$ The photon packets $\left(10^{9}\right)$ were launched following the scheme of the first (SG) geometry. In contrast, the detection scheme was constrained to individual detection fibers located at five different SDS.

The MC simulations were conducted using a semi-infinite medium geometry. To speed up the simulations, all photon packets that drifted more than 0.8 cm laterally and axially from the source center were terminated. The two termination criteria were validated and proved consistent with the semi-infinite medium geometry.

### 2.2 Scattering Phase Functions and Their Implementation in Monte Carlo Simulations

Since the SRR in the subdiffusive regime significantly depends on the phase function, several phase functions that have been


Fig. 1 Linear array optical fiber probe used to acquire the SRR profiles in the subdiffusive regime. The materials and source-detector configuration of the probe are presented in different colors.
proposed for biological tissues were considered. In addition to the common Henyey-Greenstein (HG) phase function $p_{\mathrm{HG}}$, we used the modified Henyey-Greenstein (MHG) phase function, which also accounts for the angular dependence of the Rayleigh scattering: ${ }^{12}$
$p_{\mathrm{MHG}}(x)=\beta p_{\mathrm{HG}}(x)+(1-\beta) \frac{3}{2} x^{2}$,
where $x$ represents the cosine of the scattering angle and $\beta$ adjusts the relative contribution of the HG phase function and Rayleigh scattering. Another promising phase function for describing scattering in biological tissues is the Gegenbauer kernel (GK) phase function ${ }^{19,25,26}$

$$
\begin{align*}
p_{\mathrm{GK}}(x)= & 2 \alpha_{\mathrm{GK}} g_{\mathrm{GK}} \frac{\left(1-g_{\mathrm{GK}}^{2}\right)^{2 \alpha_{\mathrm{GK}}}}{\left[\left(1+g_{\mathrm{GK}}\right)^{2 \alpha_{\mathrm{GK}}}-\left(1-g_{\mathrm{GK}}\right)^{2 \alpha_{\mathrm{GK}}}\right]} \\
& \times \frac{1}{\left(1+g_{\mathrm{GK}}^{2}-2 g_{\mathrm{GK}} x\right)^{1+\alpha_{\mathrm{GK}}}}, \tag{2}
\end{align*}
$$

where $\alpha_{\mathrm{GK}}$ and $g_{\mathrm{GK}}$ are the parameters of the GK phase function. Additionally, we used phase functions for spherical particles based on the Mie theory, ${ }^{27}$ which depend on the diameter and the refractive index of the spherical particles, the refractive index of the surrounding medium and the wavelength of the incident light.

Existing MC light propagation simulations primarily use the HG phase function to sample the scattering angles when a photon packet undergoes a scattering event. In this aspect, the HG phase function is convenient, since it allows analytical expression of the cumulative probability distribution (CPD) from which the scattering angles can be sampled. Analytical expression for the CPD can also be obtained for the GK phase function. ${ }^{28}$ In contrast, the CPD of MHG and Mie phase functions can only be derived numerically, and thus, a lookup table-based approach was used for sampling the scattering angle. ${ }^{29}$

### 2.3 Inverse Monte Carlo Model

The IMC model based on the lookup table approach introduced by Palmer et al. and Hennessy et al. ${ }^{30,31}$ was used to estimate the optical properties from the acquired SRR profiles $\mathbf{R}=$ $R\left(\mathrm{SDS}_{1}\right), R\left(\mathrm{SDS}_{2}\right), \ldots, R\left(\mathrm{SDS}_{5}\right)$, where $R\left(\mathrm{SDS}_{i}\right)$ is the reflectance acquired at the $i^{\prime}$ th SDS. In both studies, the lookup tables were introduced by only considering $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$. A similar basic IMC (b-IMC) methodology was adopted in the initial part of this study. In order to comply with biological tissue
characteristics ${ }^{32}$, the lookup table values for each SDS ranged from 0 to $25 \mathrm{~cm}^{-1}$ for $\mu_{\mathrm{a}}$ and from 5 to $70 \mathrm{~cm}^{-1}$ for $\mu_{\mathrm{s}}^{\prime}$ using 30 uniform steps. The b-IMC model was based on the HG phase function with the anisotropy factor $g$ set to 0.8 . A constant value of $g$ was selected in accordance with the first similarity relation for $\mu_{\mathrm{s}}^{\prime}$, which states that regardless of the individual values of $g$ and scattering coefficient $\mu_{\mathrm{s}}$, the same values of $\mu_{\mathrm{s}}^{\prime}$ result in the same reflectance. ${ }^{33-35}$

It has been shown, however, that higher similarity relations are important when the SDS of the optical probe becomes comparable to the transport mean free path length. ${ }^{12}$ By following the lookup table methodology, the b-IMC model was extended (e-IMC) by an additional similarity parameter $\gamma$ that was proposed by Bevilacqua and Depeursinge. ${ }^{12}$ The e-IMC model lookup tables included a third dimension for $\gamma$, which spanned from 1.6 to 2.3 in 20 uniform steps, taking into account the biological variations. ${ }^{16,22}$ The e-IMC model was based on the GK phase function that allowed the full span of $\gamma$ values at $g=0.8$. On the other hand, the MHG and HG phase functions are less useful, since they only support $\gamma$ values of up to 2 , and HG phase function does not allow independent selection of $g$ and $\gamma$.

The simplified geometry (SG) was utilized by the IMC models when applied to the synthetic $\mathbf{R}$. In contrast, the realistic geometry (RG) was used, when the IMC models were applied to the measured $\mathbf{R}$.

As the number of independent measurements used in this study is relatively low (five different SDS), formulation of a new CF is essential for robust estimation of optical properties. In this study, two CFs were compared
$\mathrm{CF}_{1}=\sum_{i=1}^{5}\left\{R_{\text {meas }}\left(\mathrm{SDS}_{i}\right)-R_{\text {sim }}\left(\mathrm{SDS}_{i}\right)\right\}^{2}$,
and
$\mathrm{CF}_{2}=\sum_{i=1}^{5}\left\{\log \left[R_{\text {meas }}\left(\operatorname{SDS}_{i}\right)\right]-\log \left[R_{\text {sim }}\left(\operatorname{SDS}_{i}\right)\right]\right\}^{2}$,
where $R_{\text {meas }}$ and $R_{\text {sim }}$ are the measured and simulated reflectance, respectively. The choice of the second CF is based on the dependence of reflectance on the SDS, which decreases with the SDS for a few orders of magnitude. Each CF was minimized by a trust-region-reflective algorithm available in

MATLAB ${ }^{\circledR}$ (Mathworks Inc.) as a function Isqnonlin. ${ }^{36}$ Values of each optimized parameter were constrained to the range defined in the corresponding lookup table. An initial estimate was obtained by an exhaustive search over all the lookup table entries. Subsequently, the estimate was refined by optimization and spline interpolation of the SRR over the IMC model parameters.

### 2.4 Synthetic Datasets of Spatially Resolved Reflectance Profiles

To objectively evaluate the performance of the IMC models, synthetic datasets $\mathcal{R}$ of SRR profiles $\mathbf{R}$ were computed according to the parameters in Table 1. Additionally, the synthetic datasets included computed $\mathbf{R}$ of turbid phantoms, later used for validation (see Sec. 2.6, Table 2). Each phantom comprised 150 synthetic $\mathbf{R}$ for wavelengths from 450 to 800 nm .

It should be noted that an arbitrary combination of $g$ and $\gamma$ cannot be obtained for all the used phase functions. For HG phase function, the relation between $g$ and $\gamma$ (i.e., $\gamma=g+1$ ) results in only six valid combinations of $g$ and $\gamma$ (Table 1, second line). In the case of MHG phase function, a mathematical constrain (i.e., $\gamma<1+g$ ) leads to only 14 combinations (Table 1, third line). Lastly, combinations of $g$ and $\gamma,(0.75$, $2.25)$ and ( $0.95,1.65$ ), are not valid for the GK phase function. Thus, only 18 valid combinations of $g$ and $\gamma$ can be obtained (Table 1, fifth line). The datasets $\mathcal{R}_{\text {HG-I }}$ and $\mathcal{R}_{\text {GK-I }}$ were introduced for the performance verification of the b-IMC and e-IMC inverse models, respectively. For these datasets, the error of estimated parameters should be close to zero, given a proper formulation of the CF is employed.

### 2.5 Experimental Setup for Reflectance Measurements

SRR profiles in the subdiffusive regime were acquired by a cus-tom-made linear array optical fiber probe (FiberTech Optica Inc., Ontario, Canada) with five SDSs (220, 440, 660, 880, and $1100 \mu \mathrm{~m}$ ) and an outer diameter of 6.0 mm (Fig. 1). The diameter of the optical fiber cores and the numerical aperture of the fibers was $200 \mu \mathrm{~m}$ and 0.22 , respectively. The optical fiber probe was placed in a holder, which enabled repeatable repositioning of the probe within the cylindrical beaker containing the liquid turbid phantom. The cylindrical beaker with

Table 1 Optical properties used for synthetic datasets $\mathcal{R}$ of SRR profiles $\mathbf{R}$.

| Synthetic dataset | Phase function | $\mu_{\mathrm{a}}\left(\mathrm{cm}^{-1}\right)$ | $\mu_{\mathrm{s}}^{\prime}\left(\mathrm{cm}^{-1}\right)$ | $g$ | $\gamma$ | Number of R |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $\mathcal{R}_{\text {HG-I }}$ | Henyey-Greenstein | 0 to 25 (15 values) | 5 to 70 (15 values) | 0.8 | 1.8 | 225 |
| $\mathcal{R}_{\text {HG-II }}$ | Henyey-Greenstein | 0 to 20 <br> (5 values) | 10 to 60 <br> (5 values) | $\begin{aligned} & 0.7,0.75,0.8 \\ & 0.85,0.9,0.95 \end{aligned}$ | $\begin{aligned} & 1.7,1.75,1.8 \\ & 1.85,1.9,1.95 \end{aligned}$ | 150 |
| $\mathcal{R}_{\text {MHG }}$ | Modified Henyey-Greenstein | 0 to 20 <br> (5 values) | 10 to 60 <br> (5 values) | $\begin{gathered} 0.75,0.8,0.85 \\ 0.9,0.95 \end{gathered}$ | $\begin{aligned} & 1.65,1.75 \\ & 1.85,1.95 \end{aligned}$ | 350 |
| $\mathcal{R}_{\text {GK-I }}$ | Gegenbauer Kernel | 0 to 25 (10 values) | 5 to 70 (10 values) | 0.8 | 1.65 to 2.25 <br> (10 values) | 1000 |
| $\mathcal{R}_{\text {GK-II }}$ | Gegenbauer Kernel | 0 to 20 <br> (5 values) | 10 to 60 <br> (5 values) | $\begin{gathered} 0.75,0.8,0.85 \\ 0.9,0.95 \end{gathered}$ | $\begin{aligned} & 1.65,1.85, \\ & 2.05,2.25 \end{aligned}$ | 450 |

Table 2 Microsphere number density $n$, reduced scattering coefficient $\mu_{\mathrm{s}}^{\prime}$ and absorption coefficient $\mu_{\mathrm{a}}$ at 630 nm for each phantom P comprising $0.51-$ and $0.99-\mu \mathrm{m}$ diameter polystyrene microspheres and, for the absorbing phantoms, also green molecular dye.

|  | $0.51 \pm 0.01 \mu \mathrm{~m}$ |  |  |  | $0.99 \pm 0.03 \mu \mathrm{~m}$ |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| P | $n\left(\right.$ spheres $/ \mathrm{ml} \times 10^{10}$ ) | $\begin{gathered} \mu_{\mathrm{s}}^{\prime}\left(\mathrm{cm}^{-1}\right) @ \\ 630 \mathrm{~nm} \end{gathered}$ | $\begin{gathered} \mu_{a}\left(\mathrm{~cm}^{-1}\right) @ \\ 630 \mathrm{~nm} \end{gathered}$ | $P$ | $n\left(\right.$ spheres $/ \mathrm{ml} \times 10^{10}$ ) | $\begin{gathered} \mu_{\mathrm{s}}^{\prime}\left(\mathrm{cm}^{-1}\right) @ \\ 630 \mathrm{~nm} \end{gathered}$ | $\begin{gathered} \mu_{\mathrm{a}}\left(\mathrm{~cm}^{-1}\right) @ \\ 630 \mathrm{~nm} \end{gathered}$ |
| 1 | 3.54 | 9.52 | 0 | 7 | 0.486 | 7.61 | 0 |
| 2 | 6.27 | 16.9 | 0 | 8 | 0.852 | 13.3 | 0 |
| 3 | 7.13 | 19.2 | 0 | 9 | 0.975 | 15.3 | 0 |
| 4 | 8.90 | 24.0 | 0 | 10 | 1.21 | 19.0 | 0 |
| 5 | 11.6 | 31.2 | 0 | 11 | 1.58 | 24.7 | 0 |
| 6 | 14.3 | 38.5 | 0 | 12 | 1.95 | 30.6 | 0 |
| 13 | 3.55 | 9.56 | 10.1 | 19 | 0.483 | 7.56 | 10.1 |
| 14 | 6.23 | 16.8 | 9.24 | 20 | 0.851 | 13.3 | 9.24 |
| 15 | 7.13 | 19.2 | 8.95 | 21 | 0.971 | 15.2 | 8.96 |
| 16 | 8.89 | 23.9 | 10.3 | 22 | 1.21 | 19.0 | 10.3 |
| 17 | 11.6 | 31.2 | 9.25 | 23 | 1.57 | 24.6 | 9.29 |
| 18 | 13.3 | 35.8 | 8.59 | 24 | 1.95 | 30.5 | 8.22 |

a diameter of 19.5 mm and a height of 30 mm was internally coated with a black matte paint to reduce reflections from the container walls and eliminate stray light pollution. Additionally, the optical fiber probe was held at a position far away from the beaker walls to satisfy the semi-infinite medium assumption for the turbid phantoms. A broadband halogen light source (AvaLight-Hal LS, Avantes, The Netherlands) was coupled to the source fiber. The collected light from each of the five detector fibers was transmitted to a custom-made multiplexer, which comprised a motorized linear stage that aligned the selected detector fiber with a fiber leading to the spectrometer (AvaSpec-2048TEC-FT, Avantes). The recorded signal from each detector fiber was corrected for the sensor dark current and normalized by the Spectralon white signal (Avantes). The SRR profiles were acquired in the wavelength range from 450 to 800 nm .

### 2.6 Turbid Phantoms

Turbid phantoms with known optical properties are commonly used to objectively evaluate experimental setups and computational methodologies in biomedical optics. ${ }^{37}$ In this study, we prepared 24 water-based turbid phantoms comprising a mixture of absorbers and scatterers. A green molecular dye found in fountain pen inks (Live Line Green) was used as the absorber. The molecular dye was thoroughly tested for its stability through several weeks and proved to be stable even when exposed to direct sunlight. ${ }^{38}$ The scattering component of the dye was negligible due to its molecular nature. A $1-\mathrm{mm}$ cuvette and a cuvette holder (CVH100/M, Thorlabs) were used to measure the absorption coefficient $\mu_{\mathrm{a}}$ of the molecular dye (without the scatterer) by observing the attenuation of a collimated light beam through the cuvette. The absorption coefficient was computed according to the Beer-Lambert law. Polystyrene microspheres
(diameter $0.51 \pm 0.01$ and $0.99 \pm 0.03 \mu \mathrm{~m}$, Polysciences Inc.) were used as the scatterer. The respective $\mu_{\mathrm{s}}$ and phase functions were calculated according to the Mie theory and the narrow size distribution of the microspheres (nearly monodisperse) provided by the manufacturer. The wavelength dependence of the polystyrene refractive index was taken from Nikolov et al. ${ }^{39}$

The prepared turbid phantoms were divided into two groups. The first group of phantoms $\mathrm{P}_{1}$ to $\mathrm{P}_{12}$ comprised only scatterers (phantoms $\mathrm{P}_{1}$ to $\mathrm{P}_{6}$ comprised 0.51 and phantoms $\mathrm{P}_{7}$ to $\mathrm{P}_{12}$ comprised $0.99-\mu \mathrm{m}$ microspheres, Table 2). The second group of phantoms $\mathrm{P}_{13}$ to $\mathrm{P}_{24}$ comprised scatterers and green molecular dye (phantoms $\mathrm{P}_{13}$ to $\mathrm{P}_{18}$ comprised 0.51 and phantoms $\mathrm{P}_{19}$ to $\mathrm{P}_{24}$ comprised $0.99-\mu \mathrm{m}$ microspheres, Table 2). The values of $\mu_{\mathrm{a}}$ for the second group of phantoms ranged between 8.2 and $10.3 \mathrm{~cm}^{-1}$ at 630 nm (absorption peak). The microsphere number density $n$ and corresponding $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$ for all the prepared turbid phantoms are listed in Table 2.

## 3 Results and Discussion

### 3.1 Cost Function Evaluation

The performance and robustness of the proposed CFs were evaluated on the synthetic dataset $\mathcal{R}_{\text {HG-I }}$ by deploying the b -IMC model. The b-IMC model was selected for its dependence on only two parameters ( $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$ ) and, hence, easier visualization of the CF (Fig. 2). The presented example ( $\mathbf{R}$ from the dataset $\mathcal{R}_{\text {HG-I }}$ with the true values of $\mu_{\mathrm{a}}=12.5 \mathrm{~cm}^{-1}$ and $\mu_{\mathrm{s}}^{\prime}=18.9 \mathrm{~cm}^{-1}$ ) clearly shows that the CF significantly affects the estimation of optical properties by the IMC models. The $\mathrm{CF}_{2}$ [Eq. (4)] exhibits improved localization of the minimum with respect to the $\mathrm{CF}_{1}$. Moreover, the relative root-mean-square (RMS) error obtained by the b-IMC model using the $\mathrm{CF}_{1}$ for the entire dataset $\mathcal{R}_{\mathrm{HG}-\mathrm{I}}$ is $2.7 \%$ and $2.8 \%$ for $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$,


Fig. 2 CF dependence on the values of optical properties in terms of absorption $\mu_{\mathrm{a}}$ and reduced scattering $\mu_{\mathrm{s}}^{\prime}$ coefficients for (a) $\mathrm{CF}_{1}$ from Eq. (3) and for (b) $\mathrm{CF}_{2}$ from Eq. (4).
respectively. In contrast, the relative RMS error obtained by the $\mathrm{CF}_{2}$ was one order of magnitude lower, i.e., $0.32 \%$ and $0.33 \%$ for $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$, respectively. For the sake of completeness, the performance of the two CFs was also compared by the e-IMC model. As with the b-IMC model, the relative RMS errors obtained by the e-IMC model based on the $\mathrm{CF}_{1}(5.6 \%$, $4.4 \%$, and $4.7 \%$ for $\mu_{\mathrm{a}}, \mu_{\mathrm{s}}^{\prime}$, and $\gamma$, respectively) were one order of magnitude higher than the errors obtained by the e-IMC model based on the $\mathrm{CF}_{2}(0.60 \%, 0.58 \%$, and $0.76 \%$ for $\mu_{\mathrm{a}}, \mu_{\mathrm{s}}^{\prime}$, and $\gamma$, respectively). The obtained results clearly show the superiority of the $\mathrm{CF}_{2}$ over $\mathrm{CF}_{1}$. Consequently, $\mathrm{CF}_{2}$ was used in all subsequent calculations involving the b-IMC and e-IMC models.

### 3.2 Influence of the Phase Function on the Subdiffusive Reflectance and on the Inverse Monte Carlo Models

Subdiffusive reflectance is known to significantly depend on the phase function. As explained in Sec. 1, the similarity parameter
$\gamma$ has been suggested to account for some of the subdiffusive reflectance variability that arises from the phase functions. In other words, for a constant $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$, the phase functiondependent changes in the subdiffusive reflectance should be reduced when the additional similarity parameter $\gamma$ is kept constant as well.

In order to study the beneficial effect of $\gamma$, we introduced relative variability maps of the subdiffusive reflectance as a function of $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$ at a particular SDS. The relative variability maps were derived as a ratio between the standard deviation and the mean of the reflectance maps across different phase functions. Figure 3 shows the relative variability maps at three SDS of 220,660 , and $1100 \mu \mathrm{~m}$. In the top row (constant $g$ ), the HG, MHG, and GK phase functions were used with $g$ set to 0.8 and $\gamma$ varied from 1.7 to 1.9 . In the bottom row (constant $\gamma$ ), the HG, MHG, and GK phase functions were used with $\gamma$ set to 1.7 and $g$ varied from 0.7 to 0.9 . In comparison to $g, \gamma$ significantly reduces the variability of the reflectance for $\mu_{\mathrm{s}}^{\prime}$ exceeding $\sim 15 \mathrm{~cm}^{-1}$ when different phase functions are


Fig. 3 Relative variability maps of the subdiffusive reflectance at (a) $220 \mu \mathrm{~m}$, (b) $660 \mu \mathrm{~m}$, and (c) $1100 \mu \mathrm{~m}$ SDS for different phase functions, as a function of the absorption $\mu_{\mathrm{a}}$ and reduced scattering $\mu_{\mathrm{s}}^{\prime}$ coefficients. In the top row (constant $g$ ), HG, MHG, and GK phase functions were used with $g=0.8$ and $\gamma$ varied from 1.7 to 1.9. In the bottom row (constant $\gamma$ ), HG, MHG, and GK phase functions were used with $\gamma=1.7$ and $g$ varied from 0.7 to 0.9 .
used. Moreover, although $g$ is frequently described in the literature as an important observable of the angular scattering distribution in turbid media, the subdiffusive reflectance is more dependent on the changes of $\gamma$ than $g$. Similar conclusions were recently made by Calabro and Bigio ${ }^{15}$ and Bodenschatz et al. ${ }^{40}$ The reason for the increased sensitivity of the subdiffusive reflectance to the phase function observed at the low reduced scattering coefficients can be attributed to the longer photon mean free path lengths. In this case, the photons that contribute to the reflectance signal exhibit only a few scattering events, with one of those events likely to be a large-angle deflection. Therefore, reflectance is highly dependent on the largeangle section of the phase function, which is only approximately accounted for by $\gamma$. The effect can be observed in the reflectance variability map in Fig. 3 (bottom row) computed for phase functions that despite having a common $\gamma$ still exhibit significant differences in the large-angle section. To account for this shortcoming, Bodenschatz et al. ${ }^{40}$ have proposed a parameter $\sigma$, which reduced the variability of reflectance for a constant $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$ and different phase functions in the spatial frequency domain. However, in the same study, $\sigma$ was not found to significantly reduce the variability in the spatially resolved domain in comparison to $\gamma$. We believe that the performance of the e-IMC model for $\mu_{\mathrm{s}}^{\prime}$ below $\sim 15 \mathrm{~cm}^{-1}$ could be significantly improved by introducing higher order similarity parameters beyond $\gamma,{ }^{41}$ e.g., $\delta=\left(1-g_{3}\right) /\left(1-g_{1}\right){ }^{42}$ where $g_{3}$ is the third Legendre moment of the phase function. Keeping the values of $\gamma$ and $\delta$ constant should in principle significantly reduce the relative variability of subdiffusive reflectance computed for different phase functions at any given value of $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$.

Another interesting observation can be made at the smallest SDS, where a region of significantly decreased relative reflectance variability occurs. In this region, the reflectance seems unaffected by the choice of the phase function as long as $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$ are kept constant. This observation, named as the "isobestic point," was also made by Calabro and Bigio, ${ }^{15}$ and it is
believed to occur at $\mu_{\mathrm{s}}^{\prime} \mathrm{SDS} \approx 0.7$. In this study, the SDS of $220 \mu \mathrm{~m}$ evaluates to an isobestic point of $32 \mathrm{~cm}^{-1}$, which is consistent with the results in Fig. 3.

To evaluate the effect of the phase function on the estimation of optical properties, the synthetic dataset $\mathcal{R}_{\text {GK-I }}$ was analyzed by the b-IMC and e-IMC models. The $\mathcal{R}_{\text {GK-I }}$ dataset is based on the GK phase function with the value of $g$ set to 0.8 and $\gamma$ varied from 1.65 to 2.25 , which is similar to the conditions used for producing the results in the top row of Fig. 3, where $g$ was constant. Figure 4 (top row) shows the relative errors obtained for the estimated $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$ by the b-IMC model from the dataset $\mathcal{R}_{\text {GK-I }}$. The values of $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$ used in $\mathcal{R}_{\mathrm{GK}-\mathrm{I}}$ are color-coded and can be deduced from the corresponding colorbars. It can be observed that the accuracy of the estimated optical properties decreases significantly due to the variations in $\gamma$. The relative errors are the lowest for $\gamma=1.8$, where the GK phase function simplifies to HG phase function. For all the other values of $\gamma$, the performance is significantly degraded. In contrast, a significant improvement in the accuracy of the optical properties estimated by the e-IMC model from the same dataset $\mathcal{R}_{\text {GK-I }}$ can be observed in Fig. 4 (bottom row). The relative errors of the estimated optical properties obtained by the e-IMC model are mostly within $2 \%$ for all of the selected values of $\gamma$.

The results of b-IMC model presented in Fig. 4 clearly suggest that using only the first similarity relation will lead to large relative errors of the estimated optical properties. For example, although the dataset $\mathcal{R}_{\mathrm{GK}-\mathrm{I}}$ is simulated using a phase function with a constant $g$, the changes in the reflectance due to changes in phase function arising from $\gamma$ cannot be sufficiently accounted for by the first similarity relation. This observation could explain one of the possible sources of errors in studies that utilized b-IMC-like models for optical fiber probes with similar SDS as used in this study. ${ }^{31,43,44}$ We believe that the listed studies could be improved by using additional parameters such as $\gamma$ in addition to $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$ in the IMC models to account for


Fig. 4 Relative errors of (a) the absorption $\mu_{\mathrm{a}}$ and (b) reduced scattering $\mu_{\mathrm{s}}^{\prime}$ coefficients estimated by the b-IMC model (top row) and e-IMC model (bottom row) for the synthetic reflectance dataset $\mathcal{R}_{\text {GK-I }}$. The true values of $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$ used in $\mathcal{R}_{\mathrm{GK}-\mathrm{I}}$ are color coded and can be deduced from the colorbars.
some of the phase function variability that influences the reflectance at small SDS.

### 3.3 Performance of the Basic and Extended Inverse Monte Carlo Models on Synthetic Datasets of Spatially Resolved Reflectance Profiles

To further examine the effect of $\gamma$ on the estimated optical properties, the b-IMC and e-IMC models were extensively evaluated and compared on synthetic datasets of $\mathbf{R}$ (Sec. 2.5, Table 1). The variability of phase functions captured by the synthetic datasets should account for the variability observed in biological tissues. Consequently, similar errors can be expected for measured $\mathbf{R}$ of biological tissues.

Figure 5 shows the values of $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$ estimated by the b-IMC and e-IMC models with respect to the true values used to compute the synthetic datasets. The performance of the e-IMC model is evidently superior to the b-IMC model, since the values obtained by the b-IMC model are significantly more dispersed along the ideal relationship line between the true and estimated parameter values. Moreover, the e-IMC model offers an additional estimate of the similarity parameter $\gamma$ [Fig. 5(e)].

Quantitative results obtained for each synthetic dataset in terms of the two IMC models are summarized in Table 3. As expected, both IMC models performed with relative RMS errors well below $1 \%$ when tested on the datasets $\mathcal{R}_{\text {HG-I }}$ and $\mathcal{R}_{\text {GK-I }}$, respectively. The two datasets were obtained by the same phase functions that were used to create the lookup tables. The performance of the b-IMC model clearly degrades for the dataset $\mathcal{R}_{\mathrm{HG}-\mathrm{II}}$ with $g$ varying between 0.7 and 0.95 . In contrast, the e-IMC model does not seem to be significantly affected by the variation in $g$, which is in agreement with the results in Fig. 3 (bottom row). The e-IMC model performs similarly on the $\mathcal{R}_{\text {GK-II }}$ dataset. However, the RMS errors of the IMC models further increase when applied to the MHG and Mie phase function-based datasets ( $\mathcal{R}_{\mathrm{MHG}}$, pure scattering $\mathrm{P}_{1}$ to $\mathrm{P}_{12}$, and scattering and absorbing $\mathrm{P}_{13}$ to $\mathrm{P}_{24}$ datasets). Once more, the b-IMC model is much more affected by the changes in the phase function and exhibits twice the relative RMS error of the e-IMC model. The quantitative advantage of the e-IMC model over the b-IMC model can be observed in Table 3 that summarizes the results across all the datasets (all datasets).

Despite the improved result, the performance of e-IMC model on the dataset of scattering and absorbing phantoms


Fig. 5 The values of (a) $\mu_{\mathrm{a}}$ and (b) $\mu_{\mathrm{s}}^{\prime}$ estimated by the b -IMC model and the values of (c) $\mu_{\mathrm{a}}$, (d) $\mu_{\mathrm{s}}^{\prime}$, and (e) $\gamma$ estimated by the e-IMC model with respect to the corresponding true values that were used to compute the synthetic datasets of SRR profiles. The ideal relationship is denoted by a black solid line. The synthetic datasets are presented with different colors and markers (see the legend).

Table 3 The RMS errors of the estimated optical properties obtained by the b-IMC and e-IMC models on synthetic datasets of SRR profiles R.

| Synthetic datasets of R | b-IMC |  | e-IMC |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  | RMS error $\mu_{\mathrm{a}}$ | RMS error $\mu_{\text {s }}^{\prime}$ | RMS error $\mu_{\mathrm{a}}$ | RMS error $\mu_{\mathrm{s}}^{\prime}$ | RMS error $\gamma$ |
| $\mathcal{R}_{\text {HG-I }}$ | 0.030 (0.32\%) | 0.11 (0.33\%) | 0.073 (1.0\%) | 0.21 (0.70\%) | 0.013 (0.74\%) |
| $\mathcal{R}_{\text {HG-II }}$ | 0.32 (3.4\%) | 0.68 (2.6\%) | 0.12 (1.2\%) | 0.39 (1.3\%) | 0.029 (1.6\%) |
| $\mathcal{R}_{\text {GK-I }}$ | 0.95 (9.7\%) | 2.3 (6.8\%) | 0.066 (0.60\%) | 0.23 (0.58\%) | 0.015 (0.76\%) |
| $\mathcal{R}_{\text {GK-II }}$ | 1.24 (13.6\%) | 2.5 (10\%) | 0.16 (1.7\%) | 0.51 (1.8\%) | 0.034 (1.6\%) |
| $\mathcal{R}_{\text {MHG }}$ | 0.60 (6.6\%) | 1.4 (6.1\%) | 0.29 (3.0\%) | 1.04 (3.3\%) | 0.062 (3.5\%) |
| Pure Scat. $\mathrm{P}_{1}-\mathrm{P}_{12}$ | 0 (NA) | 0.86 (4.1\%) | 0 (NA) | 0.49 (1.6\%) | 0.071 (3.3\%) |
| Scat. and Abs. $P_{13}-P_{24}$ | 0.91 (42\%) | 1.8 (13\%) | 0.35 (14.8\%) | 0.74 (6.1\%) | 0.10 (4.5\%) |
| All datasets | 0.75 (21\%) | 1.7 (8.7\%) | 0.22 (7.4\%) | 0.59 (3.6\%) | 0.071 (3.3\%) |
| $\underline{\text { All }-\left\{\mathrm{P}_{1}, \mathrm{P}_{7}, \mathrm{P}_{13}, \mathrm{P}_{19}\right\}}$ | 0.68 (16.4\%) | 1.7 (6.5\%) | 0.12 (3.2\%) | 0.56 (2.0\%) | 0.049 (2.3\%) |

$\mathrm{P}_{13}$ to $\mathrm{P}_{24}$ is still significantly degraded. Previous results (bottom row in Fig. 3) point out that $\gamma$ does not fully eliminate the variability of reflectance for low reduced scattering coefficients (especially below $\sim 15 \mathrm{~cm}^{-1}$ ), when different phase functions are used. Since Fig. 3 only includes HG, MHG, and GK phase functions, we also constructed similar relative subdiffusive reflectance variability maps that depend only on the GK and Mie phase functions. Specifically, Fig. 6 shows the relative variability map for three different phase functions with $\gamma$ set to 2.1. The first-phase function was GK with $g=0.8$, which was also used to create the e-IMC lookup table. The second-phase function was Mie for $0.51-\mu \mathrm{m}$ polystyrene spheres at 705.8 nm , which yields $g=0.79$. Finally, the third-phase function was Mie for $0.99-\mu \mathrm{m}$ polystyrene spheres at 499.5 nm , which yields $g=0.93$. Figure 6 highly resembles the bottom row of Fig. 3, thus again suggesting that $\gamma$ cannot fully account for the variability of the phase function for low $\mu_{\mathrm{s}}^{\prime}$. Moreover, since the GK phase function is used in the e-IMC model, an SRR profile simulated by a different phase function with the same value of $\gamma$ could be different due to the phase function properties that
cannot be accounted for by the first two Legendre moments encapsulated in $\gamma$. These differences between the e-IMC model and the SRR profiles then propagate into errors of the estimated $\mu_{\mathrm{a}}, \mu_{\mathrm{s}}^{\prime}$, and $\gamma$. The observation is further confirmed by removing the phantoms $\mathrm{P}_{1}, \mathrm{P}_{7}, \mathrm{P}_{13}, \mathrm{P}_{19}$, that exhibited $\mu_{\mathrm{s}}^{\prime}$ below $15 \mathrm{~cm}^{-1}$ over the majority of the wavelength range, from the summarized RMS errors (All - $\left\{\mathrm{P}_{1}, \mathrm{P}_{7}, \mathrm{P}_{13}, \mathrm{P}_{19}\right\}$; Table 3). As a result, the summarized RMS errors drop significantly and are now $0.12 \mathrm{~cm}^{-1}(3.2 \%), 0.56 \mathrm{~cm}^{-1}(2.0 \%)$, and 0.049 (2.3\%) for $\mu_{\mathrm{a}}, \mu_{\mathrm{s}}^{\prime}$, and $\gamma$, respectively.

### 3.4 Performance of the Extended Inverse Monte Carlo Model on Measured Spatially Resolved Reflectance Profiles

Finally, we evaluated the e-IMC model on measured $\mathbf{R}$ acquired from turbid phantoms (Sec. 2.6). To compare the measured reflectance to the MC simulated reflectance at a particular SDS, a calibration procedure was required. Briefly, the MC simulated reflectance is normalized against the initial number


Fig. 6 The relative variability maps of the subdiffusive reflectance at (a) $220 \mu \mathrm{~m}$, (b) $660 \mu \mathrm{~m}$, and (c) $1100 \mu \mathrm{~m}$ SDS for different phase functions at a constant $\gamma=2.1$. The employed phase functions included the GK phase function with $g$ set to 0.8 (the value employed by the e-IMC lookup table) and Mie phase functions for 0.51 - and $0.99-\mu \mathrm{m}$ polystyrene microspheres at 705.8 and 499.5 nm , respectively.


Fig. 7 The values of (a) $\mu_{\mathrm{a}}$, (b) $\mu_{\mathrm{s}}^{\prime}$, and (c) $\gamma$ estimated by the e-IMC model from the measured $\mathbf{R}$ of turbid phantoms with respect to the corresponding true values. The ideal relationship is denoted by a black solid line. Different phantoms are presented with different colors (see the legend).
of launched photon packets, while the measured reflectance is normalized against a reflectance standard. Consequently, to account for the reflectance properties of the standard, the reflectance of the turbid phantoms with known optical properties is measured and simulated. The calibration factor is introduced as the ratio between the MC simulated and measured reflectance. Ideally, the calibration factor for a given wavelength and SDS should remain the same across all the turbid phantoms. In this study, the purely scattering phantoms $P_{1}$ to $P_{12}$ were used for computing the calibration factors. The variations of the calibration factors at all spectral bands and SDS were within $2 \%$, and mostly arising from the variations in the measurements.

The performance of the e-IMC model was evaluated on the remaining turbid phantoms $\mathrm{P}_{13}$ to $\mathrm{P}_{24}$ with nonzero $\mu_{\mathrm{a}}$. The estimated values of the optical properties with respect to the corresponding true values are shown in Fig. 7 and are similar to those in Figs. 5(c)-5(e). Moreover, the RMS errors of $\mu_{\mathrm{a}}, \mu_{\mathrm{s}}^{\prime}$, and $\gamma$ obtained for the turbid phantoms are $0.38 \mathrm{~cm}^{-1}(15.6 \%)$, $0.71 \mathrm{~cm}^{-1}(5.8 \%)$, and $0.10(4.8 \%)$, respectively, and tightly follow the results obtained for the corresponding synthetic datasets (Table 4). In addition, by removing the turbid phantoms $\mathrm{P}_{13}$ and $\mathrm{P}_{19}$ ( $\mu_{\mathrm{s}}^{\prime}$ below $15 \mathrm{~cm}^{-1}$ ) from the summarized RMS
errors, nearly a twofold improvement in terms of relative RMS error is attained. Consequently, the RMS errors stay within the $10 \%$ margin, however, only for a subregion of optical properties where $\mu_{\mathrm{s}}^{\prime}$ exceeds $\sim 15 \mathrm{~cm}^{-1}$. The obtained results in this section are consistent with the results shown in Figs. 3 and 6, where $\gamma$ accounts for the phase functions variability only for $\mu_{\mathrm{s}}^{\prime}$ values exceeding $\sim 15 \mathrm{~cm}^{-1}$.

The related studies from Hennessy et al. ${ }^{31}$ and Rajaram et al. ${ }^{43}$ estimated $\mu_{\mathrm{a}}$ and $\mu_{\mathrm{s}}^{\prime}$ with relative RMS errors of $0.74 \%$ and $1.74 \%$, and $11.6 \%$ and $5.9 \%$, respectively. Both studies used optical fiber probes with small SDS between 250 and $300 \mu \mathrm{~m}$. According to these results, the proposed e-IMC model gives slightly higher relative errors for the turbid phantoms $P_{13}$ to $P_{24}$. The results by Hennessy et al. are especially surprising since the employed IMC model was based on the HG phase function at a constant $g=0.85$ (similar to our b-IMC model), while the validation was performed on turbid phantoms containing polystyrene microspheres that follow the Mie phase function. We have shown that the reflectance at small SDS and constant $g$ can vary significantly, if different phase functions are used (Fig. 3). Moreover, we have observed a significant degradation of the b-IMC model on synthetic datasets (Table 3).

Table 4 The RMS errors of the estimated optical properties obtained by the e-IMC models on synthetic (left) and measured (right) SRR profiles of turbid phantoms.

|  | e-IMC, synthetic |  |  | e-IMC, measured |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Phantoms | RMS error $\mu_{\mathrm{a}}$ | RMS error $\mu_{\mathrm{s}}^{\prime}$ | RMS error $\gamma$ | RMS error $\mu_{\mathrm{a}}$ | RMS error $\mu_{\mathrm{s}}^{\prime}$ | RMS error $\gamma$ |
| Scat. and Abs. | 0.35 (14.8\%) | 0.74 (6.1\%) | 0.10 (4.5\%) | 0.38 (15.6\%) | 0.71 (5.8\%) | 0.10 (4.8\%) |
| $\underline{\text { Scat. and Abs. - }\left\{\mathrm{P}_{13}, \mathrm{P}_{19}\right\}}$ | 0.14 (6.3\%) | 0.63 (2.7\%) | 0.066 (3.0\%) | 0.21 (8.4\%) | 0.59 (2.5\%) | 0.072 (3.3\%) |

One of the possible causes for a better performance of the IMC models in the related studies is the use of spectral constrains in terms of tissue chromophore concentrations and spectral dependence of the $\mu_{\mathrm{s}}^{\prime}$. Spectral constrains were required because only one SDS was utilized, thus the full reflectance spectrum was used to estimate only a few parameters. This, however, significantly reduced the number of optimized parameters in comparison to the number of available measurements. It is important to note that unlike the IMC models presented in the related studies, the e-IMC model does not require spectral constrains. In this way, the e-IMC model requires no prior knowledge of the chromophores and scatterers in the turbid medium to estimate $\mu_{\mathrm{a}}, \mu_{\mathrm{s}}^{\prime}$, and $\gamma$. As such, the e-IMC can be utilized only at particular wavelengths. In addition, spectral constrains can be easily introduced into the e-IMC model. We believe that this should in principle further reduce the relative error of the estimated quantities since there would be significantly more measurements and less parameters to optimize. For human tissues, this would most commonly include modeling $\mu_{\mathrm{a}}$ by the absorption spectra of chromophores, such as oxygenated and deoxygenated hemoglobin, melanin, carotenoids, while $\mu_{\mathrm{s}}^{\prime}$ can be modeled as $\mu_{\mathrm{s}}^{\prime}(\lambda)=A\left(\lambda / \lambda_{0}\right)^{-B} .{ }^{32}$ The spectral dependence of $\gamma$ in tissue is close to constant across the visible spectrum and could thus be adequately modeled by a linear or quadratic function. ${ }^{16,45}$

Finally, the turbid phantoms used in this study to experimentally test the e-IMC model were based on polystyrene microspheres, which follow the Mie phase function. Polystyrene microspheres are very practical to use since they are available in standardized nearly monodisperse suspensions. In this way, the scattering coefficient and the phase function can easily be calculated. However, monodisperse solutions do not exactly mimic the tissue optical properties in terms of the shape of the phase function. As a result, the accuracy of the estimated optical properties for tissues can somewhat differ from the accuracy obtained for such turbid phantoms. Since biological tissues exhibit phase functions similar to GK or MHG, more accurate results can be expected by the e-IMC model (similar to the synthetic dataset $\mathcal{R}_{\text {GK-II }}$ or $\mathcal{R}_{\mathrm{MHG}}$ in Table 3).

## 4 Conclusion

This study offers a simple yet effective approach for estimation of optical properties in the subdiffusive regime where the reflectance significantly depends on the phase function. The subdiffusive reflectance variability due to the phase function can be reduced by taking into account an additional similarity parameter $\gamma$, which carries additional information about the turbid medium phase function. Although $\gamma$ did not guarantee reduced reflectance variability for reduced scattering coefficients under $\sim 15 \mathrm{~cm}^{-1}$, it has proved beneficial when used in the IMC model. In comparison to the b-IMC model that depends only on the absorption and reduced scattering coefficients, the e-IMC model extended by $\gamma$ showed increased accuracy when used on synthetic datasets and measured SRR profiles of turbid phantoms. For a subset of optical properties where the reduced scattering coefficients exceeded $\sim 15 \mathrm{~cm}^{-1}$, the relative RMS errors of the estimated absorption and reduced scattering coefficients, and the similarity parameter $\gamma$ for measured SSR profiles were $8.4 \%, 2.5 \%$, and $3.3 \%$, respectively.

The main advantage of the e-IMC model is that in conjunction with the proposed CF , the absorption and reduced scattering coefficients and $\gamma$ can be estimated from SRR profiles acquired at only five SDS. This can reduce the acquisition time and, due
to the fast estimation of optical properties by the lookup table approach, potentially offer a faster and a more detailed insight into human tissues in a clinical setting. Moreover, the e-IMC model, unlike many other proposed IMC models, offers estimation of optical properties without any prior knowledge of the tissue chromophores or the spectral dependence of the reduced scattering coefficient or $\gamma$. Consequently, the e-IMC model can be used for analysis of turbid media that have not yet been extensively studied. While the e-IMC model presented in this study is intended for semi-infinite media, multilayered models can be supported by extending the lookup tables. Likewise, the e-IMC model can be extended to estimate similarity parameters beyond $\gamma$, which could further improve the accuracy of the inverse model, or any of the additional parameters of the turbid medium or biological tissue, provided that the correlations among the free parameters are small. The main concern with the e-IMC model is its limited use for reduced scattering coefficients below $\sim 15 \mathrm{~cm}^{-1}$. Since many of the studies to date have used $\gamma$, we believe they might have suffered from similar limitations. By considering higher order similarity parameters in addition to $\gamma$, this limitation might be overcome. With this study, we attempted to point out the advantages and the limitations of the e-IMC model for estimation of optical properties by SRR spectroscopy in the subdiffusive regime. The fact that $\gamma$ has a limited use requires further investigation for a new or additional parameters that would better encapsulate the phase function information.

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## References

1. R. Marchesini et al., "In vivo spectrophotometric evaluation of neoplastic and non-neoplastic skin pigmented lesions-I. Reflectance measurements," Photochem. Photobiol. 53(1), 77-84 (1991).
2. G. Lu and B. Fei, "Medical hyperspectral imaging: a review," J. Biomed. Opt. 19(1), 010901 (2014).
3. S.-P. Lin et al., "Measurement of tissue optical properties by the use of oblique-incidence optical fiber reflectometry," Appl. Opt. 36(1), 136-143 (1997).
4. U. Utzinger and R. R. Richards-Kortum, "Fiber optic probes for biomedical optical spectroscopy," J. Biomed. Opt. 8(1), 121-147 (2003).
5. R. Reif, O. A'Amar, and I. J. Bigio, "Analytical model of light reflectance for extraction of the optical properties in small volumes of turbid media," Appl. Opt. 46(29), 7317-7328 (2007).
6. T. J. Farrell, B. C. Wilson, and M. S. Patterson, "The use of a neural network to determine tissue optical properties from spatially resolved diffuse reflectance measurements," Phys. Med. Biol. 37(12), 2281-2286 (1992).
7. A. Kienle et al., "Spatially resolved absolute diffuse reflectance measurements for noninvasive determination of the optical scattering and absorption coefficients of biological tissue," Appl. Opt. 35(13), 2304-2314 (1996).
8. R. M. P. Doornbos et al., "The determination of in vivo human tissue optical properties and absolute chromophore concentrations using spatially resolved steady-state diffuse reflectance spectroscopy," Phys. Med. Biol. 44(4), 967-981 (1999).
9. T. J. Pfefer et al., "Reflectance-based determination of optical properties in highly attenuating tissue," J. Biomed. Opt. 8(2), 206-215 (2003).
10. D. Arifler et al., "Spatially resolved reflectance spectroscopy for diagnosis of cervical precancer: Monte Carlo modeling and comparison to clinical measurements," J. Biomed. Opt. 11(6), 064027 (2006).
11. K.-B. Sung et al., "Accurate extraction of optical properties and top layer thickness of two-layered mucosal tissue phantoms from spatially resolved reflectance spectra," J. Biomed. Opt. 19(7), 077002 (2014).
12. F. Bevilacqua and C. Depeursinge, "Monte Carlo study of diffuse reflec tance at source-detector separations close to one transport mean free path," J. Opt. Soc. Am. A 16(12), 2935-2945 (1999).
13. J. R. Mourant et al., "Influence of the scattering phase function on light transport measurements in turbid media performed with small sourcedetector separations," Opt. Lett. 21(7), 546-548 (1996)
14. S. C. Kanick et al., "Scattering phase function spectrum makes reflectance spectrum measured from intralipid phantoms and tissue sensitive to the device detection geometry," Biomed. Opt. Express 3(5), 10861100 (2012).
15. K. W. Calabro and I. J. Bigio, "Influence of the phase function in generalized diffuse reflectance models: review of current formalisms and novel observations," J. Biomed. Opt. 19(7), 075005 (2014)
16. A. J. Radosevich et al., "Subdiffusion reflectance spectroscopy to measure tissue ultrastructure and microvasculature: model and inverse algorithm," J. Biomed. Opt. 20(9), 097002 (2015).
17. U. A. Gamm et al., "Quantification of the reduced scattering coefficient and phase-function-dependent parameter $\gamma$ of turbid media using multidiameter single fiber reflectance spectroscopy: experimental validation,' Opt. Lett. 37(11), 1838-1840 (2012).
18. S. C. Kanick et al., "Sub-diffusive scattering parameter maps recovered using wide-field high-frequency structured light imaging," Biomed. Opt Express 5(10), 3376 (2014).
19. P. Krauter et al., "Optical phantoms with adjustable subdiffusive scattering parameters," J. Biomed. Opt. 20(10), 105008 (2015).
20. A. Liemert and A. Kienle, "Exact and efficient solution of the radiative transport equation for the semi-infinite medium," Sci. Rep. 3 (2013).
21. L. Wang, S. L. Jacques, and L. Zheng, "MCML-Monte Carlo modeling of light transport in multi-layered tissues," Comput. Methods Programs Biomed. 47(2), 131-146 (1995).
22. F. Bevilacqua et al., "In vivo local determination of tissue optical properties: applications to human brain," Appl. Opt. 38(22), 4939-4950 (1999).
23. E. Alerstam, T. Svensson, and S. Andersson-Engels, "Parallel computing with graphics processing units for high-speed Monte Carlo simulation of photon migration," J. Biomed. Opt. 13(6), 060504 (2008).
24. P. Naglič et al., "Limitations of the commonly used simplified laterally uniform optical fiber probe-tissue interface in Monte Carlo simulations of diffuse reflectance," Biomed. Opt. Express 6(10), 3973 (2015)
25. K. W. Calabro and W. Cassarly, "Modeling scattering in turbid media using the Gegenbauer phase function," Proc. SPIE 9333, 93330F (2015)
26. L. O. Reynolds and N. J. McCormick, "Approximate two-parameter phase function for light scattering," J. Opt. Soc. Am. 70(10), 1206-1212 (1980).
27. C. F. Bohren and D. R. Huffman, Absorption and Scattering of Light by Small Particles, John Wiley and Sons, New York (1983).
28. A. N. Yaroslavsky et al., "Influence of the scattering phase function approximation on the optical properties of blood determined from the integrating sphere measurements," J. Biomed. Opt. 4(1), 47-53 (1999)
29. D. Toublanc, "Henyey-Greenstein and Mie phase functions in Monte Carlo radiative transfer computations," Appl. Opt. 35(18), 3270-3274 (1996).
30. G. M. Palmer and N. Ramanujam, "Monte Carlo-based inverse model for calculating tissue optical properties. Part I. Theory and validation on synthetic phantoms," Appl. Opt. 45(5), 1062-1071 (2006).
31. R. Hennessy et al., "Monte Carlo lookup table-based inverse model for extracting optical properties from tissue-simulating phantoms using diffuse reflectance spectroscopy," J. Biomed. Opt. 18(3), 037003 (2013).
32. S. L. Jacques, "Optical properties of biological tissues: a review," Phys. Med. Biol. 58(11), R37-R61 (2013).
33. R. Graaff et al., "Condensed Monte Carlo simulations for the descrip tion of light transport," Appl. Opt. 32(4), 426-434 (1993).
34. R. Graaff et al., "Similarity relations for anisotropic scattering in absorbing media," Opt. Eng. 32(2), 244-252 (1993).
35. A. Kienle and M. S. Patterson, "Determination of the optical properties of turbid media from a single Monte Carlo simulation," Phys. Med. Biol. 41(10), 2221-2227 (1996).
36. T. Coleman and Y. Li, "An interior trust region approach for nonlinear minimization subject to bounds," SIAM J. Optim. 6(2), 418-445 (1996).
37. B. W. Pogue and M. S. Patterson, "Review of tissue simulating phantoms for optical spectroscopy, imaging and dosimetry," J. Biomed. Opt. 11(4), 041102 (2006)
38. B. Cugmas and P. Nagli, "Durability of ink absorbers in water-based phantoms used in diffuse reflectance spectroscopy," Elektrotehniški Vestnik 83(3), 93-98 (2016).
39. I. D. Nikolov and C. D. Ivanov, "Optical plastic refractive measurements in the visible and the near-infrared regions," Appl. Opt. 39(13), 20672070 (2000)
40. N. Bodenschatz et al., "Quantifying phase function influence in subdiffusively backscattered light," J. Biomed. Opt. 21(3), 035002 (2016).
41. D. R. Wyman, M. S. Patterson, and B. C. Wilson, "Similarity relations for anisotropic scattering in Monte Carlo simulations of deeply penetrating neutral particles," J. Comput. Phys. 81(1), 137-150 (1989).
42. H. Tian, Y. Liu, and L. Wang, "Influence of the third-order parameter on diffuse reflectance at small source-detector separations," Opt. Lett. 31(7), 933 (2006)
43. N. Rajaram, T. H. Nguyen, and J. W. Tunnell, "Lookup table-based inverse model for determining optical properties of turbid media," J. Biomed. Opt. 13(5), 050501 (2008).
44. X. Zhong, X. Wen, and D. Zhu, "Lookup-table-based inverse model for human skin reflectance spectroscopy: two-layered Monte Carlo simulations and experiments," Opt. Express 22(2), 1852-1864 (2014).
45. P. Thueler et al., "In vivo endoscopic tissue diagnostics based on spectroscopic absorption, scattering, and phase function properties," J. Biomed. Opt. 8(3), 495-503 (2003).

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