Perspective on optical imaging for functional assessment in musculoskeletal extremity trauma surgery

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Abstract

Significance: Extremity injury represents the leading cause of trauma hospitalizations among adults under the age of 65 years, and long-term impairments are often substantial. Restoring function depends, in large part, on bone and soft tissue healing. Thus, decisions around treatment strategy are based on assessment of the healing potential of injured bone and/or soft tissue. However, at the present, this assessment is based on subjective clinical clues and/or cadaveric studies without any objective measure. Optical imaging is an ideal method to solve several of these issues.

Aim: The aim is to highlight the current challenges in assessing bone and tissue perfusion/viability and the potentially high impact applications for optical imaging in orthopaedic surgery.

Approach: The perspective will review the current challenges faced by the orthopaedic surgeon and briefly discuss optical imaging tools that have been published. With this in mind, it will suggest key research areas that could be evolved to help make surgical assessments more objective and quantitative.

Results: Orthopaedic surgical procedures should benefit from incorporation of methods to measure functional blood perfusion or tissue metabolism. The types of measurements though can vary in the depth of tissue sampled, with some being quite superficial and others sensing several millimeters into the tissue. Most of these intrasurgical imaging tools represent an ideal way to improve surgical treatment of orthopaedic injuries due to their inherent point-of-care use and their compatibility with real-time management.

Conclusion: While there are several optical measurements to directly measure bone function, the choice of tools can determine also the signal strength and depth of sampling. For orthopaedic surgery, real-time data regarding bone and tissue perfusion should lead to more effective patient-specific management of common orthopaedic conditions, requiring deeper penetrance commonly seen with indocyanine green imaging. This will lower morbidity and result in decreased variability associated with how these conditions are managed.

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1 Introduction

A reliable assessment of tissue viability is critical to effectively treating patients who sustain traumatic extremity injury. Despite their relatively inert appearance, bone, tendons, and ligaments are, physiologically speaking, highly active. Bones, specifically, receive ~10% of the entire cardiac output of blood volume.¹ Dysvascular tissues have limited potential to regenerate, heal, or fight infection due to insufficient delivery of inflammatory cells, growth factors, osteoprogenitor cells, endogenous immune cells, and antibiotics. It is clear that, in orthopaedic injury, nonviable or poorly perfused bone and soft tissue inhibits healing and increases the risk for infection. Because of this, assessing bone and soft tissue viability is critical to making effective treatment decisions, and the comparative translucency of these tissues to red and near-infrared light makes optical methods feasible.

2 Current Paradigm and Challenges

To date, the eyes and hands of the surgeon remain the dominant “imaging modality” used to make decisions regarding the health of soft tissue or bone. Methods currently used to assess tissue and bone perfusion are subjective and entail non-quantitative clinical cues. These techniques include clinical judgement based on the color and swelling of skin, presence of skin wrinkles, color, and turgor of deep tissues, measurements of prior cadaveric studies (for example, regarding perfusion of the meniscus in the knee),^{2,3} extent of soft tissue stripping off bone, and the “paprika sign” (defined as scattered pinpoint bleeding on bone).^{4,5} The subjective or imprecise nature of these assessments leads to substantial variation in treatment and a lack of advancement in objectively driven treatment protocols.

Imaging blood perfusion to soft tissue and bone can be done using traditional imaging modalities, such as positron emission tomography/computed tomography (PET/CT)^{6,7} and contrast-enhanced magnetic resonance imaging (MRI).⁸ These imaging modalities provide detailed and accurate information of the fractured or diseased bony and soft tissue anatomical structure, yet they have issues with limits to spatial resolution and artifacts from metallic implants. The logistical limitations are the cost recovery, the need to schedule them as a separate procedure, and the limits to using them for routine real-time assessment or repeated re-examination.^{6,7} MR-based techniques are limited in orthopaedic patients due to metal artifact and signal dropout in association with metal implants. A limited number of studies have used PET/CT or MRI to measure blood flow to bone for surgical indications, including infection, osteomyelitis, and nonunion.^{8–13} However, there is limited to no access to these imaging methods in real-time in the operating room. At the present time, fluoroscopy and plain radiographs are used regularly in the operating room to visualize bones and assess fracture alignment and/or hardware placement. In contrast to these radiologic modalities, optical imaging can obtain metabolic information that provides physiological abnormalities of the tissue on microvascular perfusion and/or bone and tissue viability. The main limitation of optical imaging is the inability to achieve both high spatial resolution and high tissue penetrance: any added value, then, is either in high-resolution imaging at low penetrance or more macroscopic assessment of tissue function at a more moderate to medium penetrance. Thus, optical imaging can be a unique complementary imaging modality to the conventional imaging modalities.

3 Potential Optical Imaging and Sensing Needs

3.1 Surgical Timing/Skin Assessment

High energy fractures are frequently associated with significant insult to the soft tissues surrounding bone. Historically, immediately performing open reduction and internal fixation of injuries such as tibial plafond, tibial plateau, and calcaneus fractures has been associated with unacceptably high rates of infections and wound complications.^{14–17} Based on these experiences, staged protocols in which fractures were temporized in traction, splints, or external fixators until the soft tissue envelope had recovered enough to undergo open reduction internal fixation were introduced.^{18–21} Unfortunately, no objective definitive clinical signs exist to determine the timing for definitive fixation.²² Surgeons frequently use clinical clues such as the presence of skin wrinkles or epithelialization of fracture blisters, but there are no evidence-based objective signs or thresholds to indicate that the soft tissues are appropriate for a surgical incision and open fracture fixation. Staged protocols have resulted in decreased wound complications; however, they have recently been challenged with a resurgence of early fixation for a variety of reasons. For example, there are valid concerns regarding quality of reduction (which becomes increasingly difficult the longer the delay to internal fixation), increased operative time, increased healthcare costs, and pin site infections associated with the use of external fixators, and these have led a number of surgeons to question whether acute definitive internal fixation might be more appropriate in certain patients. While this view is supported by multiple recent studies showing that fractures treated within 72 h of surgery have comparable outcomes compared with staged fixation,^{23–25} these studies were all retrospective reviews, and the criterion for early fixation was surgeon or facility dependent. It is clear that not all soft tissues are appropriate for acute fixation. Therefore, an objective tool that could effectively measure the soft tissue recovery with thresholds identified in association with appropriateness for definitive internal fixation may significantly improve the current practice. Optical imaging tools that image tissue perfusion of indocyanine green (ICG) have longstanding success regarding assessment of soft tissue viability and perfusion in the setting of plastic surgical indications such as free osseous flap perfusion^{26,27} or breast reconstruction.^{28,29} They are, therefore, well suited for this application.

3.2 Assessing the Viability of Deep Tissue in Traumatic Wounds

High-energy open fractures, with bone and deep soft tissue exposed to environmental contamination, can be at an extraordinarily high risk of a complication, such as infection and nonunion, occurring in 10% to 60% of patients.^{30–36} Complications convert a difficult 6-month recovery into a several-year (or longer) recovery that may include unplanned surgical procedures, prolonged morbidity, loss of function, and loss of limb. Bone and soft tissue devitalization is believed to be a critical determinant in the development of complication following traumatic injury. Deficient perfusion or blood flow prevents delivery of inflammatory cells, growth factors, osteoprogenitor cells, endogenous immune cells, and systemically delivered antibiotics,^{37–43} which limit the potential of the injured extremity to regenerate, heal, or fight infection. Because of this, the cornerstone of management of these injuries includes surgical debridement of all devitalized bone and soft tissue. However, the subjective clinical signs (such as color, turgor, and soft tissue stripping) remain the only signs surgeons can currently use to evaluate bone or soft tissue perfusion or viability. These highly subjective signs lead to substantial variation in the extent of debridement and a high potential for either under- or overdebridement, both of which have potentially catastrophic downstream consequences. What is needed is an intraoperative system that can assess vascular perfusion of bone in the surgical field in real-time, which could guide surgeons in the right amount of bone and soft tissue to debride. Although there are challenges in association with the three-dimensional (3-D) nature of traumatic wounds and open fractures, optical imaging techniques^{44,45} are uniquely suited to collect data on the inflow/outflow kinetics of complex tissues.

3.3 Assessing Perfusion of Deep Tissue in Infection

In the setting of bone infection, or osteomyelitis, it is clear that there are two interconnected problems with regards to effective treatment: (1) location and debridement of all devitalized or poorly perfused bone and (2) location and debridement of all infected bone. In the current paradigm, MRI may be used to assess the extent of bony edema on T2-weighted sequences and marrow replacement on T1-weighted sequences when there is not pre-existing hardware in place. However, more commonly, metallic hardware prevents MRI from providing useful information due to signal dropout around metallic implants. Furthermore, there is not a one-to-one relationship between either vascular perfusion or microbial infiltration in bone and bony edema on T2-weighted sequences or marrow replacement on T1-weighted sequences. The standard of care is otherwise removal of hardware, surgical debridement, and, in the setting of unhealed fracture, staged fracture stabilization using either antibiotic coated or non-antibiotic coated implants. Surgical debridement is carried out using the previously described signs including the color of bone/tissues, turgor, and “paprika sign” or using a burr to assess for bony bleeding. The failure rate of treatment of these issues is unacceptably high, resulting in reoperation and/or amputation 30% of the time in the setting of fracture or fusion.^{46–48} Because, to date, there are no methods available to measure bone perfusion or bacterial penetrance *in vivo*, there is very little understanding of which of these variables is necessary to guide debridement. Further, there are no intraoperative tools to assess how effective performance is in these debridement procedures. Similar to issues associated with evaluating vascular perfusion of open fractures, the 3-D nature of infected wound beds presents challenges. However, optical imaging systems are uniquely suited to assess perfusion of bone and deep soft tissues in operating room settings in real-time.^{27,44,45,49}

3.4 Assessing Soft Tissue for Healing Potential

All injured tissue requires adequate perfusion to deliver growth factors necessary for healing. This is true for skin and bone (described above) as well as soft tissue injuries, such as ligament ruptures, meniscal tears, and cartilage injury. This is the rationale for the high failure rate of primary repair of the anterior cruciate ligament (ACL). This unacceptable failure rate led to the abandonment of primary repair in favor of ACL reconstruction using autograft or allograft such as patellar tendon or hamstring.^{50–56} However, ACL reconstruction is not without issues, and some continue to advocate for primary repair to preserve the native ligament, which maintains proprioception^{57,58} and prevents complications in graft harvesting, tunnel widening, and revisions.^{59,60} These issues with reconstruction have led to methods to support a primary repair, including the design of an extracellular matrix scaffold activated with the patients’ blood as an alternate way to deliver growth factors and promote tissue healing.⁶¹ Several authors have noted that the location of the tear may play a role in outcomes following primary repair, with primary repair of more proximal tears performing better than mid-substance or distally based ACL tears.^{62–67} Likely, this differential rate of healing of a primary ligament repair between proximally located and other tears is associated with the vascular perfusion of the ligament. Given this, if there was a way to assess vascular perfusion and therefore healing potential arthroscopically, it may be possible to preserve the native ligament in appropriate patients and more effectively manage this increasingly common injury.

Similarly, the standard treatment for a meniscal tear is based on cadaveric studies evaluating the vascular penetrance or vascular perfusion of the meniscus.^{2,3,68} From these cadaveric studies, the meniscus was divided according to vascularity into red–red (well perfused), red–white (marginally perfused), or white–white (poorly perfused) zones.^{2,3,69,70} This classification, based on perfusion assessed in cadaveric menisci, is used to indicate whether there is adequate perfusion to allow the meniscus to heal after repair versus location where perfusion is not adequate and whether the area with the meniscal tear should be debrided or removed surgically. However, it is clear that meniscus preservation is the first choice when there is adequate perfusion to prevent the risk of secondary osteoarthritis.^{71–74} Furthermore, several papers have noted that there is variability in the extent of meniscal perfusion in association with age and other patient-specific variables and that the zones described above are not universally accurate.^{75–80}

In both of these injury types, as well as other soft tissue orthopaedic injuries, the translation of optical imaging techniques to an arthroscope could inform appropriate surgical management (repair versus reconstruction or debridement) in a patient-specific manner rather than basing this decision on vascular perfusion of the average cadaveric specimen. This advancement could simultaneously expand the number of patients eligible for repair, reduce failure rates, and provide both patients and surgeons with reassurance that the correct surgical procedure was performed.

4 Optical Tools Utilized to Date

When optical imaging is used for maximum benefit, light of different wavelengths or different interaction mechanisms can be used to capture biophysical changes in the tissue, occurring in the vascular as well as extravascular and cellular matrix compartments.^{81–83} The key to most of the applications, as with all imaging systems, is to have a good biophysical understanding of the compartmentalization of the endogenous or exogenous chromophores that contribute to each image so that the images can be interpreted appropriately. When compared with conventional medical imaging modalities, such as MRI, PET, or x-ray CTCT, optical imaging detects changes in light reflectance, absorption, and scattering of the tissue and thus offers advantages of non-ionizing, low-cost, and portable imaging for routine, low-cost, longitudinal monitoring of response to therapy or during human surgery.^{81–83} The logistics of use of optical imaging, being real time and at the point of care, make it more comparable to C-arm x-ray or ultrasound. Therefore, any determination of the value of optical imaging in tissue injury care needs to be done within the context of what information can be assessed at the point of surgery decision making.

The challenge of applying optical tools to bone and soft tissue assessment has been the high scattering nature of both tissues, limiting the spatial depth dependence of the information. Clearly surface camera-based imaging tools interrogate a range of depths in the millimeter to centimeter range,⁴⁵ with rapidly decreasing sensitivity and spatial resolution with small increases in depth. So, the two major attractions to optical tools come in the categories of one of three rough regimes:

- (1) microscopic imaging and sensing (micron sampling depth and spatial resolution),
- (2) surface imaging during surgery (micron-mm sampling depth and spatial resolution),
- (3) deep tissue sensing during surgery (mm-cm sampling depth and spatial resolution).

These regimes are defined by the type of tool used and dictate what information can be obtained during imaging. The matching of the capabilities of each tool with the needs in each application are essential to evaluate.

Table 1 presents an overview of the tools that have been tried in soft and hard tissue trauma and disease studies. The engineering aspect of the contrast mechanism is used to categorize the tools in column 1. However, in terms of utility and function for trauma surgery, it is likely much more functional to consider the depth of interaction, column 3, because this is the factor that will determine what biomedical information the tool can supply. For example, optical coherence tomography (OCT) has been widely used^{91,92} in bone imaging but with penetrance on the order of <1 mm, thereby allowing its use only for largely research studies and unlikely as a major surgical tool. Similarly, ultraviolet (UV) fluorescence imaging provides information bone necrosis, and the addition of antibiotics has been shown to enhance the ability of this fluorescence to provide diagnostic value about osteonecrosis. However, the penetrance of both UV and antibiotic fluorescence to just tens to hundreds of microns (μm) does limit their use to situations in which the imaging is directly on the bone surface in question.

In terms of molecular information about bone, perhaps no other measure has the potential to provide high feature information than Raman spectroscopy.^{99–105} The measures of bone regrowth have been well studied, and measurements through even centimeter thick tissues have been demonstrated with spatially offset Raman or Raman near-infrared (NIR) tomography.^{106,109} However, practical issues in the very low signal strength and the specificity of the signal have limited the routine application of this technique to research studies to date. More exotic methods such as

Table 1 Listing of optical imaging tools utilized in soft and hard tissue trauma and disease studies, categorized by the type of contrast, depth regime, and a brief strengths and weaknesses list.

Contrast mechanism	Probe technology	Depth regime	Strengths/weaknesses	References
Endogenous signals	UV fluorescence	μm	No exogenous contrast needed; assessment of bone necrosis; superficial tissues only.	84, 85
	Scatter, dichroic, polarization, OCT	μm to 1 mm	No exogenous contrast needed; high spatial resolution; matrix structures and flow; superficial tissues only.	86–92
	NIRS (absorption and scattering)	Several mm to cm	No exogenous contrast needed; functional information of hemodynamics and tissue oxygenation; need to scan probe over region.	93–96
	NIR imaging (reflection)	Several mm	No exogenous contrast needed; 3-D-model-based surface-shape tracking.	97, 98
	Raman, NIR-hyperspectral and infrared (IR)	μm to cm	High molecular specificity; very low signal levels; specificity/sensitivity tradeoff.	99–109
	Terahertz (THz)	μm	Superficial sensing; largely water content based.	110, 111
Contrast agents	Antibiotics	μm	Sensing of osteonecrosis or bone growth; well-tolerated; multiday use for contrast.	112–116
	Fluorescence recovery after photobleaching	μm to mm	Inter cellular solute flow sensing; used with standard contrast agents; superficial imaging assessment.	117–120
	ICG	Several mm to cm	Vascular/tissue perfusion; often used qualitatively but can be provide a binary diagnostic; can provide flow kinetics.	27, 121, 122
	Molecular contrast and nanoparticles	μm to cm	Matrix and biology targeting; limited to largely preclinical to date, although evolving rapidly.	123–126
Hybrid imaging modalities	Optical/ICG	Several mm to cm	Commercially available and growing in usage; primarily used for soft tissue due to higher perfusion.	27, 121, 122
	Photoacoustic/optoacoustic	Several mm to cm	Primarily studied in soft tissues; some potential for cancellous bone; high degree of image artifacts.	127, 128
	X-ray/Raman	Several mm to cm	High potential for chemical specificity and contrast agents; very low signal levels.	129

terahertz imaging have been tried, although their specificity to biological information beyond water content of the tissue is unclear and they are constrained to fairly superficial tissue thicknesses (μm to mm).

At the other end of the penetrance scale (mm to cm), near-infrared spectroscopy (NIRS) can be used through several centimeters of tissue, albeit with very limited spatial resolution, for deep tissue sensing of absorption and scattering features. Time-resolved NIRS, which measures photon time-of-flight allowing for selection of late-arriving photons to increase depth sensitivity, may offer improvements in resolution and quantification and has been used recently in joint imaging.⁹⁶ So, while NIRS has its limitations, it is likely a more useful tool for macroscopic tissue assessment and is largely limited to the measurement of blood, oxygen saturation, water, and lipid content of the tissue or scattering features related to the tissue matrix composition. Thus, it can be used in conjunction with ICG as a dynamic contrast-enhanced NIRS.¹³⁰ Future work might examine the value of NIRS for osteonecrosis assessment or flow. The assessment of blood flow can be done microscopically with OCT imaging. However, macroscopic bone blood flow has been one of the challenges. Recently, the flow assessment of imaging the fluorescence from ICG has been shown to provide predictive value for flow and even potentially differentiate between endosteal and periosteal flow.^{27,44,45,49}

Perhaps the most promising tools are ones in which there is a hybrid approach to imaging information, so the tool is not being evaluated as a single diagnostic, but rather is complementary to existing clinical tools or as a combination of more than one tool. The most obvious of these is the use of ICG fluorescence imaging in which it has been added onto color laparoscopy imaging, and the two can be codisplayed on a video monitor for the surgeon to use.¹³¹ This technique has high penetrance, with several commercial systems approved for use.¹³² The creation of new molecular-specific optical contrast agents is evolving rapidly with many clinical trials ongoing. These should be eventual contrast agents for fluorescence-guided surgery in trauma, even if many of the initial studies are devoted to diseases such as cancer or cardiovascular disease.

Photoacoustic imaging exists as a hybrid optical-ultrasound tool, which can in some cases provide both optical contrast as well as ultrasound images, although like ultrasound it has its strengths in soft tissue imaging and can have large artifacts from the interfaces between hard and soft tissues or voids within either tissue. Still, the photoacoustic mode of imaging vascularity and even flow can be significant and provide exquisite detail on vascular patterns and function when deployed appropriately.^{127,128,133} The clinical utility of these systems is evolving rapidly, and evaluating their use in trauma surgery should be a high priority.

Figure 1 shows an overview of the needs in trauma surgery, based upon the depth of penetrance of the imaging tool and the biological timeline of the disease. This figure provides a basic template from which to think about the tools and their value to trauma or disease diagnostics. While fine structured fractures might be seen with high-resolution tools, this is the realm of x-ray imaging or microscopy with optics such as OCT.^{91,92} Vascular function such as flow or perfusion to the tissue can be assessed microscopically or macroscopically, based upon the scope of the

Fig. 1 The matching process of the capabilities of each optical tool with the needs in each application.

problem, but most clinical evaluations would require macroscopic sensing such as with NIRS, fluorescence, or photoacoustics. Biological functions such as regrowth or necrosis have two very different sampling scales, either microscopic methods such as UV and antibiotic fluorescence or macroscopic tools such as NIRS and contrast agent fluorescence.^{27,44,45,49} Again, for most clinical issues, the sampling of macroscopic signals is likely needed, and the combination with existing clinical tools such as ultrasound or x-ray would be beneficial.

5 Conclusions

Research is needed to optimize optical imaging techniques for tissue trauma surgical applications, and the developments described in Sec. 4 are each important to test. Based on the safety profile and previous implementation success to other surgical domains, the barriers to the translation of these techniques to orthopaedic surgery are relatively low. We believe that using optical imaging to provide surgeons with real-time objective data regarding bone and tissue perfusion will lead to more effective patient-specific management of common orthopaedic conditions with lower morbidity and will result in decreased variability associated with how these conditions are managed. Information provided should go beyond the basic soft and hard tissue structures available with x-ray imaging and incorporate functional flow and perfusion or tissue metabolism features that are likely to have a higher specific correlation to the outcome of the procedure. Optical tools have the best opportunity to impact surgery because of their inherent point-of-care use, their relatively low capital costs, and their compatibility with intraprocedure measurement.

Disclosures

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References

1. I. McCarthy, "The physiology of bone blood flow: a review," *J. Bone Joint. Surg. Am.* **88**(Suppl. 3), 4–9 (2006).
2. C. Clark and J. Ogden, "Development of the menisci of the human knee joint. Morphological changes and their potential role in childhood meniscal injury," *JBJS* **65**(4), 538–547 (1983).
3. S. P. Arnoczky and R. F. Warren, "Microvasculature of the human meniscus," *Am. J. Sports Med.* **10**(2), 90–95 (1982).
4. G. Cierny, 3rd, J. T. Mader, and J. J. Penninck, "A clinical staging system for adult osteomyelitis," *Clin. Orthop. Relat. Res.* **414**, 7–24 (2003).
5. B. Parsons and E. Strauss, "Surgical management of chronic osteomyelitis," *Am. J. Surg.* **188**(1), 57–66 (2004).
6. F. F. Stecker et al., "Early dynamic ¹⁸F-FDG PET/CT to diagnose chronic osteomyelitis following lower extremity fractures. A pilot study," *Nuklearmedizin* **53**(3), 117–122 (2014).
7. M. Freesmeyer et al., "First experience with early dynamic ¹⁸F-NaF-PET/CT in patients with chronic osteomyelitis," *Ann. Nucl. Med.* **28**(4), 314–321 (2014).
8. C. Fischer et al., "Dynamic contrast-enhanced magnetic resonance imaging (DCE-MRI) for the prediction of non-union consolidation," *Injury* **48**(2), 357–363 (2017).
9. A. Christersson, S. Larsson, and J. Sorensen, "Presurgical localization of infected avascular bone segments in chronic complicated posttraumatic osteomyelitis in the lower extremity using dual-tracer PET/CT," *EJNMMI Res.* **8**(1), 65 (2018).

10. C. Fischer et al., "Dynamic contrast-enhanced sonography and dynamic contrast-enhanced magnetic resonance imaging for preoperative diagnosis of infected nonunions," *J. Ultrasound Med.* **35**(5), 933–942 (2016).
11. D. Liao et al., "Dynamic contrast-enhanced magnetic resonance imaging for differentiating osteomyelitis from acute neuropathic arthropathy in the complicated diabetic foot," *Skeletal Radiol.* **47**(10), 1337–1347 (2018).
12. G. Muller et al., "Increased perfusion in dynamic gadolinium-enhanced MRI correlates with areas of bone repair and of bone necrosis in patients with Kienbock's disease," *J. Magn. Reson. Imaging* **50**(2), 481–489 (2018).
13. O. Schoierer et al., "Dynamic contrast-enhanced magnetic resonance imaging can assess vascularity within fracture non-unions and predicts good outcome," *Eur. Radiol.* **24**(2), 449–459 (2014).
14. B. Wyrsch et al., "Operative treatment of fractures of the tibial plafond. A randomized, prospective study," *J. Bone Joint. Surg. Am.* **78**(11), 1646–1657 (1996).
15. S. M. Teeny and D. A. Wiss, "Open reduction and internal fixation of tibial plafond fractures. Variables contributing to poor results and complications," *Clin. Orthop. Relat. Res.* **292**, 108–117 (1993).
16. M. A. McFerran et al., "Complications encountered in the treatment of pilon fractures," *J. Orthop. Trauma* **6**(2), 195–200 (1992).
17. D. N. Oviada and R. K. Beals, "Fractures of the tibial plafond," *J. Bone Joint. Surg. Am.* **68**(4), 543–551 (1986).
18. M. Sirkin et al., "A staged protocol for soft tissue management in the treatment of complex pilon fractures," *J. Orthop. Trauma* **13**(2), 78–84 (1999).
19. D. L. Helfet et al., "Intraarticular "pilon" fracture of the tibia," *Clin. Orthop. Relat. Res.* **298**, 221–228 (1994).
20. V. J. Leone, R. T. Ruland, and B. P. Meinhard, "The management of the soft tissues in pilon fractures," *Clin. Orthop. Relat. Res.* **292**, 315–320 (1993).
21. M. Blauth et al., "Surgical options for the treatment of severe tibial pilon fractures: a study of three techniques," *J. Orthop. Trauma* **15**(3), 153–160 (2001).
22. J. Bear, N. Rollick, and D. Helfet, "Evolution in management of tibial pilon fractures," *Curr. Rev. Musculoskeletal Med.* **11**(4), 537–545 (2018).
23. T. O. White et al., "The results of early primary open reduction and internal fixation for treatment of OTA 43. C-type tibial pilon fractures: a cohort study," *J. Orthop. Trauma* **24**(12), 757–763 (2010).
24. X. Tang et al., "Comparison of early and delayed open reduction and internal fixation for treating closed tibial pilon fractures," *Foot Ankle Int.* **35**(7), 657–664 (2014).
25. F. A. Liporace and R. S. Yoon, "Decisions and staging leading to definitive open management of pilon fractures: where have we come from and where are we now?" *J. Orthop. Trauma* **26**(8), 488–498 (2012).
26. I. Valerio et al., "Vascularized osseous flaps and assessing their bipartate perfusion pattern via intraoperative fluorescence angiography," *J. Reconstr. Microsurg.* **31**(1), 45–53 (2015).
27. J. T. Nguyen et al., "Bone flap perfusion assessment using near-infrared fluorescence imaging," *J. Surg. Res.* **178**(2), e43–e50 (2012).
28. M. Griffiths, M. P. Chae, and W. M. Rozen, "Indocyanine green-based fluorescent angiography in breast reconstruction," *Gland Surg.* **5**(2), 133–149 (2016).
29. S. J. Mirhaidari et al., "A prospective study of immediate breast reconstruction with laser-assisted indocyanine green angiography," *Plast. Reconstr. Surg. Glob. Open* **6**(9), e1774 (2018).
30. M. J. Bosse et al., "An analysis of outcomes of reconstruction or amputation after leg-threatening injuries," *New Engl. J. Med.* **347**(24), 1924–1931 (2002).
31. A. M. Harris et al., "Complications following limb-threatening lower extremity trauma," *J. Orthop. Trauma* **23**(1), 1–6 (2009).
32. J. J. Keeling et al., "Short-term outcomes of severe open wartime tibial fractures treated with ring external fixation," *JBJS* **90**(12), 2643–2651 (2008).
33. A. Lerner, L. Fodor, and M. Soudry, "Is staged external fixation a valuable strategy for war injuries to the limbs?" *Clin. Orthop. Relat. Res.* **448**, 217–224 (2006).

34. B. D. Owens et al., "Combat wounds in operation Iraqi freedom and operation enduring freedom," *J. Trauma Acute Care Surg.* **64**(2), 295–299 (2008).
35. K. Merritt, "Factors increasing the risk of infection in patients with open fractures," *J. Trauma* **28**(6), 823–827 (1988).
36. E. P. Dellinger et al., "Risk of infection after open fracture of the arm or leg," *Arch. Surg.* **123**(11), 1320–1327 (1988).
37. N. D. Charkes, M. Brookes, and P. T. Makler Jr, "Studies of skeletal tracer kinetics: II. Evaluation of a five-compartment model of [^{18}F]fluoride kinetics in rats," *J. Nucl. Med.* **20**, 1150–1157 (1979).
38. P. M. Gross, D. D. Heistad, and M. L. Marcus, "Neurohumoral regulation of blood flow to bones and marrow," *Am. J. Physiol.-Heart Circul. Physiol.* **237**(4), H440–H448 (1979).
39. R. Wootton, J. Reeve, and N. Veall, "The clinical measurement of skeletal blood flow," *Clin. Sci. Mol. Med.* **50**(4), 261–268 (1976).
40. Y. Fujikawa et al., "The human osteoclast precursor circulates in the monocyte fraction," *Endocrinology* **137**(9), 4058–4060 (1996).
41. G. Z. Eghbali-Fatourehchi et al., "Circulating osteoblast-lineage cells in humans," *New Engl. J. Med.* **352**(19), 1959–1966 (2005).
42. I. B. Mazo and U. H. von Andrian, "Adhesion and homing of blood-borne cells in bone marrow microvessels," *J. Leukocyte Biol.* **66**(1), 25–32 (1999).
43. C. Martin et al., "Chemokines acting via CXCR2 and CXCR4 control the release of neutrophils from the bone marrow and their return following senescence," *Immunity* **19**(4), 583–593 (2003).
44. J. T. Elliott et al., "Intraoperative fluorescence perfusion assessment should be corrected by a measured subject-specific arterial input function," *J. Biomed. Opt.* **25**(6), 066002 (2020).
45. S. Jiang et al., "Endosteal and periosteal blood flow quantified with dynamic contrast-enhanced fluorescence to guide open orthopaedic surgery," *Proc. SPIE* **11222**, 112220F (2020).
46. M. T. Ovaska et al., "Predictors of poor outcomes following deep infection after internal fixation of ankle fractures," *Injury* **44**(7), 1002–1006 (2013).
47. E. Rightmire, D. Zurakowski, and M. Vrahas, "Acute infections after fracture repair," *Clin. Orthop. Relat. Res.* **466**(2), 466–472 (2008).
48. R. M. Reilly et al., "Are antibiotic nails effective in the treatment of infected tibial fractures?" *Injury* **47**(12), 2809–2815 (2016).
49. A. Zaheer et al., "In vivo near-infrared fluorescence imaging of osteoblastic activity," *Nat. Biotechnol.* **19**(12), 1148–1154 (2001).
50. C. Andersson, M. Odensten, and J. Gillquist, "Knee function after surgical or nonsurgical treatment of acute rupture of the anterior cruciate ligament: a randomized study with a long-term follow-up period," *Clin. Orthop. Relat. Res.* **264**, 255–263 (1991).
51. C. Andersson et al., "Surgical or non-surgical treatment of acute rupture of the anterior cruciate ligament. A randomized study with long-term follow-up," *J. Bone Joint Surg. Am.* **71**(7), 965–974 (1989).
52. L. Engebretsen et al., "A prospective, randomized study of three surgical techniques for treatment of acute ruptures of the anterior cruciate ligament," *Am. J. Sports Med.* **18**(6), 585–590 (1990).
53. T. Grøntvedt et al., "A prospective, randomized study of three operations for acute rupture of the anterior cruciate ligament. Five-year follow-up of one hundred and thirty-one patients," *JBJS* **78**(2), 159–169 (1996).
54. T. Grøntvedt and L. Engebretsen, "Comparison between two techniques for surgical repair of the acutely torn anterior cruciate ligament: a prospective, randomized follow-up study of 48 patients," *Scand. J. Med. Sci. Sports* **5**(6), 358–363 (1995).
55. M. Odensten et al., "Surgical or conservative treatment of the acutely torn anterior cruciate ligament; a randomized study with a short-term follow-up," in *Surgery and Arthroscopy of the Knee*, E. L. Trickey and P. Hertel, Eds., pp. 131, Springer, New York (1986).
56. R. Sandberg et al., "Operative versus non-operative treatment of recent injuries to the ligaments of the knee. A prospective randomized study," *J. Bone Joint Surg. Am.* **69**(8), 1120–1126 (1987).

57. M. S. Dhillon, K. Bali, and S. Prabhakar, "Differences among mechanoreceptors in healthy and injured anterior cruciate ligaments and their clinical importance," *Muscles, Ligaments Tendons J.* **2**(1), 38 (2012).
58. D. Barrett, "Proprioception and function after anterior cruciate reconstruction," *J. Bone Joint Surg.* **73**(5), 833–837 (1991).
59. C. Aga et al., "Tunnel widening in single-versus double-bundle anterior cruciate ligament reconstructed knees," *Knee Surgery, Sports Traumatol., Arthrosc.* **25**(4), 1316–1327 (2017).
60. T. B. Griffith et al., "Outcomes of repeat revision anterior cruciate ligament reconstruction," *Am. J. Sports Med.* **41**(6), 1296–1301 (2013).
61. M. M. Murray et al., "Bridge-enhanced anterior cruciate ligament repair: two-year results of a first-in-human study," *Orthop. J. Sports Med.* **7**(3), 2325967118824356 (2019).
62. J. K. Weaver et al., "Primary knee ligament repair—revisited," *Clin. Orthop. Relat. Res.* **199**, 185–191 (1985).
63. M. F. Sherman et al., "The long-term followup of primary anterior cruciate ligament repair: defining a rationale for augmentation," *Am. J. Sports Med.* **19**(3), 243–255 (1991).
64. J. L. Marshall, R. F. Warren, and T. L. Wickiewicz, "Primary surgical treatment of anterior cruciate ligament lesions," *Am. J. Sports Med.* **10**(2), 103–107 (1982).
65. U. Heim, B. Bachmann, and K. Infanger, "Reinsertion of the anterior cruciate ligaments or primary ligament plasty," *Helvetica Chirurgica ACTA* **48**, 703–708 (1981).
66. T. Strand et al., "Knee function following suture of fresh tear of the anterior cruciate ligament," *Acta Orthop. Scand.* **55**(2), 181–184 (1984).
67. M. Marcacci et al., "Notes on 53 cases of immediate suture of acute lesions of the anterior cruciate ligament," *Ital. J. Sports Traumatol.* **7**(2), 69–79 (1985).
68. D. E. Cooper, S. P. Arnoczky, and R. F. Warren, "Meniscal repair," *Clin. Sports Med.* **10**(3), 529–548 (1991).
69. A. F. Anderson et al., "Interobserver reliability of the International Society of Arthroscopy, Knee Surgery and Orthopaedic Sports Medicine (ISAKOS) classification of meniscal tears," *Am. J. Sports Med.* **39**(5), 926–932 (2011).
70. Y. Dong et al., "The effect of meniscal tears and resultant partial meniscectomies on the knee contact stresses: a finite element analysis," *Comput. Methods Biomech. Biomed. Eng.* **17**(13), 1452–1463 (2014).
71. M. Englund et al., "Patient-relevant outcomes fourteen years after meniscectomy: influence of type of meniscal tear and size of resection," *Rheumatology* **40**(6), 631–639 (2001).
72. I. Pengas et al., "Total meniscectomy in adolescents: a 40-year follow-up," *J. Bone Joint Surg.* **94**(12), 1649–1654 (2012).
73. C. A. Petty and J. H. Lubowitz, "Does arthroscopic partial meniscectomy result in knee osteoarthritis? A systematic review with a minimum of 8 years' follow-up," *Arthrosc.: J. Arthrosc. Relat. Surg.* **27**(3), 419–424 (2011).
74. H. Roos et al., "Knee osteoarthritis after meniscectomy: prevalence of radiographic changes after twenty-one years, compared with matched controls," *Arthritis Rheumat.* **41**(4), 687–693 (1998).
75. J. J. Tenuta and R. A. Arciero, "Arthroscopic evaluation of meniscal repairs: factors that effect healing," *Am. J. Sports Med.* **22**(6), 797–802 (1994).
76. K. L. Vanderhave et al., "Meniscus tears in the young athlete: results of arthroscopic repair," *J. Pediatric Orthop.* **31**(5), 496–500 (2011).
77. M. H. Rubman, F. R. Noyes, and S. D. Barber-Westin, "Arthroscopic repair of meniscal tears that extend into the avascular zone," *Am. J. Sports Med.* **26**(1), 87–95 (1998).
78. C. M. Mintzer, J. C. Richmond, and J. Taylor, "Meniscal repair in the young athlete," *Am. J. Sports Med.* **26**(5), 630–633 (1998).
79. F. R. Noyes et al., "Greater than 10-year results of red-white longitudinal meniscal repairs in patients 20 years of age or younger," *Am. J. Sports Med.* **39**(5), 1008–1017 (2011).
80. G. R. Barrett, "Clinical results of meniscus repair in patients 40 years and older," *Arthrosc.: J. Arthrosc. Relat. Surg.* **14**(8), 824–829 (1998).
81. S. Jiang et al., "Predicting breast tumor response to neoadjuvant chemotherapy with diffuse optical spectroscopic tomography prior to treatment," *Clin. Cancer Res.* **20**(23), 6006–6015 (2014).

82. A. Taruttis and V. Ntziachristos, "Translational optical imaging," *Am. J. Roentgenol.* **199**(2), 263–271 (2012).
83. G. D. Luker and K. E. Luker, "Optical imaging: current applications and future directions," *J. Nucl. Med.* **49**(1), 1–4 (2008).
84. O. Ristow et al., "Comparison of auto-fluorescence and tetracycline fluorescence for guided bone surgery of medication-related osteonecrosis of the jaw: a randomized controlled feasibility study," *Int. J. Oral Maxillofac Surg.* **46**(2), 157–166 (2017).
85. O. Ristow and C. Pautke, "Auto-fluorescence of the bone and its use for delineation of bone necrosis," *Int. J. Oral Maxillofac Surg.* **43**(11), 1391–1393 (2014).
86. D. Chen et al., "Laser-based gas absorption spectroscopy in decaying hip bone: water vapor as a predictor of osteonecrosis," *J. Biomed. Opt.* **24**(6), 065001 (2019).
87. A. Pifferi et al., "Optical biopsy of bone tissue: a step toward the diagnosis of bone pathologies," *J. Biomed. Opt.* **9**(3), 474–480 (2004).
88. A. J. Makowski et al., "Polarization in Raman spectroscopy helps explain bone brittleness in genetic mouse models," *J. Biomed. Opt.* **19**(11), 117008 (2014).
89. M. F. Wood et al., "Polarization birefringence measurements for characterizing the myocardium, including healthy, infarcted, and stem-cell-regenerated tissues," *J. Biomed. Opt.* **15**(4), 047009 (2010).
90. X. Wu et al., "High-speed polarization imaging of dynamic collagen fiber realignment in tendon-to-bone insertion region," *J. Biomed. Opt.* **23**(11), 116002 (2018).
91. C. Kasseck et al., "Comparison of optical coherence tomography, microcomputed tomography, and histology at a three-dimensionally imaged trabecular bone sample," *J. Biomed. Opt.* **15**(4), 046019 (2010).
92. M. Del-Valle, E. Lins, and P. Ana, "Assessment of simulated osteoporosis in alveolar bone using optical coherence tomography," *J. Biophotonics* **12**(12), e201900171 (2019).
93. T. Rashnavadi et al., "Monitoring spinal cord hemodynamics and tissue oxygenation: a review of the literature with special focus on the near-infrared spectroscopy technique," *Spinal Cord* **57**(8), 617–625 (2019).
94. R. C. Mesquita et al., "Optical monitoring and detection of spinal cord ischemia," *PLoS One* **8**(12), e83370 (2013).
95. B. S. Klosterhoff et al., "Implantable sensors for regenerative medicine," *J. Biomech. Eng.* **139**(2), 0210091 (2017).
96. S. Ioussoufovitch et al., "Quantification of joint blood flow by dynamic contrast-enhanced near-infrared spectroscopy: application to monitoring disease activity in a rat model of rheumatoid arthritis," *J. Biomed. Opt.* **25**(1), 015003 (2020).
97. A. Kashigar, J. S. Wunder, and P. C. Ferguson, "Computer-assisted orthopaedic surgery: introduction and applications in orthopaedic oncology," *Univ. Toronto Med. J.* **92**(3), 78–83 (2015).
98. S. C. Lee et al., "Multi-modal imaging, model-based tracking, and mixed reality visualisation for orthopaedic surgery," *Healthcare Technol. Lett.* **4**(5), 168–173 (2017).
99. M. Bennet et al., "Simultaneous Raman microspectroscopy and fluorescence imaging of bone mineralization in living zebrafish larvae," *Biophys. J.* **106**(4), L17–L19 (2014).
100. T. Buchwald et al., "Identifying compositional and structural changes in spongy and subchondral bone from the hip joints of patients with osteoarthritis using Raman spectroscopy," *J. Biomed. Opt.* **17**(1), 017007 (2012).
101. K. A. Dooley et al., "Stress mapping of undamaged, strained, and failed regions of bone using Raman spectroscopy," *J. Biomed. Opt.* **14**(4), 044018 (2009).
102. G. Falgayrac et al., "New method for Raman investigation of the orientation of collagen fibrils and crystallites in the Haversian system of bone," *Appl. Spectrosc.* **64**(7), 775–780 (2010).
103. B. Gong et al., "Raman spectroscopy demonstrates prolonged alteration of bone chemical composition following extremity localized irradiation," *Bone* **57**(1), 252–258 (2013).
104. M. V. Schulmerich et al., "Noninvasive Raman tomographic imaging of canine bone tissue," *J. Biomed. Opt.* **13**(2), 020506 (2008).

105. M. V. Schulmerich et al., "Transcutaneous fiber optic Raman spectroscopy of bone using annular illumination and a circular array of collection fibers," *J. Biomed. Opt.* **11**(6), 060502 (2006).
106. J. A. Timlin et al., "Spatial distribution of phosphate species in mature and newly generated Mammalian bone by hyperspectral Raman imaging," *J. Biomed. Opt.* **4**(1), 28–34 (1999).
107. A. Boskey and R. Mendelsohn, "Infrared analysis of bone in health and disease," *J. Biomed. Opt.* **10**(3), 031102 (2005).
108. M. D. Morris, A. Berger, and A. Mahadevan-Jansen, "Infrared and Raman spectroscopy," *J. Biomed. Opt.* **10**(3), 031101 (2005).
109. D. Vincke et al., "Analysis of collagen preservation in bones recovered in archaeological contexts using NIR hyperspectral imaging," *Talanta* **125**, 181–188 (2014).
110. M. Plazanet et al., "Time-domain THz spectroscopy of the characteristics of hydroxyapatite provides a signature of heating in bone tissue," *PLoS One* **13**(8), e0201745 (2018).
111. M. R. Stringer et al., "The analysis of human cortical bone by terahertz time-domain spectroscopy," *Phys. Med. Biol.* **50**(14), 3211–3219 (2005).
112. R. A. Milch, D. P. Rall, and J. E. Tobie, "Fluorescence of tetracycline antibiotics in bone," *J. Bone Joint Surg. Am.* **40-A**(4), 897–910 (1958).
113. R. F. Canalis and J. Lechago, "Tetracycline bone labeling: an improved technique using incident fluorescence," *Ann. Otol. Rhinol. Laryngol.* **91**(2 Pt 1), 160–162 (1982).
114. C. Pautke et al., "Tetracycline bone fluorescence: a valuable marker for osteonecrosis characterization and therapy," *J. Oral Maxillofac Surg.* **68**(1), 125–129 (2010).
115. S. Otto et al., "Successful surgical management of osteonecrosis of the jaw due to RANK-ligand inhibitor treatment using fluorescence guided bone resection," *J. Craniomaxillofac Surg.* **41**(7), 694–698 (2013).
116. F. Wehrhan et al., "Fluorescence-guided bone resection: a histological analysis in medication-related osteonecrosis of the jaw," *J. Craniomaxillofac Surg.* **47**(10), 1600–1607 (2019).
117. A. Akiva et al., "Intercellular pathways from the vasculature to the forming bone in the zebrafish larval caudal fin: possible role in bone formation," *J. Struct. Biol.* **206**(2), 139–148 (2019).
118. C. Price et al., "An in-situ fluorescence-based optical extensometry system for imaging mechanically loaded bone," *J. Orthop. Res.* **28**(6), 805–811 (2010).
119. L. Wang et al., "In situ measurement of solute transport in the bone lacunar-canalicular system," *Proc. Natl. Acad. Sci. U. S. A.* **102**(33), 11911–11916 (2005).
120. X. Zhou, J. E. Novotny, and L. Wang, "Modeling fluorescence recovery after photobleaching in loaded bone: potential applications in measuring fluid and solute transport in the osteocytic lacunar-canalicular system," *Ann. Biomed. Eng.* **36**(12), 1961–1977 (2008).
121. J. T. Elliott et al., "Bone-specific kinetic model to quantify periosteal and endosteal blood flow using indocyanine green in fluorescence guided orthopedic surgery," *J. Biophotonics* **12**(8), e201800427 (2019).
122. G. Tringali et al., "Two-staged frontal bone defect reconstruction: perioperative assessment of scalp vascularization using near-infrared indocyanine green video angiography (vision-sense iridium)," *World Neurosurg.* **126**, 502–507 (2019).
123. G. Campi et al., "Heterogeneous and self-organizing mineralization of bone matrix promoted by hydroxyapatite nanoparticles," *Nanoscale* **9**(44), 17274–17283 (2017).
124. Q. Chen et al., "Bone targeted delivery of SDF-1 via alendronate functionalized nanoparticles in guiding stem cell migration," *ACS Appl. Mater. Interfaces* **10**(28), 23700–23710 (2018).
125. M. Konar, J. K. Sahoo, and H. Sahoo, "Impact of bone extracellular matrix mineral based nanoparticles on structure and stability of purified bone morphogenetic protein 2 (BMP-2)," *J. Photochem. Photobiol. B* **198**, 111563 (2019).
126. V. Vratsha et al., "Mechanisms of cellular internalization of quantum Dot(R) conjugated bone formation mimetic peptide CK2.3," *Nanomaterials (Basel)* **8**(7), (2018).
127. I. Steinberg et al., "Quantitative study of optical and mechanical bone status using multi-spectral photoacoustics," *J. Biophotonics* **9**(9), 924–933 (2016).

128. L. Yang et al., "Photoacoustic and ultrasound imaging of cancellous bone tissue," *J. Biomed. Opt.* **20**(7), 076016 (2015).
129. J. L. Demers et al., "Next-generation Raman tomography instrument for non-invasive in vivo bone imaging," *Biomed. Opt. Express* **6**(3), 793–806 (2015).
130. J. T. Elliott et al., "Quantifying cerebral blood flow in an adult pig ischemia model by a depth-resolved dynamic contrast-enhanced optical method," *Neuroimage* **94**, 303–311 (2014).
131. J. T. Elliott et al., "Review of fluorescence guided surgery visualization and overlay techniques," *Biomed. Opt. Express* **6**(10), 3765–3782 (2015).
132. D. S. Av et al., "Review of fluorescence guided surgery systems: identification of key performance capabilities beyond indocyanine green imaging," *J. Biomed. Opt.* **21**(8), 080901 (2016).
133. P. Wray et al., "Photoacoustic computed tomography of human extremities," *J. Biomed. Opt.* **24**(2), 026003 (2019).

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